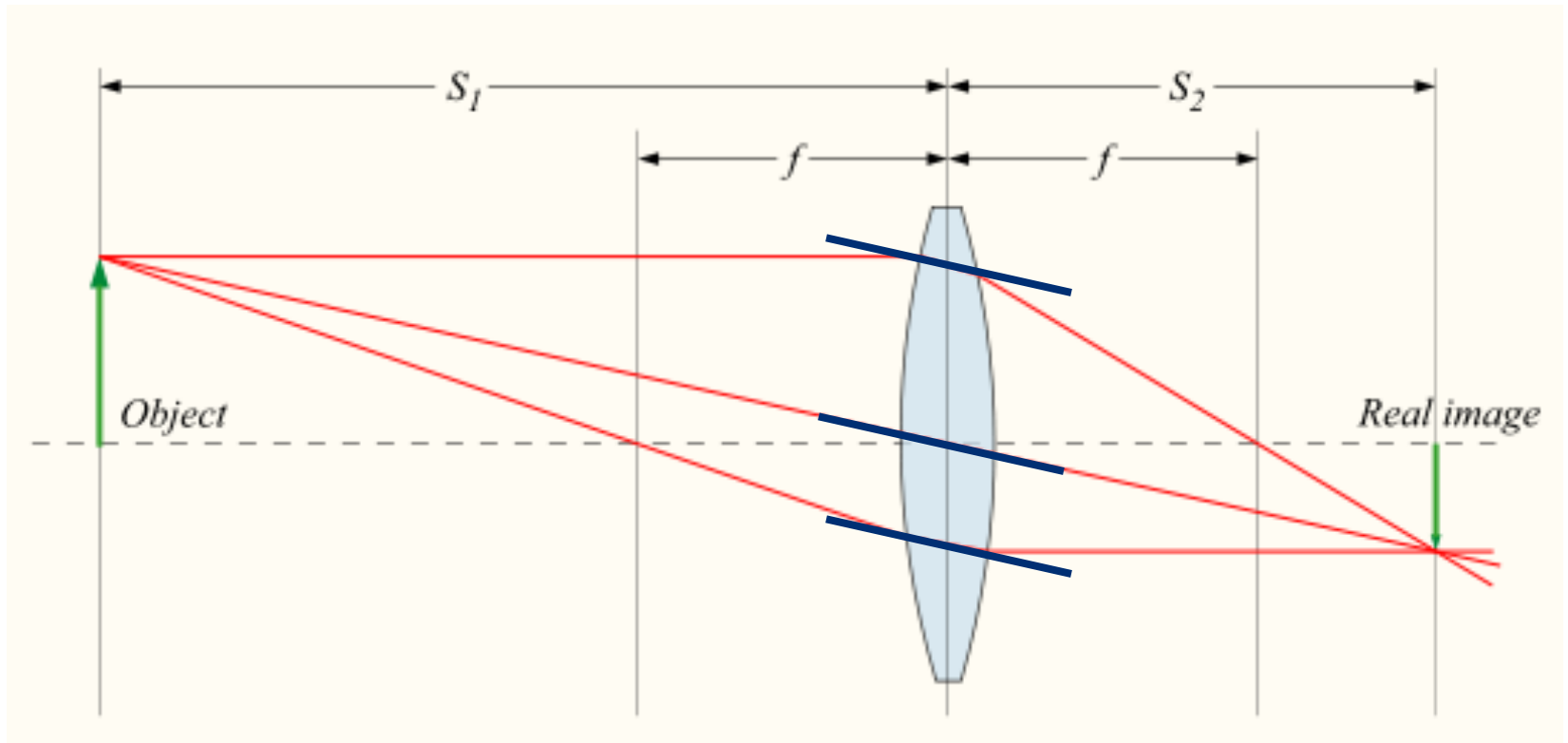


Quantitative Fluorescence  
Microscopy 2014  
*Microscope Camp*

“Every Photon is Sacred”

