

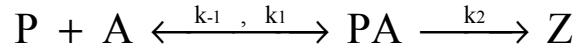
# Modelling Our System

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# Slide2: General Derivation

## Assumptions:

- Concentration of promoters is constant
- Promoter  $P$  and activator  $A$  are in equilibrium with their complex  $PA$
- The reaction forming the protein  $Z$  is irreversible
- Note: Contrarily to Michaelis-Menten, the substrate is not used up & the protein  $Z$  rebinds the promoter



$$\frac{d[PA]}{dt} = k_1[P][A] - k_{-1}[PA] = 0 \quad (\text{steady state reached quickly if } k_1 \succ k_{-1})$$

$$(1) \quad [PA] = \frac{k_1[P][A]}{k_{-1}} = [P][A]K_D \quad \text{where } K_D \equiv \frac{k_1}{k_{-1}}$$

$$\text{Note: in Michaelis-Menten } K_m = \frac{k_{-1} + k_2}{k_1} \approx \frac{k_{-1}}{k_1} = \frac{1}{K_D} \quad \text{if } k_2 \ll k_{-1}$$

As the total concentration of promoters is constant:

$$[P_0] = [P] + [PA] \quad \therefore [P] = [P_0] - [PA]$$

Substituting into (1):

$$[PA] = \frac{[P][A]}{K_D} = \frac{([P_0] - [PA])[A]}{K_D} \quad \therefore [PA] = \frac{[P_0]}{1 + \frac{K_D}{[A]}} = \frac{[A][P_0]}{[A] + K_D}$$

The rate of protein synthesis is described by:

$$\frac{d[Z]}{dt} = k_2[PA] = \frac{k_2[P_0][A]}{[A] + K_D} = \frac{V_{\max}[A]}{[A] + K_D} \quad \text{where } V_{\max} \equiv k_2[P_0]$$

# Modelling T9002

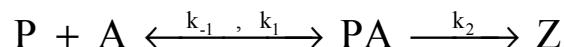


$$\frac{d[A]}{dt} = k_{\alpha}[\text{AHL}][\text{LuxR}] - k_{-\alpha}[\text{A}] = 0 \quad (\text{steady-state})$$

$\text{LuxR}$  is constitutively produced and reaches steady state before  $\text{AHL}$  is added.

$[\text{LuxR}]$  can be approximated as a constant:  $[\text{LuxR}] \approx \lambda$

$$\therefore \frac{[\text{AHL}][\text{LuxR}]}{[\text{A}]} = \frac{[\text{AHL}]}{[\text{A}]} = \frac{k_{-\alpha}}{\lambda k_{\alpha}} = \frac{1}{\lambda K_{D\alpha}} \quad \therefore (2) \quad [\text{A}] = \lambda K_{D\alpha}[\text{AHL}][\text{LuxR}]$$



The rate of protein synthesis is described by:

$$\frac{d[\text{Z}]}{dt} = k_2[\text{PA}] = \frac{k_2[\text{P}_0][\text{A}]}{[\text{A}] + K_D} = \frac{V_{\max}[\text{A}]}{[\text{A}] + K_D} \quad \text{where } V_{\max} \equiv k_2[\text{P}_0]$$

(see Slide No. 2 for derivation)

The total change in protein concentration includes protein degradation:

$$\frac{d[\text{Z}]}{dt} = \frac{V_{\max}[\text{A}]}{[\text{A}] + K_D} - \delta_2[\text{Z}]$$

$$\text{Substituting Equation (2): } \frac{d[\text{Z}]}{dt} = \frac{V_{\max} \lambda K_{D\alpha}[\text{AHL}]}{\lambda K_{D\alpha}[\text{AHL}] + K_D} - \delta_2[\text{Z}]$$

Rearranging and substituting for  $\text{Z}$ :

$$(3) \quad \frac{d[\text{GFP}]}{dt} = \frac{V_{\max}[\text{AHL}]}{[\text{AHL}] + \frac{K_D}{\lambda K_{D\alpha}}} - \delta_{\text{GFP}}[\text{GFP}]$$

## Key:

$P$ : Promoter  $p\text{LuxR}$

$A$ :  $\text{AHL}/\text{LuxR}$  complex

$PA$ :  $p\text{LuxR}/\text{AHL}/\text{LuxR}$  complex

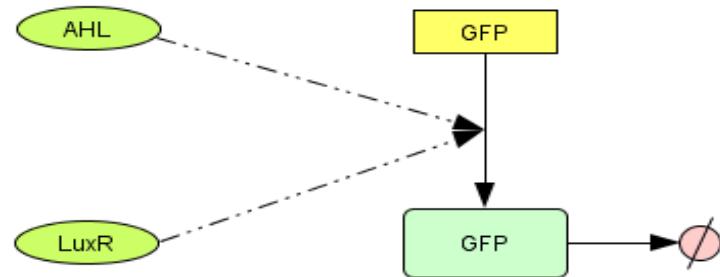
$Z$ : GFP

## Note:

$\text{AHL} + \text{LuxR} \leftrightarrow \text{AHL}/\text{LuxR}$

$\text{LuxR}$  is present in excess of  $\text{AHL}$ .

