

Photo credit: Theresa Walunas, http://www.keyboardbiologist.net/knitblog/

Based on this figure, what is expected in an illustration?

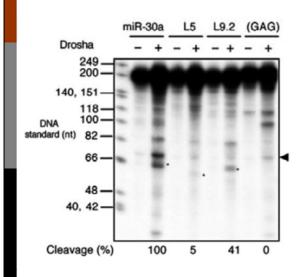
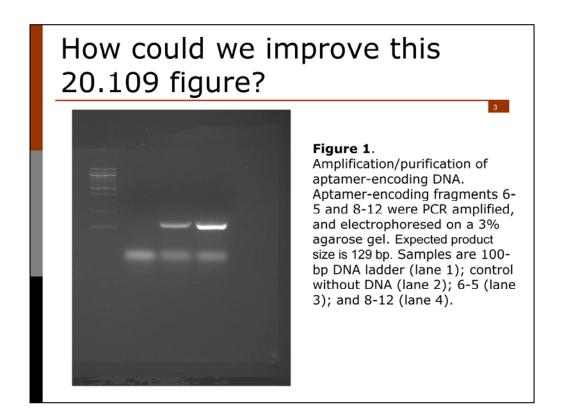


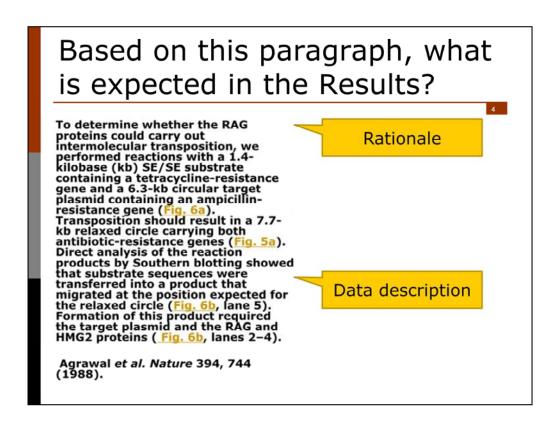
Figure 5. In vitro Drosha cleavage of pri-miR-30a transcripts. FLAG immuno-precipitates from mock-transfected 293T cells (-) or cells transfected with pCK-Drosha-FLAG(+) were incubated with a ³²P-labeled ~202 nt RNA probe encoding the indicated miR-30a variants. RNA cleavage products were recovered and resolved on a denaturing 10% polyacrylamide gel. The pre-miRNA band is indicated by an asterisk, and the background band running slightly above by an arrowhead.

Zeng et al. (2005) EMBO J. 24: 138.



This slide shows the revision. Below is the original.

Figure 1 shows the DNA ladder, control, 6-5, and 8-12.



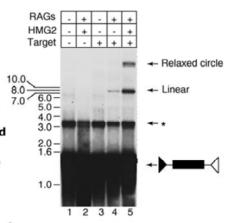
This paragraph lacks a conclusion sentence, i.e. the answer to the question posed (rationale). Providing a conclusion sentence ties together your observations, and helps smooth the transition to the next experiment.

Describe illustrations so that the reader not look at them.

To determine whether the RAG proteins could carry out intermolecular transposition, we performed reactions with a 1.4-kilobase (kb) SE/SE substrate containing a tetracycline-resistance gene and a 6.3-kb circular target plasmid containing an ampicillinresistance gene (Fig. 6a).

Transposition should result in a 7.7-kb relaxed circle carrying both antibiotic-resistance genes (Fig. 5a).

Direct analysis of the reaction products by Southern blotting showed that substrate sequences were transferred into a product that migrated at the position expected for the relaxed circle (Fig. 6b, lane 5). Formation of this product required the target plasmid and the RAG and HMG2 proteins (Fig. 6b, lanes 2-4).



Agrawal *et al. Nature* 394, 744 (1988).

Now, it's your turn.

Results

Preparation of DNA. We performed PCR to amplify the aptamerencoding fragments 6-5 and 8-12. To determine whether the DNA was amplified, we performed electrophoresis (Fig.1). The 6-5 and 8-12 samples generated bands between 100-200 bp, which is consistent with our expected band size of 129 bp. The reaction depended on the presence of template. Also present in all lanes were smaller bands, most likely primers. Taken together, the gel shows that DNA was amplified.

This slide shows the revision. Below is the original.

Results

We performed PCR to amplify the aptamer encoding fragments 6-5 and 8-12. The figure above shows that the DNA was amplified.