# The Microscope is a Combination of Lenses



# Not All Objective Lenses Are Created Equal

## Common Objective Types

10x Achromat

10x Fluorite

10x Apochromat

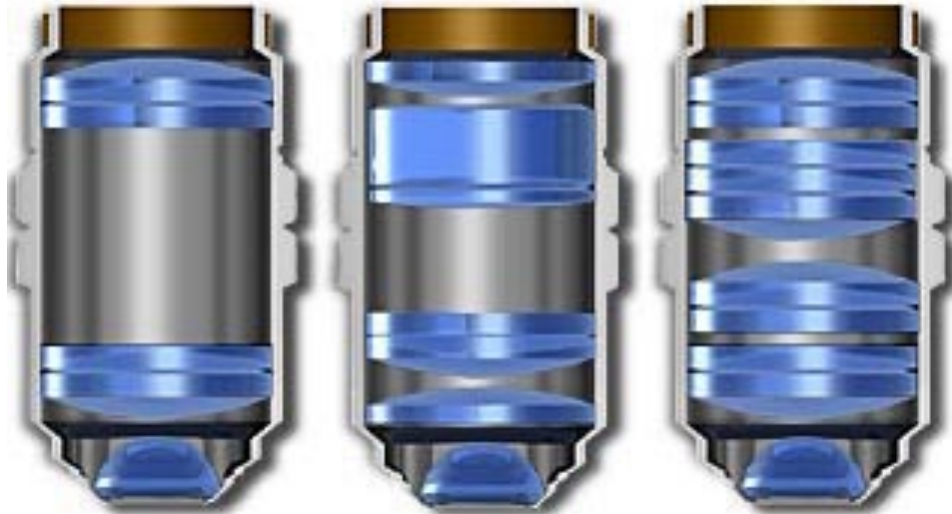


Figure 2

## Cover Glass Correction

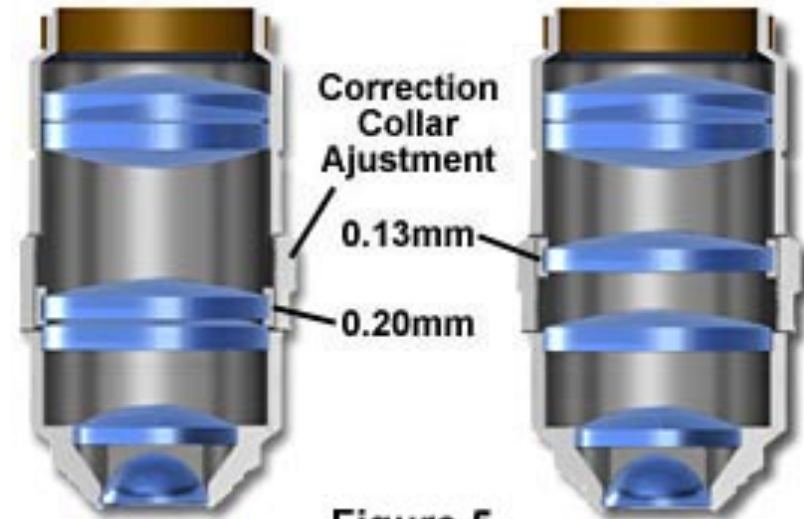


Figure 5

## Corrections:

Color - Achromat, Apochromat

Flat Field - Plan

Immersion Media

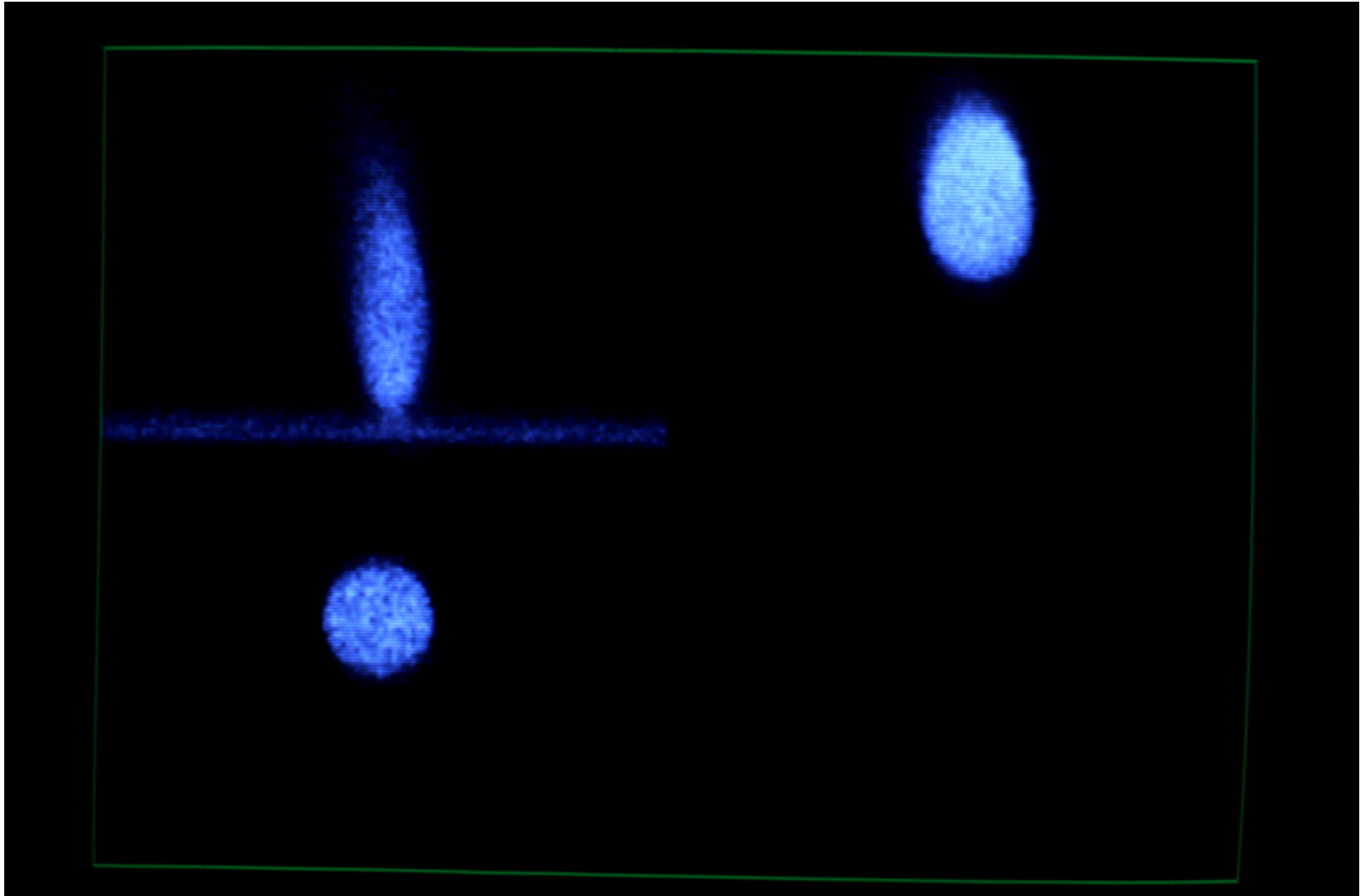
Cover Glass

Polarization

UV, IR transmission

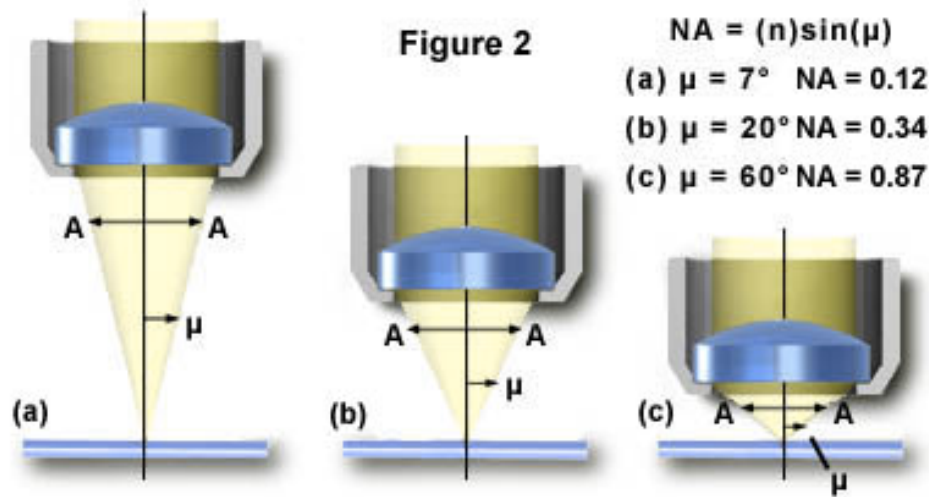
The more correction that a lens uses, the less transmission