The protein  $Z$  ( $\text{AHL}$ ) associates with  $\text{LuxR}$  to form  $A$ . Thus,  $Z$  indirectly becomes the activator  $A$ .



**Fig.1a:** Diagram for model in cell designer

**Equation:**

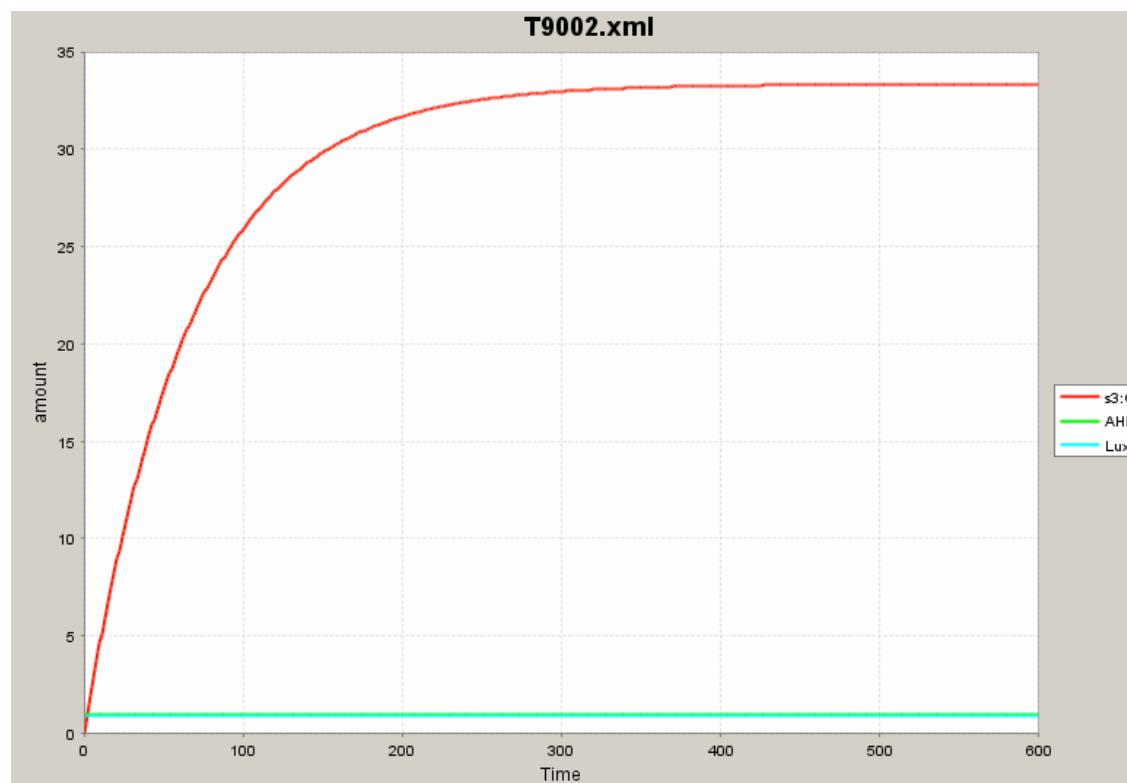
$$\frac{d[\text{GFP}]}{dt} = \frac{V_{\max} [\text{AHL}]}{[\text{AHL}] + \frac{K_D}{\lambda K_{D\alpha}}} - \delta_{\text{GFP}} [\text{GFP}]$$

**Known or measurable parameters:**

- [GFP], [AHL], GFP degradation

**Parameters to extract from model:**

- $V_{\max}$ ,  $\frac{K_D}{\lambda K_{D\alpha}}$



**Fig.1b:** Output from the model in Fig.1a

Input Values used to generate above graph:

$$\delta_{\text{GFP}} = 0.015 \text{ min}^{-1}, \frac{K_D}{\lambda K_{D\alpha}} = 1.0$$

$$V_{\max} = 1.0, [\text{AHL}] = 1.0, [\text{LuxR}] = 0.93$$

# Modelling J37015



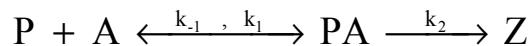
$$\frac{d[A]}{dt} = k_{\alpha}[\text{AHL}][\text{LuxR}] - k_{-\alpha}[A]$$

LuxR is constitutively produced and reaches steady state before AHL is added.

$[\text{LuxR}]$  can be approximated as a constant:  $[\text{LuxR}] \approx \lambda$

$$\frac{d[A]}{dt} = k_{\alpha}\lambda[\text{AHL}] - k_{-\alpha}[A] = 0 \quad (\text{steady-state})$$

$$\therefore \frac{[\text{AHL}]}{[A]} = \frac{k_{-\alpha}}{\lambda k_{\alpha}} = \frac{1}{\lambda K_{D\alpha}} \quad \therefore (4) \quad [A] = \lambda K_{D\alpha}[\text{AHL}]$$



The rate of protein synthesis is described by:

$$\frac{d[Z]}{dt} = k_2[\text{PA}] = \frac{k_2[P_0][A]}{[A] + K_D} = \frac{V_{\max}[A]}{[A] + K_D} \quad \text{where } V_{\max} \equiv k_2[P_0]$$

