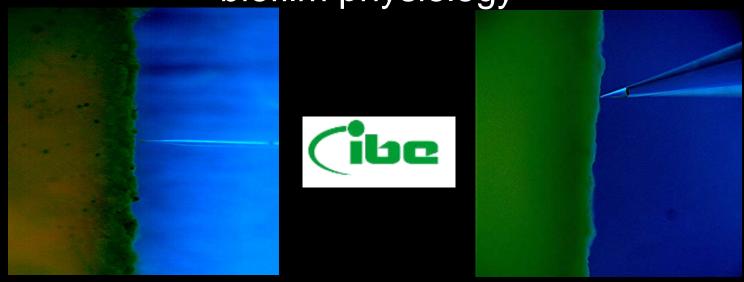


Self-referencing sensors for characterizing microbial biofilm physiology



Eric S. McLamore^{1,5}, D.M. Porterfield^{2,3,4,5}, and M.K. Banks¹

¹School of Civil Engineering

²Department of Agricultural and Biological Engineering

³Department of Horticulture & Landscape Architecture

⁴Weldon School of Biomedical Engineering

⁵Bindley Bioscience Center, Discovery Park: Physiological Sensing Facility





Biofilm Physiology

- Function
 - Ion/nutrient flux
 - Oxygen flux
- Structure
 - Shared analyses
 - Atomic force microscopy
- Why is studying biofilm physiology important?
 - Wastewater treatment
 - Toxicity studies

Biochemical (functional)

Biophysical (physiological)

Morphological (structural)





Techniques for characterizing biofilm physiology

- Respirometry
- Invasive concentration (activity) sensors
 - Oxygen, ammonium, nitrate, pH
- Classic biochemical techniques
 - In vitro detectors and assays

Non-invasive techniques with greater sensitivity and temporal resolution are required





Self-referencing sensors

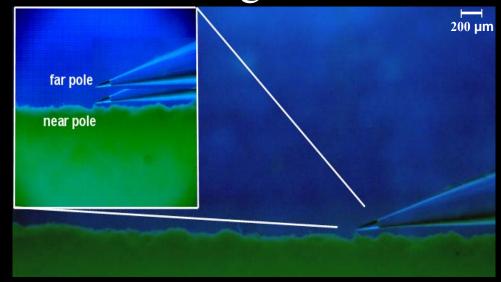
- Real time biophysical flux
 - Fixed excursion distance
 - Near pole
 - Far pole

dC = difference in concentration between near pole and far pole

$$J = -D\frac{dC}{dr}$$

dr = excursion distance

- Corrects for mechanical motion through liquid solution
 - Reference measurement
 - ≈ 0.4 cm from surface



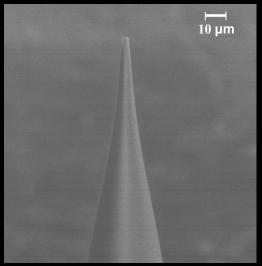


N. europaea biofilm immobilized on silicon membrane



Benefits of self-referencing technique

- Very low detection limit (fmol/cm²-sec)
- Non-invasive (non-destructive)
- Real time temporal resolution
- Direct quantification of analyte flux
 - Dynamic measurement



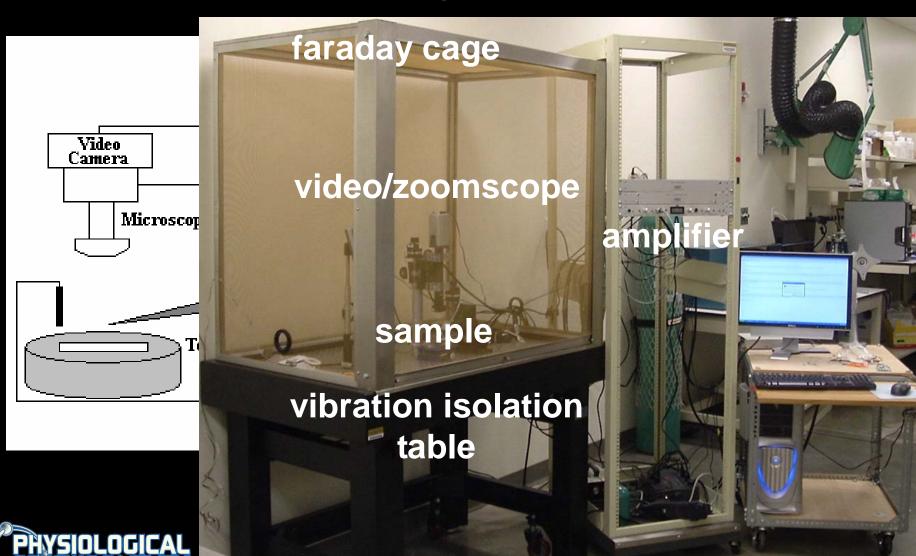
SEM of electrode tip

- Multiple sensor approach for measuring flux
 - cumulative signal error
- One sensor, one source of error