# Index mismatch





# Resolution is Dictated by Numerical Aperture



Objective Numerical Apertures

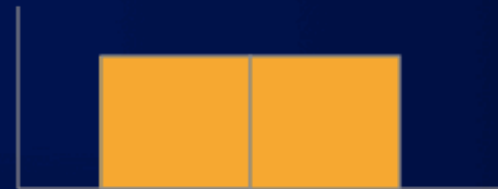
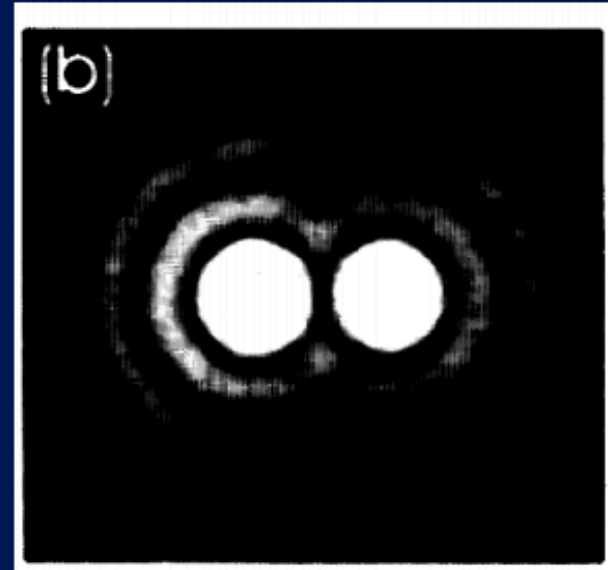
Magnification	Plan Achromat (NA)	Plan Fluorite (NA)	Plan Apochromat (NA)
0.5x	0.025	n/a	n/a
1x	0.04	n/a	n/a
2x	0.06	n/a	0.10
4x	0.10	0.13	0.20
10x	0.25	0.30	0.45
20x	0.40	0.50	0.75
40x	0.65	0.75	0.95
40x (oil)	n/a	1.30	1.00
60x	0.75	0.85	0.95
60x (oil)	n/a	n/a	1.40
100x (oil)	1.25	1.30	1.40
150x	n/a	n/a	0.90

Table 1

The smaller the NA, the bigger the focal spot,  
and the less resolution obtained –  $R \sim \lambda/NA$

# Nyquist Theorum

- How many times(*frequency*) must a sample be measured to be sure of the measurement?
  - Temporal and spatial frequency sampling may be considered the same
  - In confocal we deal with spatial frequency
- This is IMPORTANT.
  - More than 2.3 samples resel is a waste and is oversampling
  - less than 2.3 misses useful information



1 sample/resel no separation



2.3 samples/resel

**CBI**



# Resolution

$$d = \frac{0.61\lambda}{NA}$$

Therefore with 488 nm light, and a 100X 1.4 NA oil immersion optic

$$D = 0.61 \times 488 / 1.4 \\ = 212 \text{ nm}$$

Mag	resolution	
10X	0.3NA	1.1
20X	0.5NA	0.7
60X	1.4NA oil	0.2
100X	1.4NA oil	0.2

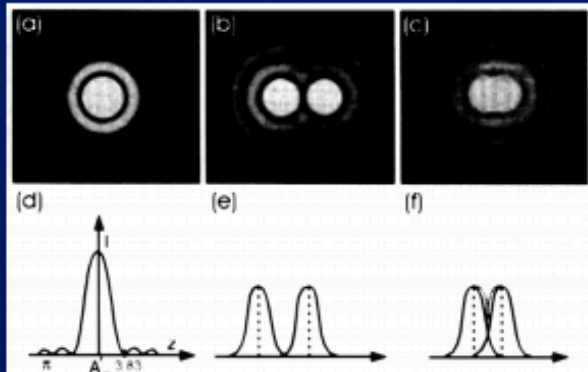
Even with short wavelength light you cannot exceed 0.2 microns resolution using the best optics available today

**IMPORTANT**

**CBI**



# Limits of confocal: Axial resolution



**Lateral resolution =  $0.61\lambda/NA$**

Diffraction limited spot  
also occurs in the axial  
or Z dimension.

$$Z_{\min} = 2\lambda n / (NA)^2$$

n = refractive index

NA = numeric aperture

$Z_{\min}$  = minimum Z resolution



60X, 1.43NA,  $\lambda = 488\text{nm}$

Lateral resolution = 208 nm

Axial resolution = 477 nm



# Köhler Illumination is Absolutely Required for Good Transmitted Light Contrast.

There are two sets of conjugate  
Optical planes in the microscope:

1. Aperture or Illumination Plane
2. Focus (Object), Image Plane

These two are Fourier transforms  
of each other -- This means that  
they are related in specific ways.