(see Slide No. 2 for derivation)

The total change in protein concentration includes protein degradation:

$$\frac{d[Z]}{dt} = \frac{V_{\max}[A]}{[A] + K_D} - \delta_1[Z]$$

$$\text{Substituting Equation (4): } \frac{d[Z]}{dt} = \frac{V_{\max}\lambda K_{D\alpha}[\text{AHL}]}{\lambda K_{D\alpha}[\text{AHL}] + K_D} - \delta_1[Z]$$

Since AHL/LuxR complex is in equilibrium with AHL, we can approximate:

$$(5) \quad \frac{d[\text{AHL}]}{dt} = \frac{V_{\max}[\text{AHL}]}{[\text{AHL}] + \cancel{K_D} / \cancel{\lambda K_{D\alpha}}} - \delta_{\text{AHL}}[\text{AHL}]$$

## Key:

P: Promoter  $p\text{LuxR}$

A: AHL/LuxR complex

PA:  $p\text{LuxR}/\text{AHL}/\text{LuxR}$  complex

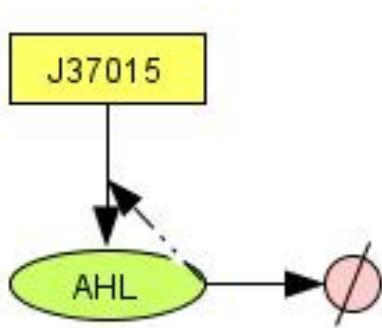
Z: AHL

## Note:



*LuxR is present in excess of AHL.*

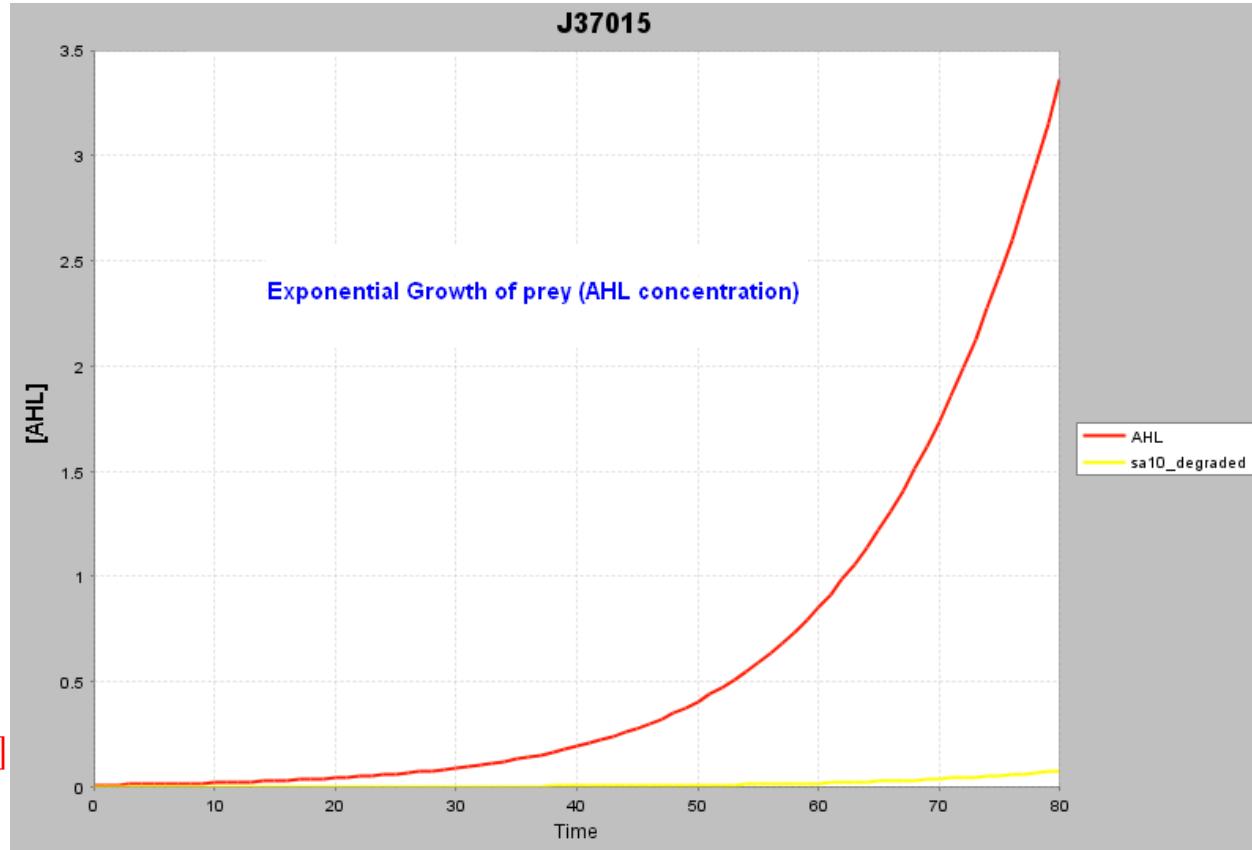
*The protein Z (AHL) associates with LuxR to form A. Thus, Z indirectly becomes the activator A.*