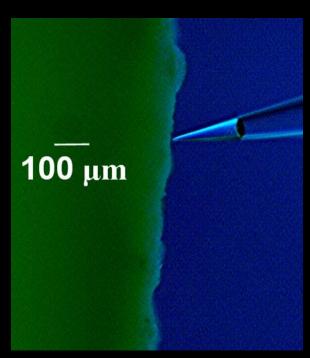
Self-referencing sensor hardware



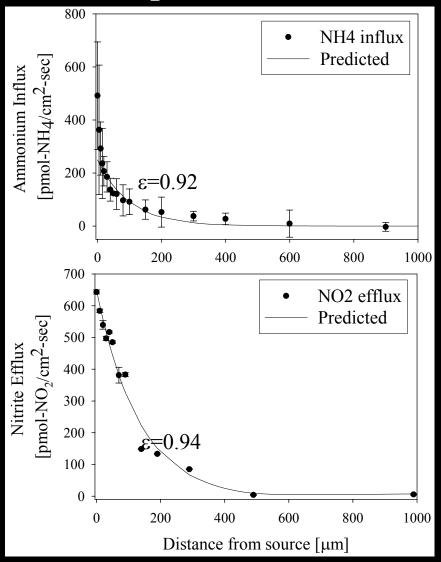
PURDUE

Sensor calibration (dynamic)

• Step-back experiments on *N. europaea* biofilm



N. europaea biofilm immobilized on silicon membrane







Classic physiological study

- Ammonia oxidizing bacteria
 - Nitrosomonas europaea
 - Ammonia monooxygenase (AMO)
 - membrane bound protein
 - copper at active site
 - Hydroxylamine oxidoreductase (HAO)
 - Produces electrons required by AMO

$$pK_a = 9.2$$

 $NH_4 \rightarrow NH_3 + H^+$

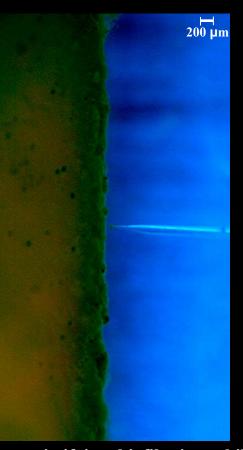
$$NH_3 + O_2 + 2H^+ \xrightarrow{AMO} NH_2OH + H_2O \xrightarrow{HAO} NO_2^- + 5H^+ + 4e^-$$





AMO inhibition

- Alternative substrates
 - AMO can oxidize over 40 substrates
- Mechanism-based (e.g., acetylene)
- Reversible inhibition
 - Visible light
 - Copper chelating agents
 - Thiourea, carbon disulfide



Mixed culture nitrifying biofilm immobilized on silicon membrane





Cell harvesting and immobilization

Microbes immobilized on oxygen-permeable

N. europaea biofilm grown in MABR

silicon membranes

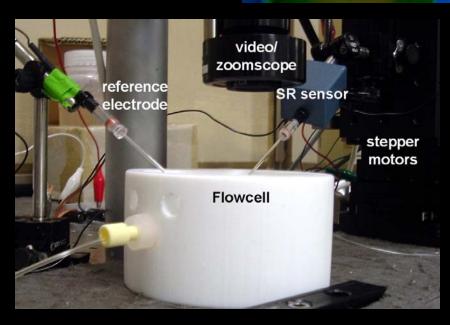
- Upflow membrane-aerated bioreactor
 - Lumen-side air flow