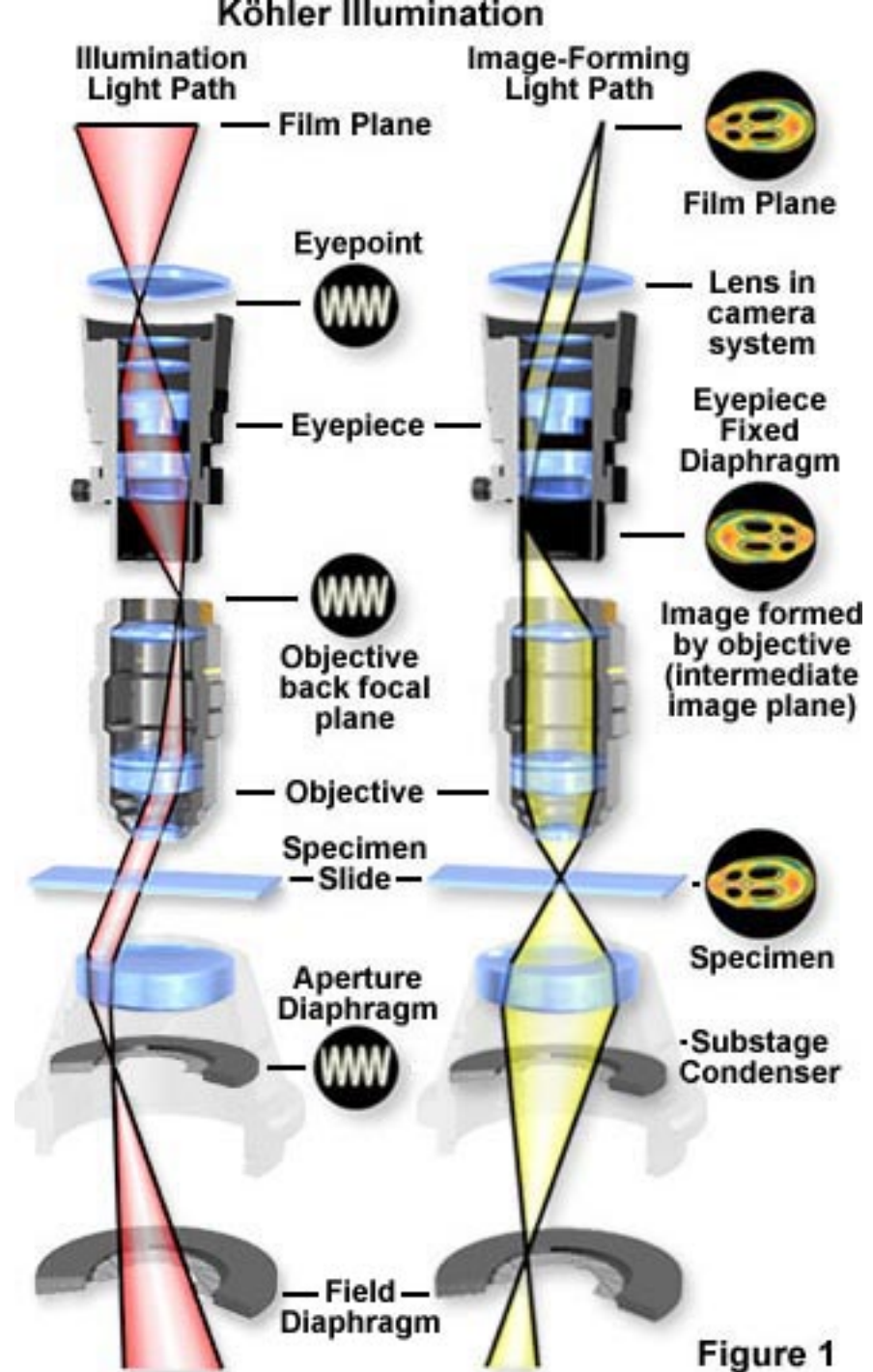
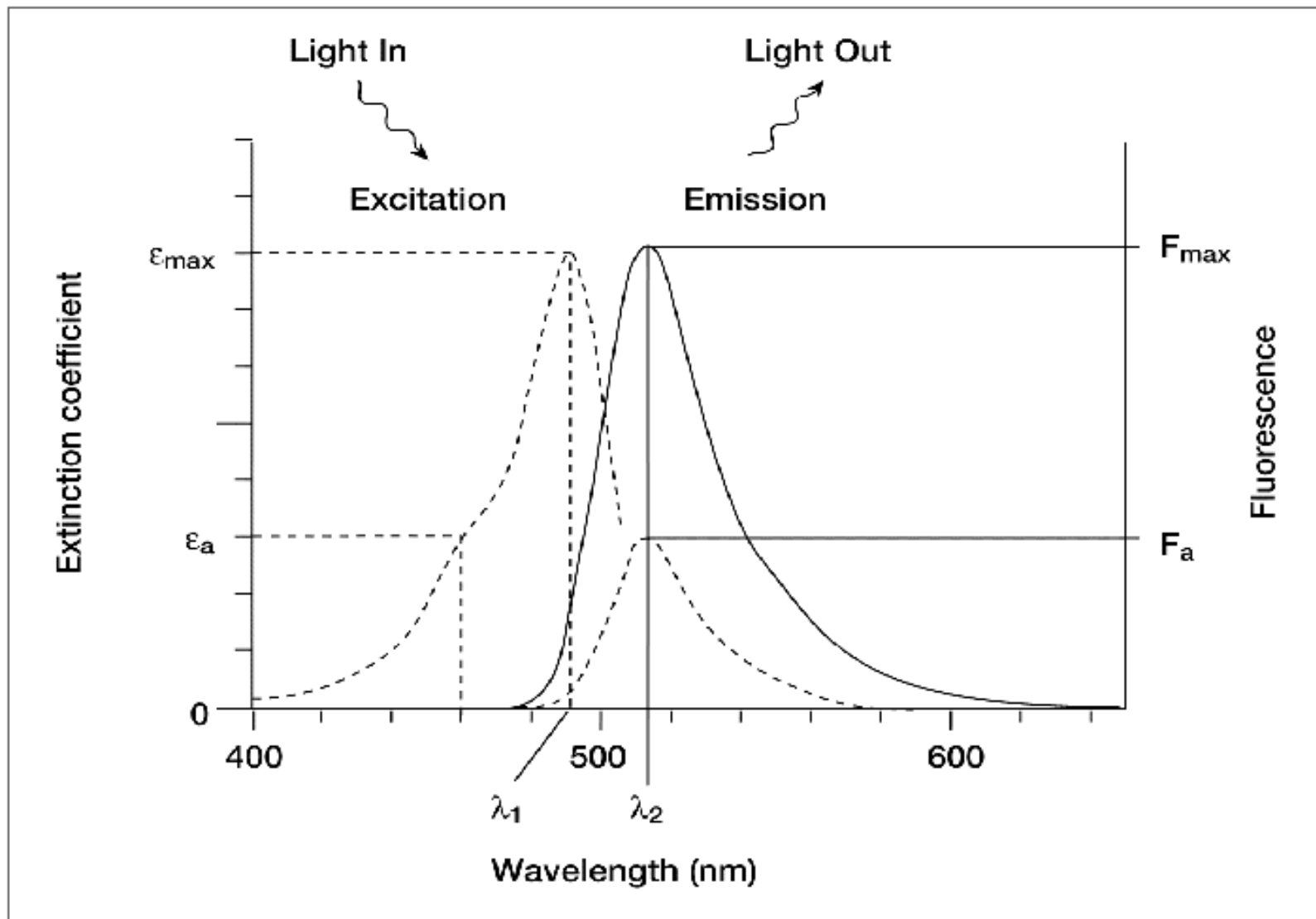