**Fig.2a:** Diagram for model  
in cell designer

**Equation:**

$$\frac{d[\text{AHL}]}{dt} = \frac{V_{\max}[\text{AHL}]}{[\text{AHL}] + \frac{K_D}{\lambda K_{D\alpha}}} - \delta_{\text{AHL}}[\text{AHL}]$$



**Fig.2b:** Output from the model in Fig.2a

**Known or measurable parameters:**

- [AHL], AHL degradation

**Parameters to extract from model:**

- $V_{\max}$ ,  $\frac{K_D}{\lambda K_{D\alpha}}$

**Values used for graph:**

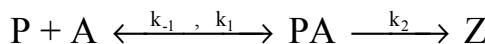
$$V_{\max} = 1.245, \frac{K_D}{\lambda K_{D\alpha}} = 15, \lambda = 1, \delta_1 = 0.0016 \text{ s}^{-1}$$

# Modelling J37016

AHL + LuxR  $\xrightleftharpoons[k_{-a}]{k_a}$  A (Assuming all stoichiometric numbers as shown)

$$\frac{d[A]}{dt} = k_a[AHL][LuxR] - k_{-a}[A] = 0 \text{ (because of steady state)}$$

$$\therefore \frac{[AHL][LuxR]}{[A]} = \frac{k_{-a}}{k_a} = \frac{1}{K_{D\alpha}} \quad \therefore (6) \quad [A] = K_{D\alpha}[AHL][LuxR]$$



The rate of protein synthesis is described by:

$$\frac{d[Z]}{dt} = k_2[PA] = \frac{k_2[P_0][A]}{[A] + K_D} = \frac{V_{\max}[A]}{[A] + K_D} \quad \text{where } V_{\max} \equiv k_2[P_0]$$

(see Slide No. 2 for derivation)

The total change in protein concentration includes protein degradation:

$$\frac{d[Z]}{dt} = \frac{V_{\max}[A]}{[A] + K_D} - \delta_2[Z]$$

$$\text{Substituting Equation (6): } \frac{d[Z]}{dt} = \frac{V_{\max}K_{D\alpha}[AHL][LuxR]}{K_{D\alpha}[AHL][LuxR] + K_D} - \delta_2[Z]$$

There are two different products being transcribed: LuxR and GFP.

Considering both products after another, keeping in mind they are measured at steady state:

$$(7) \quad \frac{d[LuxR]}{dt} = \frac{V_{\max}[AHL][LuxR]}{[AHL][LuxR] + \cancel{K_D/K_{D\alpha}}} - \delta_{LuxR}[LuxR] = 0$$

$$(8) \quad \frac{d[GFP]}{dt} = \frac{V_{\max}[AHL][LuxR]}{[AHL][LuxR] + \cancel{K_D/K_{D\alpha}}} - \delta_{GFP}[GFP] = 0$$

$$\therefore (7) = (8) \Rightarrow \delta_{LuxR}[LuxR] = \delta_{GFP}[GFP] \Rightarrow [LuxR] = \frac{\delta_{GFP}}{\delta_{LuxR}} [GFP]$$

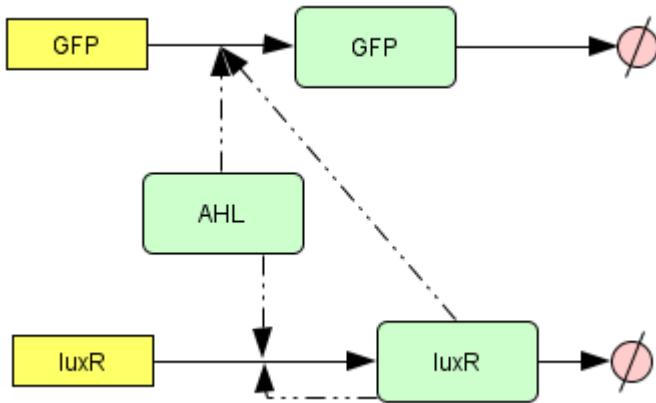
## Key:

P: Promoter *pLuxR*

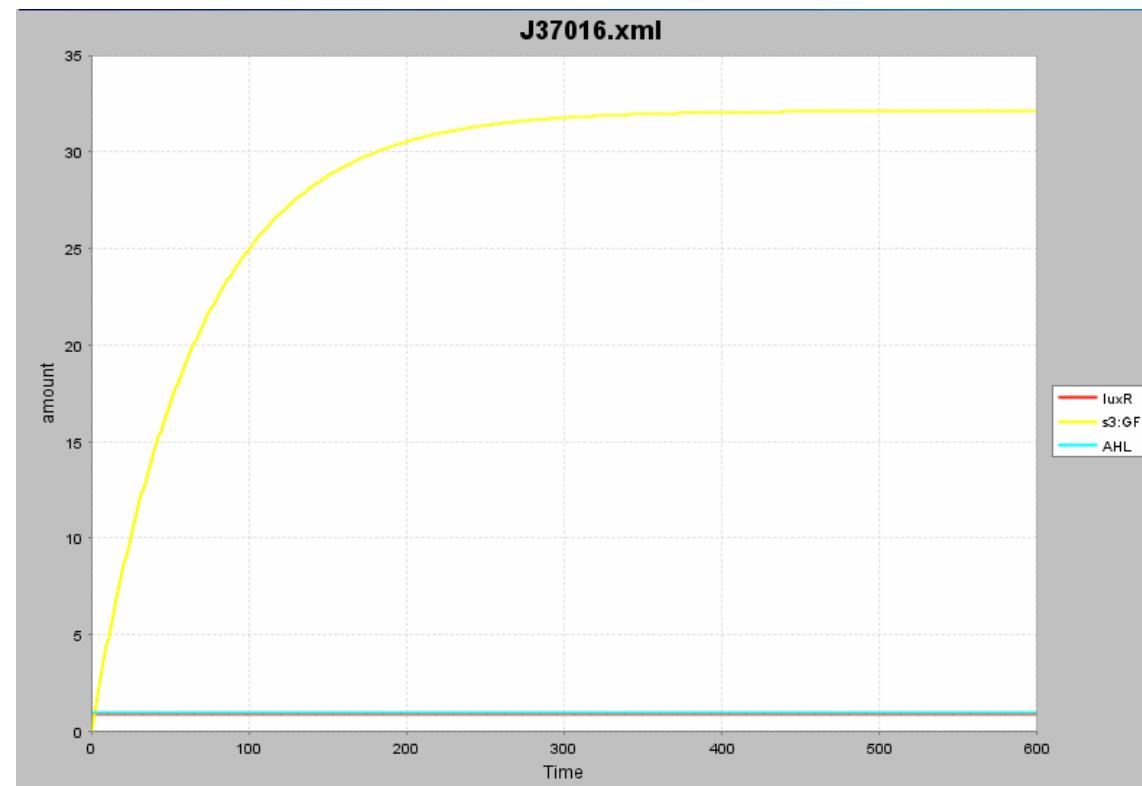
A: AHL/LuxR complex

PA: *pLuxR/AHL/LuxR complex*

Z: GFP and LuxR



**Fig.3a:** Diagram for model in cell designer



**Equation:**

$$\frac{d[GFP]}{dt} = \frac{V_{\max} [AHL][LuxR]}{[AHL][LuxR] + \frac{K_D}{K_{D\alpha}}} - \delta_{GFP}[GFP]$$

**Known or measurable parameters:**

- [AHL], GFP degradation

**Parameters to extract from model:**

- $V_{\max}$ ,  $\frac{K_D}{K_{D\alpha}}$

**Fig.3b:** Output from the model in Fig.3a

Input values used to generate above graph:

$$\delta_{GFP} = 0.015 \text{ min}^{-1}, \quad \frac{K_D}{K_{D\alpha}} = 1.0$$

$$V_{\max} = 1.0, \quad [AHL] = 1.0, \quad [LuxR] = 0.93$$

# Modelling J37022 (AHL)

## Key:

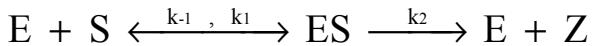
*E: Enzyme AHL-lactonase*

*S: AHL*

*ES: aiiA/AHL complex*

*Z: Acyl-HS*

True Michaelis-Menten :



$$\frac{d[ES]}{dt} = k_1[E][S] - k_{-1}[ES] - k_2[ES] = 0$$

$$(9) \quad [ES] = \frac{k_1[E][S]}{k_{-1} + k_2} = \frac{[E][S]}{K_m} \quad \text{where } K_m \equiv \frac{k_{-1} + k_2}{k_1}$$

As the total concentration of enzyme is constant:

$$[E_0] = [E] + [ES] \quad \therefore [E] = [E_0] - [ES]$$

Substituting into (1):

$$[ES] = \frac{[E][S]}{K_m} = \frac{([E_0] - [ES])[S]}{K_m} \quad \therefore [ES] = \frac{[S][E_0]}{K_m + [S]}$$

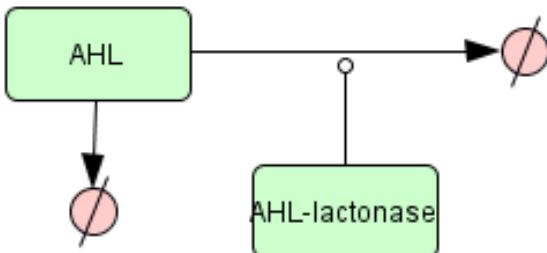
The rate of degradation of substrate (activity of enzyme)

is described by:

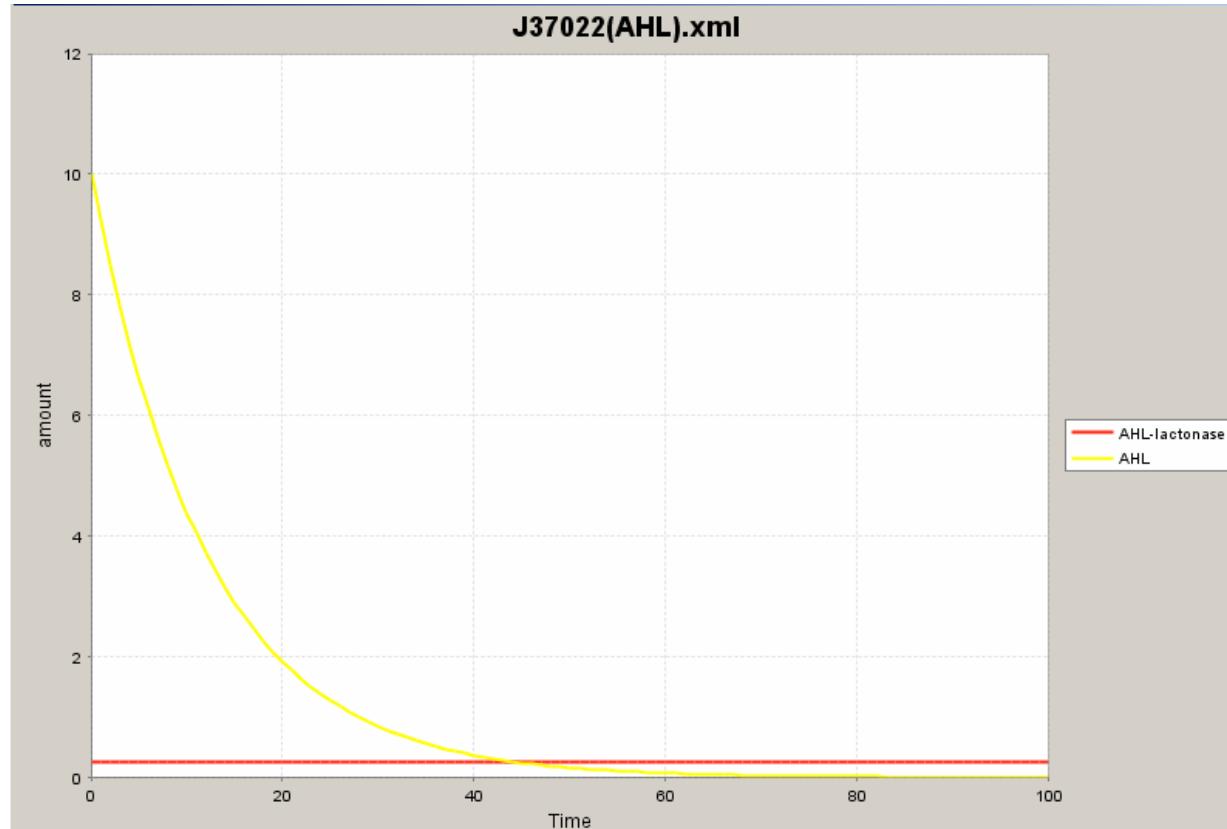
$$-\frac{d[S]}{dt} = k_2[ES] = \frac{k_2[E_0][S]}{K_m + [S]} = \frac{V_{max}[S]}{K_m + [S]} \quad \text{where } V_{max} \equiv k_2[E_0]$$

The total rate of degradation of AHL (activity of aiiA) is described by:

$$(10) \quad \frac{d[AHL]}{dt} = -\frac{V_{max}[AHL]}{K_m + [AHL]} - \delta_{AHL}[AHL] = -\frac{k_2[E_0][AHL]}{K_m + [AHL]} - \delta_{AHL}[AHL]$$



**Fig.4a:** Diagram for model in cell designer



**Fig.4b:** Output from the model in Fig.4a

**Equation:**

$$\frac{d[AHL]}{dt} = -\frac{k_2[E_0][AHL]}{K_m + [AHL]} - \delta_{AHL}[AHL]$$

**Known or measurable parameters:**

- IPTG, aiiA degradation

**Parameters to extract from model:**

- $V_{max}$ ,  $K_D$

Input values used to generate above graph:

$$\delta_{AHL} = 0.00048, \quad K_m = 331.95$$

$$V_{max} = 1.0, \quad [AHL] = 1.0$$

$$k_2 = 1000.0, \quad E_0 = 1.0$$

# Modelling J37022 (aiiA)

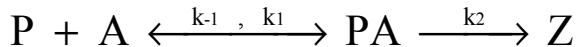
## Key:

*P*: Promoter *LacI*

*A*: *IPTG*

*PA*: *LacI/IPTG complex*

*Z*: *aiiA*



The rate of protein synthesis is described by:

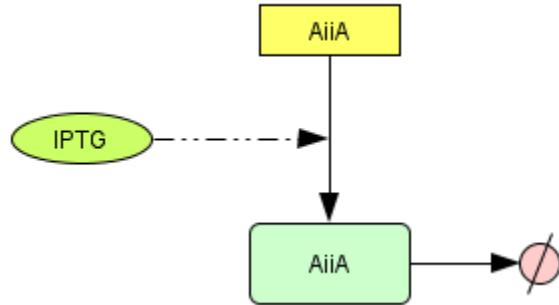
$$\frac{d[Z]}{dt} = k_2[PA] = \frac{k_2[P_0][A]}{[A] + K_D} = \frac{V_{\max}[A]}{[A] + K_D} \quad \text{where } V_{\max} \equiv k_2[P_0]$$

