- Announcements
- Pre-lab Lecture
 - Module 1 overview
 - DNA extraction principles
 - Today in Lab: M1D1

Announcements

- BE (and other) seminar series:
 - Seminar posters across from BE HQ on 3rd floor
 - Full schedule linked from BE website
 - Part of professional development
- Different equipment for different volumes/tasks
- · Begin notebooks today! ->dates TBA by Mon. store

From protocol to lab notebook

- 1. Begin by adding the correct amount of water to a 200 ul PCR tube. Add that amount +1 ul to a second PCR tube.
- Next add the primers to each reaction. Be sure to change tips between additions.
- Next add template to the first reaction tube.
- 4. Finally add PCR Master Mix to each tube, pipetting up and down to mix. Leave your tubes on ice until the entire class

Statement of purpose: Today we will design primers to [do xyz task]. Then we will prepare [xyz DNA] by PCR to use as [xyz component] for later cloning.

Design primers for GFP insert (M1D1 Part 1)
See attached Word document.

PCR to make GFP insert (M1D1 Part 2)

Copy protocol and fill in exact volumes for #1.

Optionally confirm (say, with checkboxes) key details such as adding Master Mix <u>last</u>, template <u>only</u> to experimental sample.

Add unique notes: Rxn ready at 3 pm → on ice → thermal cycler started at 4.

EN example

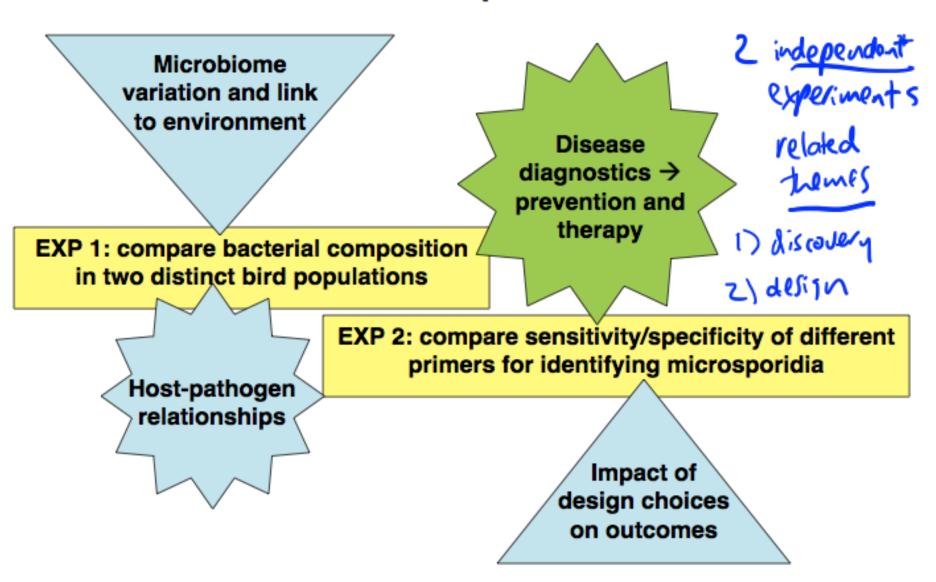
OR write out

Limit-write + take a pill

- Thanks to Shannon's UROP students!
- Protocols
- Linked Excel calculations
- Linked images
- For 20.109: front and back matter still matter

 Note: Can simultaneously use desktop and web versions, one per partner, on one laptop

Module 1 conceptual overview



Bird microbial communities: context

What is our primary research question?

What are the broader impacts of our research?

You will amplify the I 6S rRNA gene to profile
the microbiome of New England gulls
herring quil (M)
/ing-billed gulls (M, F)

Sources:



South Bay Center parking lot - South Boston Jan 4, 2014 (Photo by Darren McCollester/Getty Images)



Carson Beach, South Boston (1940): John Sanroma -Boston Public Library Flickr Stream