Figure 1

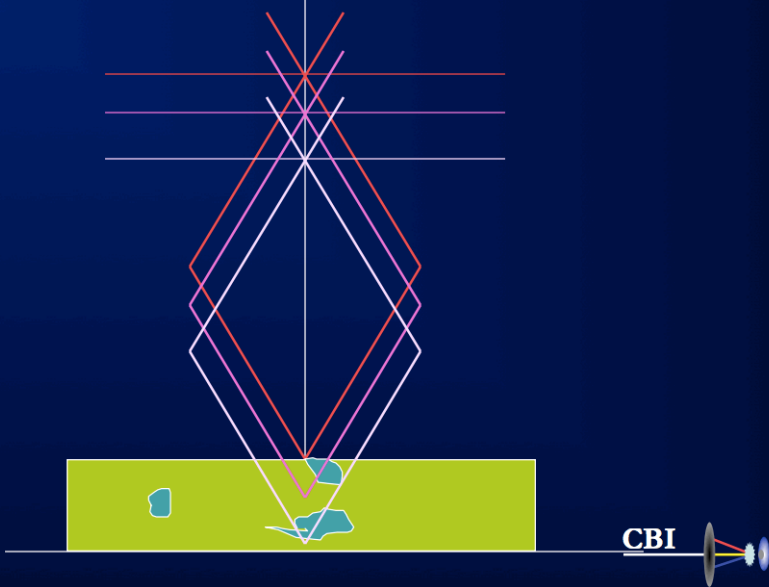
# Spectral Features of Fluorescent Molecules