Membranes transferred to flow cell for

measurement

MABR limit the formation of anoxic zones









Experimental design

position 1

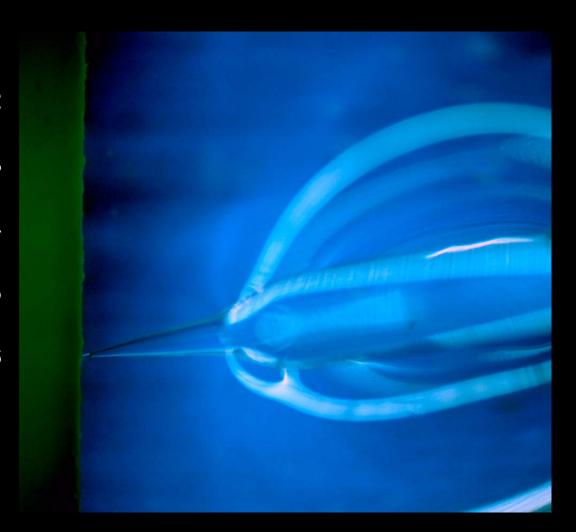
position 2

position 3

position 4

position 5

position 6



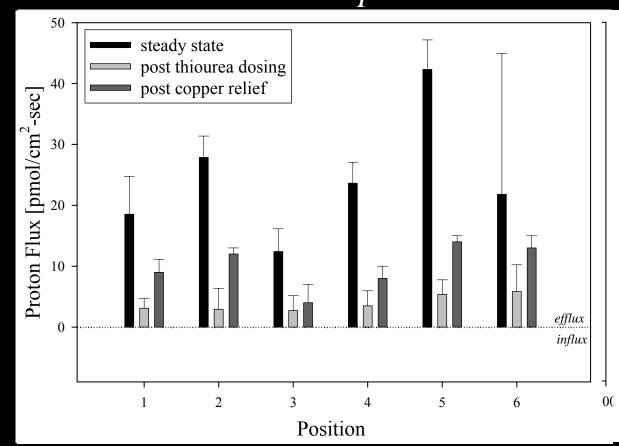




pH at biofilm surface was 6.71 ± 0.10 c

AMO inhibition study: Proton flux

Nitrosomonas europaea biofilm



Real time data collected at position 1



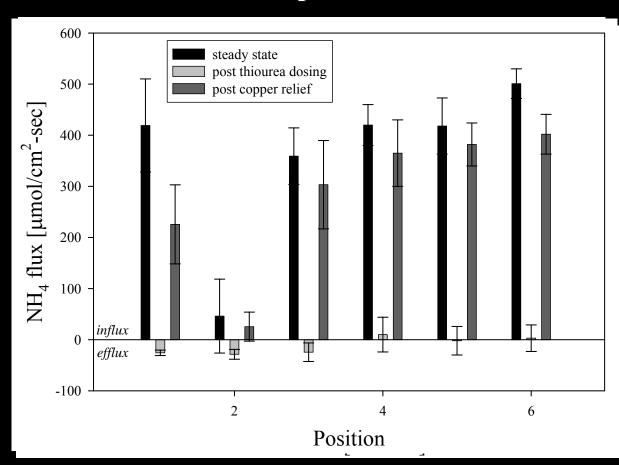


AMO inhibition study: Ammonium flux

Nitrosomonas europaea biofilm

Real time data collected at position 6

100 µM CuCl₂ sufficient for restoring steady state NH₄ influx







Conclusions

- Successfully demonstrated reversible inhibition of AMO by copper chelation in a biofilm
 - Non-invasive
 - Real time response
- More copper required to restore steady state oxygen and proton flux levels than required to restore NH₄ influx
 - Oxygen and copper are required for many cellular processes other than AMO activity
- Use of self-referencing sensors will allow noninvasive investigation of real time biofilm stress response to chemical toxin exposure





Ongoing studies

- Chemical toxicity in pure and mixed culture biofilms
 - Uncoupling of aerobic respiration
 - Glutathione gated-potassium efflux
 - Associated with deflocculation
 - Additionally measuring efflux of cross-linking ions (Ca²⁺, Mg²⁺)
 - Aerobic biodegradation of respiratory-inhibiting compounds
 - Heavy metal exposure





Acknowledgements

- M. K. Banks

Advisors

- D.M. Porterfield
- Purdue School of Civil Engineering
- Discovery Park
- Al Shipley (Applicable Electronics, Inc.)
- Jon Shaff (Cornell, USDA)
- Aeraj ul Haque, Rameez Chatni (Electric. and Comp. Engr.)

Also currently investigating effect of laminar bulk flow on sensor behavior





AMO inhibition study: Oxygen flux

Real time data collected at position 1

Nitrosomonas europaea biofilm