Samples: From Cloaca



http://www.wqed.org/birdblog/2010/04/16/anatomy-cloaca-or-vent/

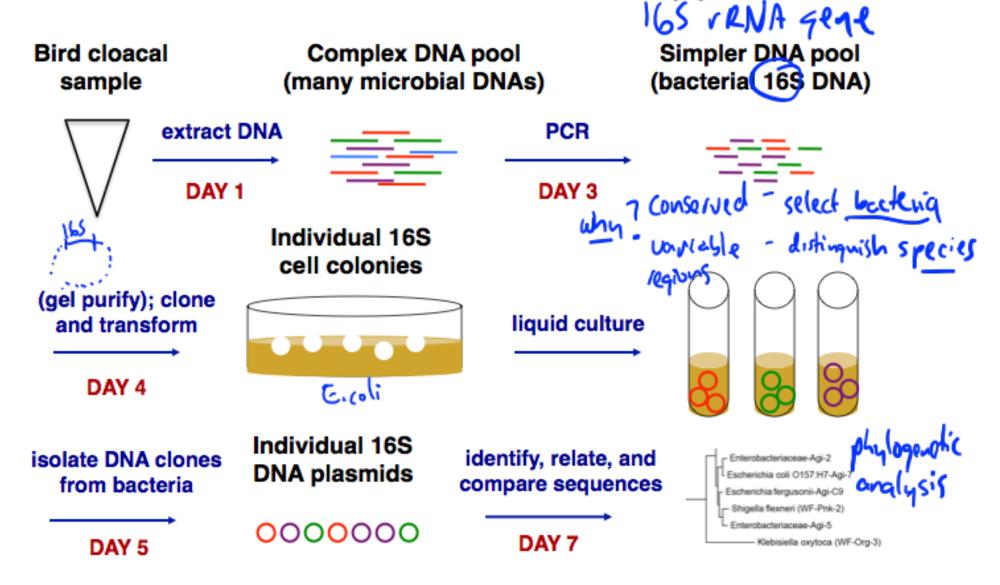
reproduction



Undisclosed to preserve bird privacy.

Thanks to Shannon H for this slide!

Bird microbial communities: approach



Step 1: DNA extraction -- background

- Stool a complex mixture! Many inhibitors of CCR
 - binders and degraders eq. | le salts or nucleares
- Rid inhibitors
 - proprietary chemicals
 - proteinase K
- Preferential isolation of microbial DNA
 − high temperature → 10°C
- Minimize cross-contamination:

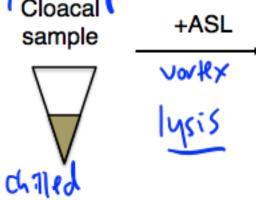
stevele filter tips : tube exchanges - label

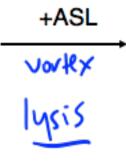
BUZI lay coats Cloacal sample

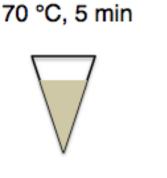
2mL

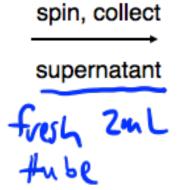
tub LS

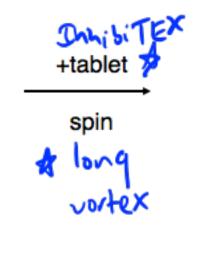
DNA extraction: initial steps

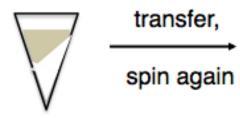


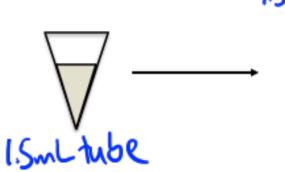














incubate: 1.5h @ 56°C

DNA extraction: later steps



Silica resin column

[qiagen.com]

t) bird DNA - Tsalt, Upt chaotropic salts disrupt 4-bonds DNA sticks to column (calombridge-see Skt)

2) keep DNA work away rest ethanol precipitates DNA (w/4) ion) Ucharge Screening

3) elute DNA - Usalt, 1 pH
electrostatic repulsion | Si-o o-p-

Today in Lab (M1D1)

- Notebook potentially collected M1D7! Ask Qs.
- Waste disposal: save all tubes, rinse 2-3x w/H₂O bottle over marked waste stream in fume hood.
- DNA extraction through enzymatic incubation
 - Many tube exchanges! May take ~ 45 min
- During 1.5 h incubation
 - Lab practical (45-50 min)
 - Prepare/label tubes for later steps
- DNA purification on column (~ 30 min)
- FNT: wiki page, start reading M1D3 paper