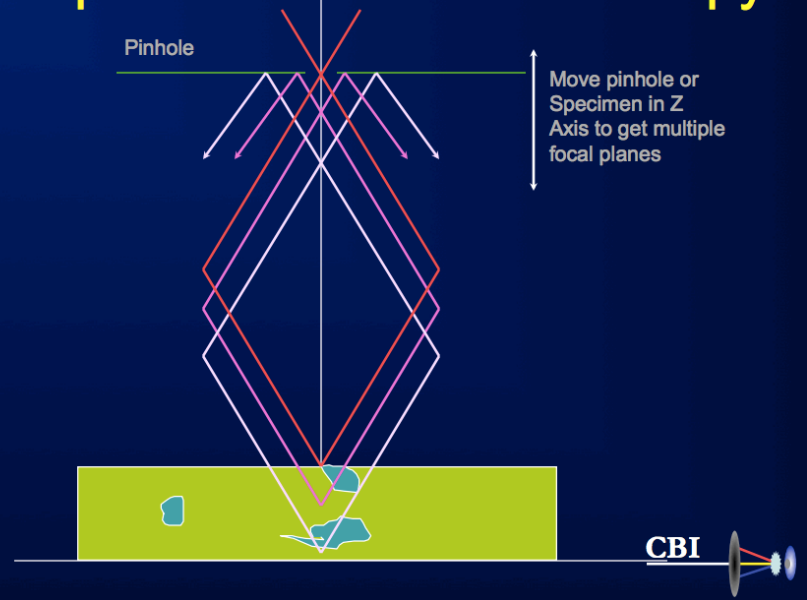


# Confocal vs Widefield

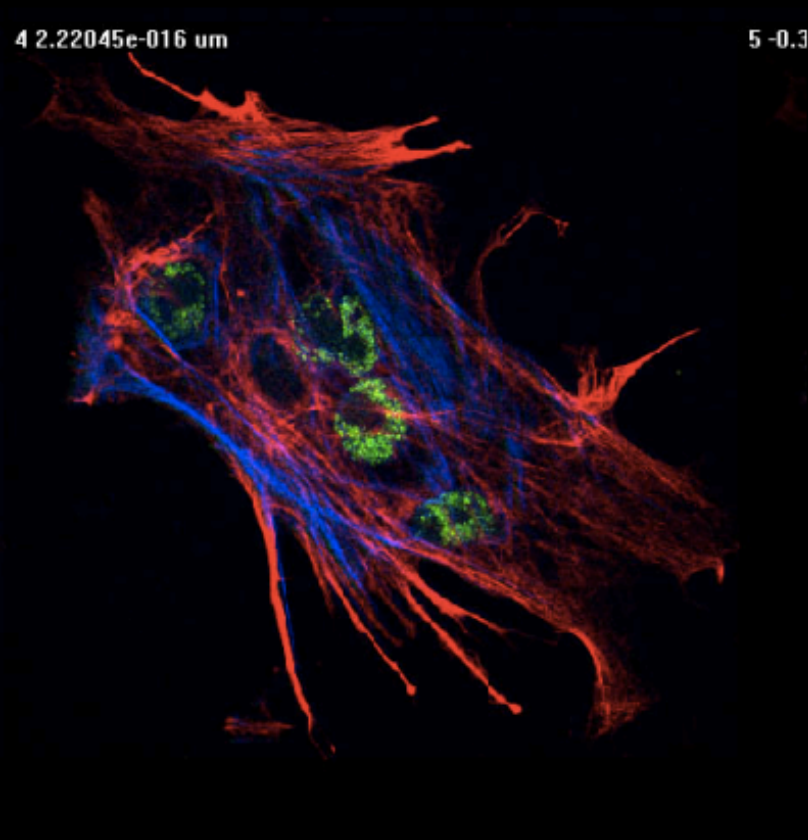
## Widefield: Multiple Focal Planes



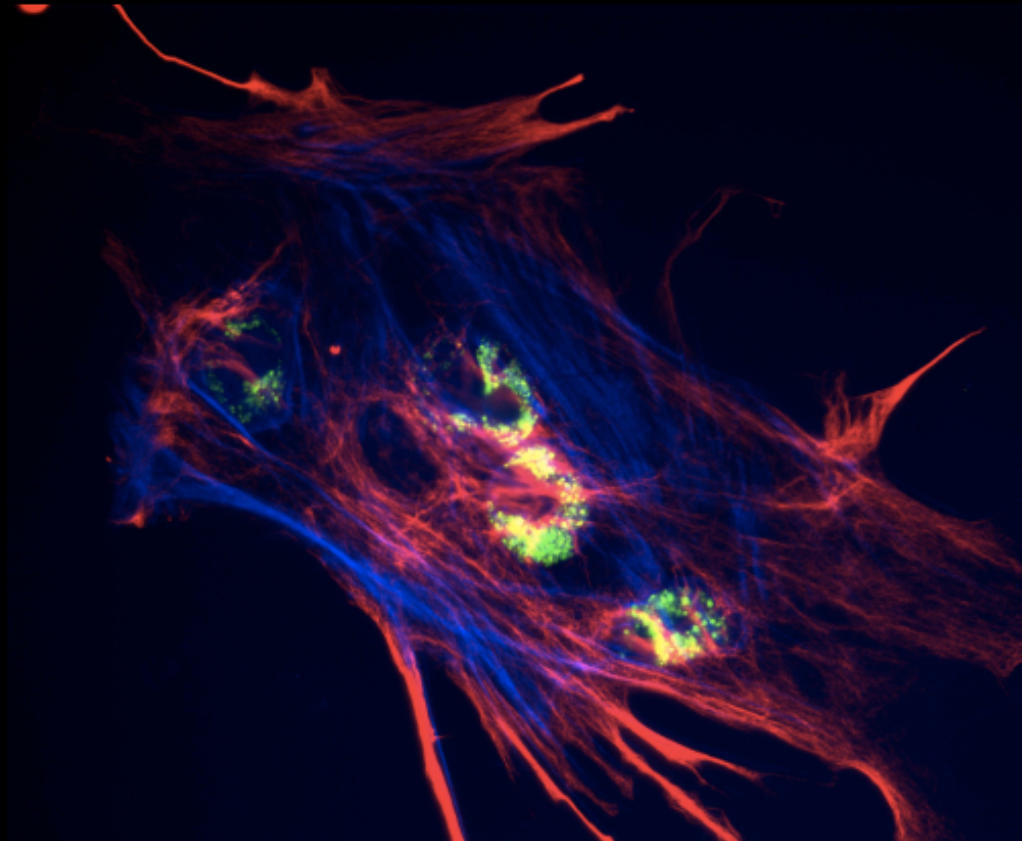
## Principal of Confocal microscopy



# Confocal vs widefield



T=10 sec min!  
Will bleach rapidly



T=0.3 secs  
Bleaches much more slowly

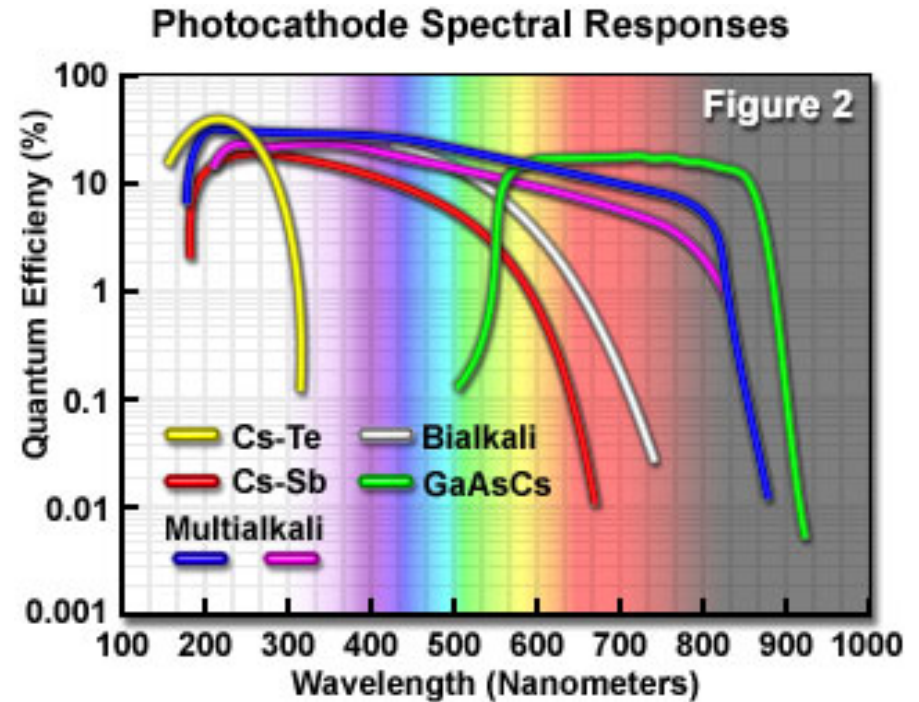
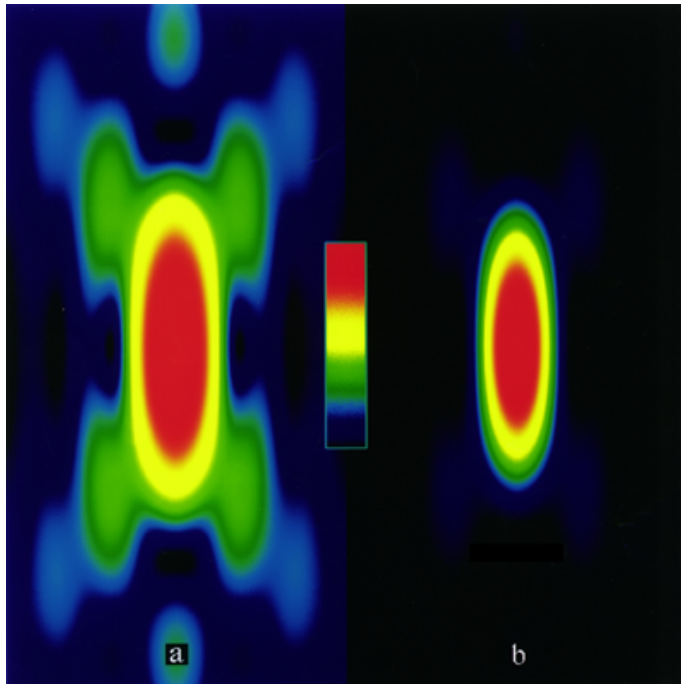
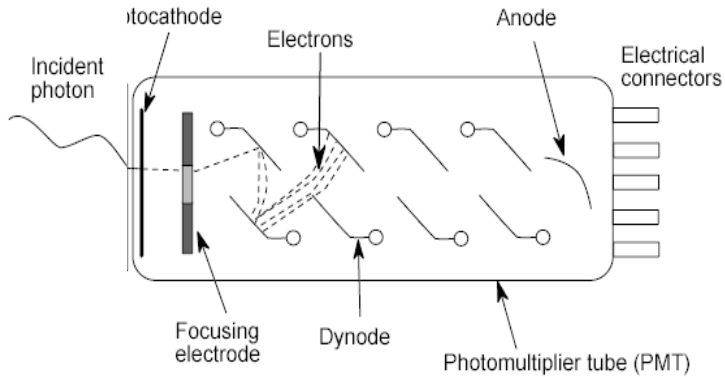