(see Slide No. 2 for derivation)

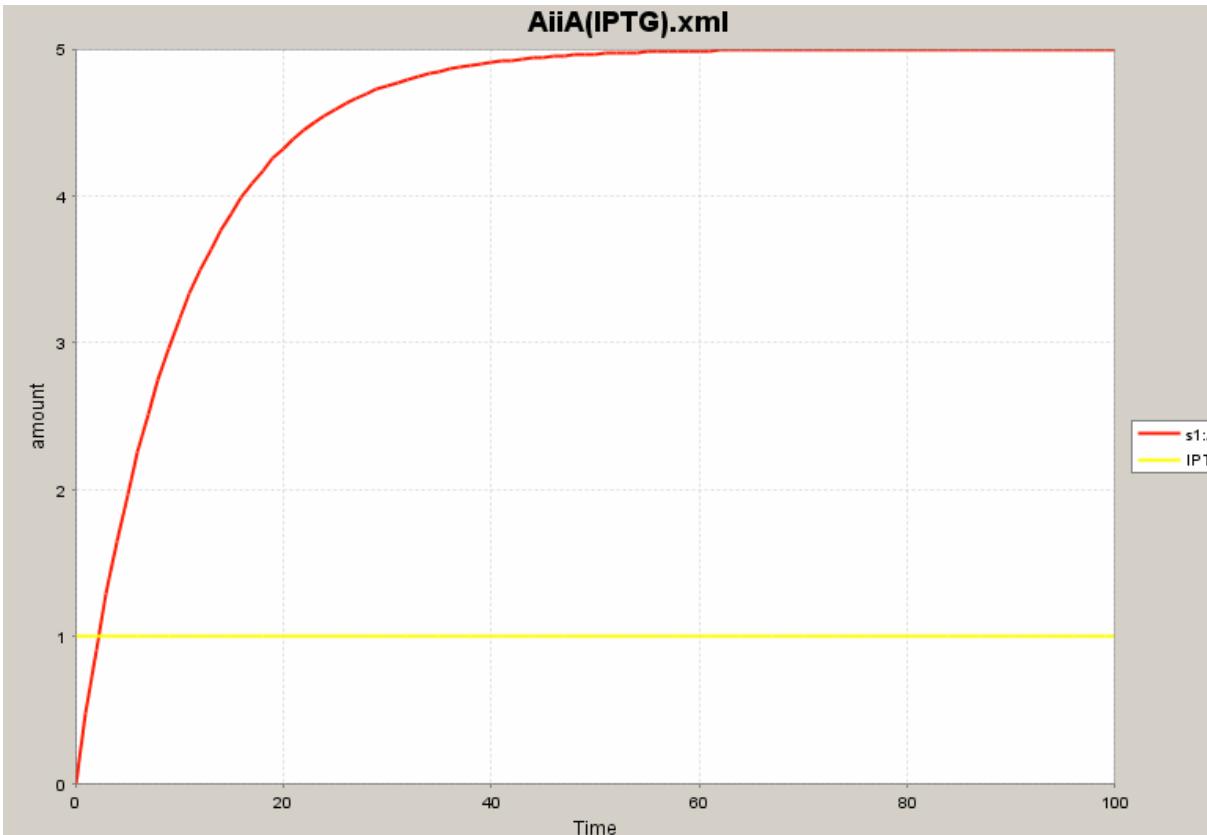
The total change in protein concentration includes protein degradation:

$$\frac{d[Z]}{dt} = \frac{V_{\max}[A]}{[A] + K_D} - \delta_{aiiA}[Z]$$

$$(11) \quad \frac{d[aiiA]}{dt} = \frac{V_{\max}[IPTG]}{[IPTG] + K_D} - \delta_{aiiA}[aiiA]$$



**Fig.5a:** Diagram for model in cell designer



**Equation:**

$$\frac{d[\text{aaiA}]}{dt} = \frac{V_{\max} [\text{IPTG}]}{[\text{IPTG}] + K_D} - \delta_{aaiA} [\text{aaiA}]$$

**Known or measurable parameters:**

- IPTG, aaiA degradation

**Parameters to extract from model:**

- $V_{\max}$ ,  $K_D$

**Fig.5b:** Output from the model in Fig.5a

Input values used to generate above graph:

$$\delta_{AiiA} = 0.1 \quad (\text{real value to be found})$$

$$K_D = 1.0 \quad (\text{real value to be found})$$

$$V_{\max} = 1.0 \quad (\text{real value to be found})$$

$$[\text{IPTG}] = 1.0$$

# Modelling the Overall System

Assumptions:

AHL is diffusing freely throughout the system

## Three resulting equations describing the Overall System:

(from the previously derived equations 5, 7, 8, 10, 11)