# Confocal Microscopy: Primary Uses

- Optical sectioning with high resolution Z sections of known thickness
- 3 Dimensional reconstruction and measurement
- Computer aided densitometry
- Fluorescence Recovery After Photo-bleaching (FRAP)
- Photo-activation

# Photomultiplier Tube (PMT)



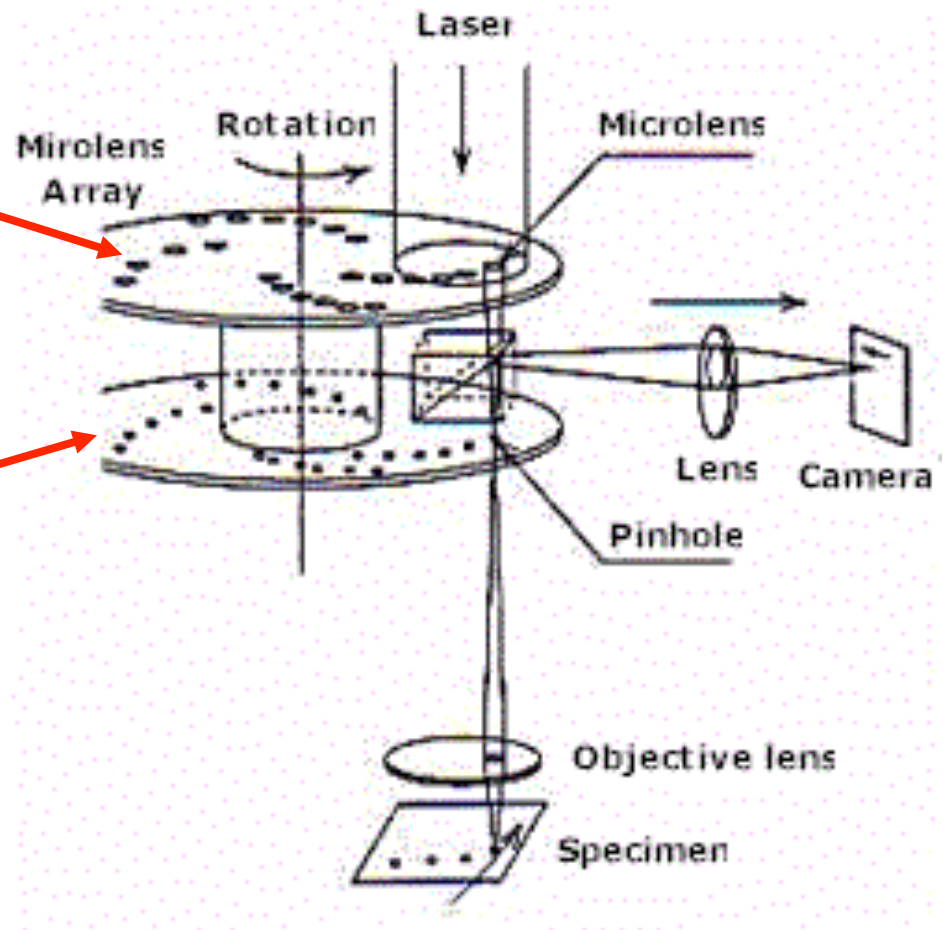
# Spinning Disk with Microlens Array

Construction of QLC100

Lens array directs >50% of the excitation light through the Nipkow Disk pinholes

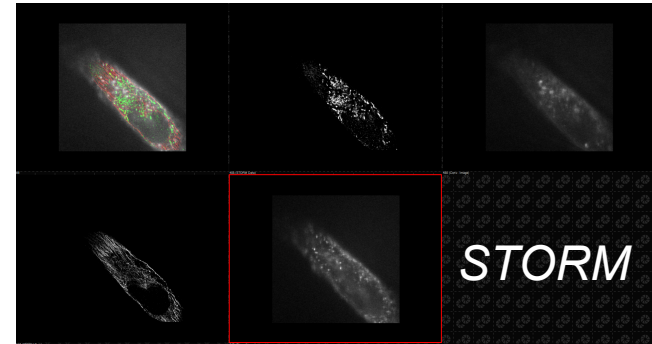
Only 1% of Nipkow disk is holes, this leads to low excitation rates