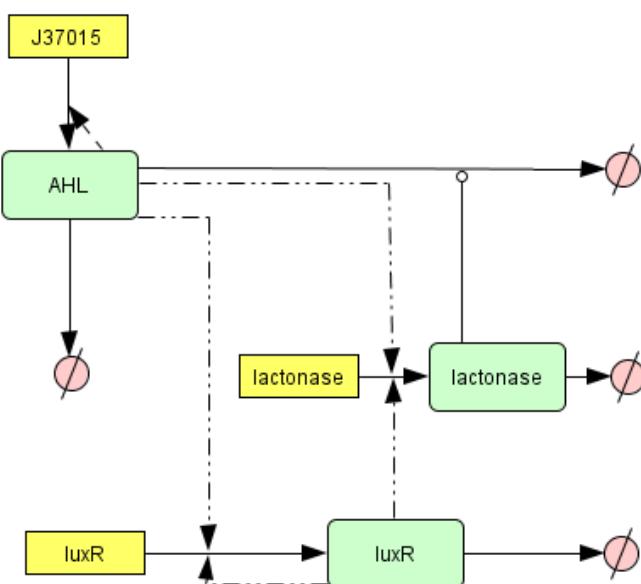
$$\frac{d[AHL]}{dt} = \frac{V_{max} [AHL]}{[AHL] + \frac{K_D}{\lambda K_{D\alpha}}} - \frac{k_2 [aiiA][AHL]}{K_m + [AHL]} - \delta_{AHL}[AHL]$$

Production of AHL

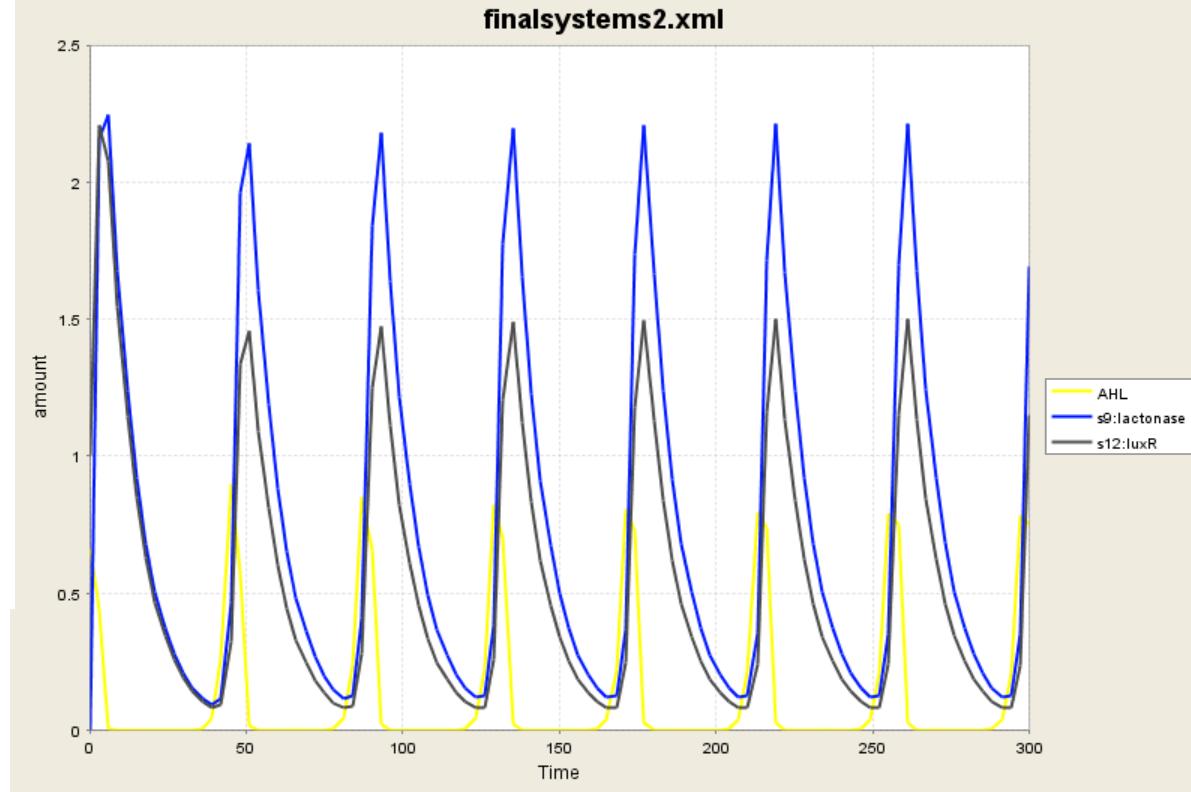
Degradation of AHL by enzyme

"Natural" degradation of AHL

$$\frac{d[aiiA]}{dt} = \frac{V_{max} [AHL][LuxR]}{[AHL][LuxR] + \frac{K_D}{K_{D\alpha}}} - \delta_{aiiA}[aiiA]$$
$$\frac{d[LuxR]}{dt} = \frac{V_{max} [AHL][LuxR]}{[AHL][LuxR] + \frac{K_D}{K_{D\alpha}}} - \delta_{2LuxR}[LuxR]$$



**Fig.6a:** Diagram for model  
in cell designer



**Fig.6b:** Output from the model in Fig.6a:  
We are getting oscillations !!!

- To gain some qualitative insight we will initially work under the rapid equilibrium approximation. This approximation assumes that the timescale of protein-protein and protein-DNA interactions are significantly faster than the other chemical reactions and thus we can consider these protein reactions to be at equilibrium