**But, faster acquisition means faster photobleaching!**

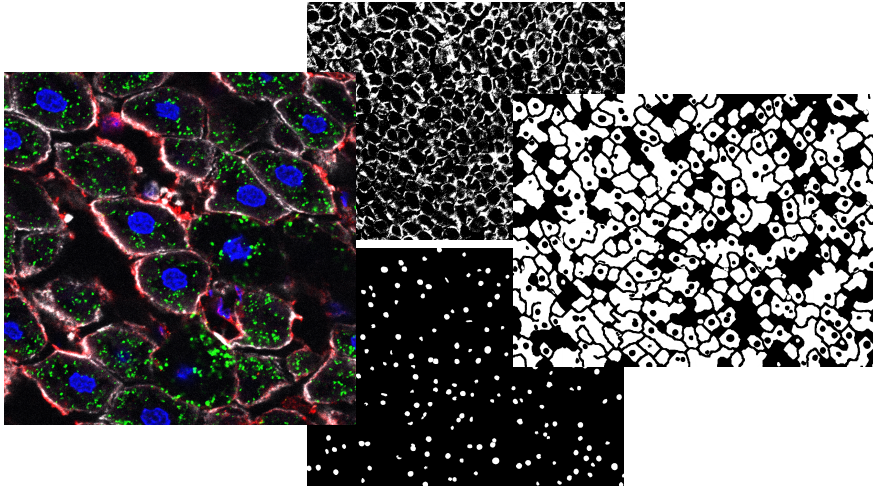


# MICROSCOPE CAMP *QFM 2014*

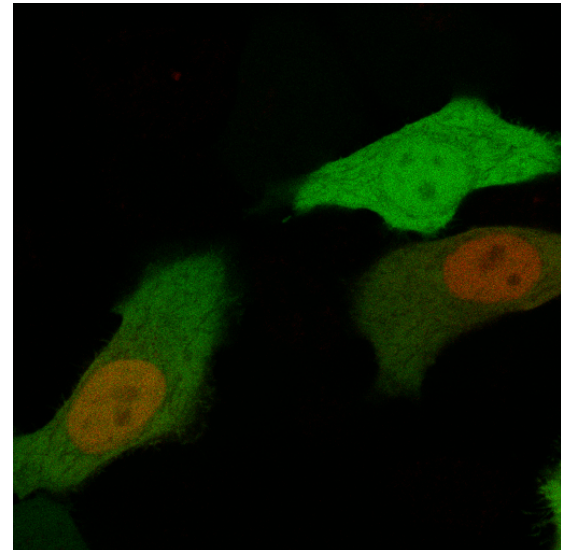
## Super Resolution



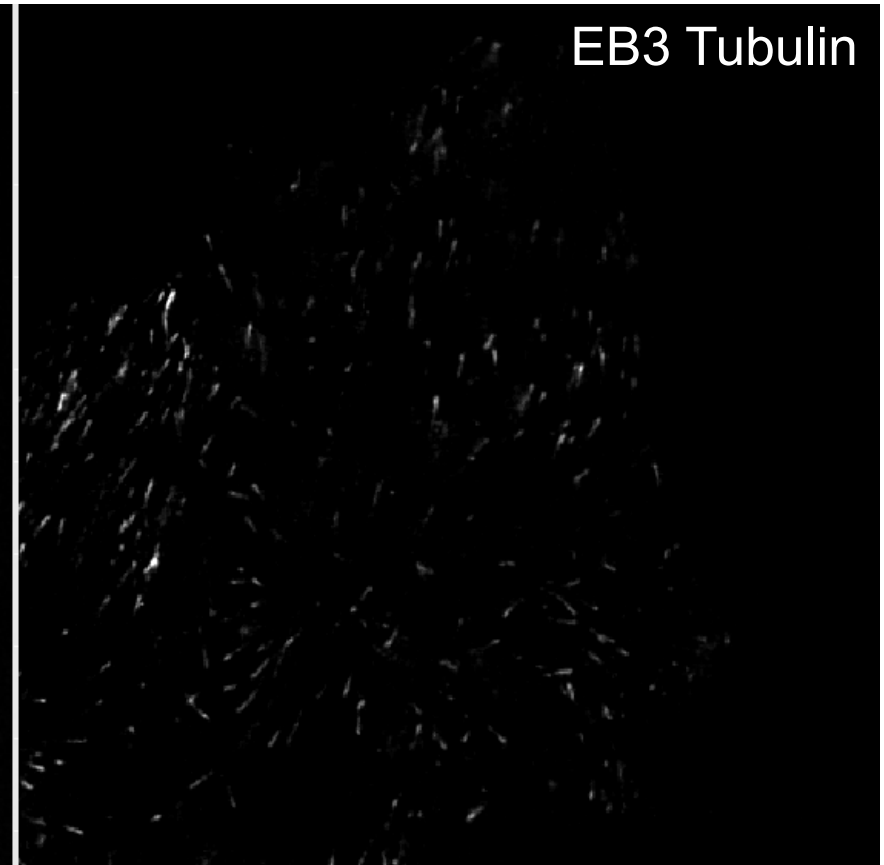
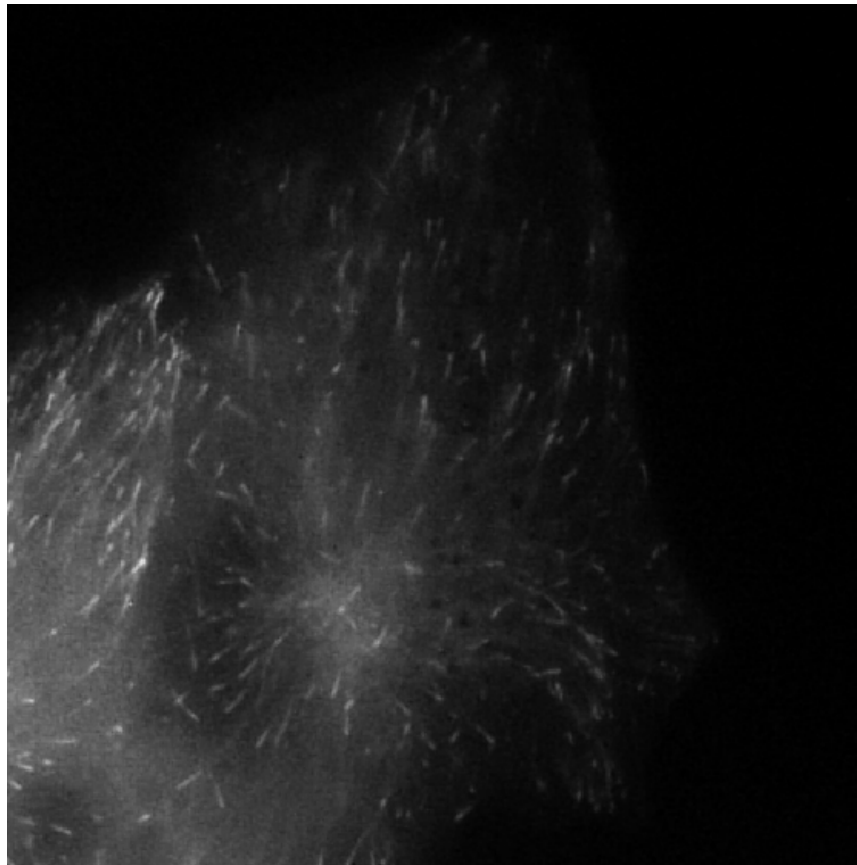
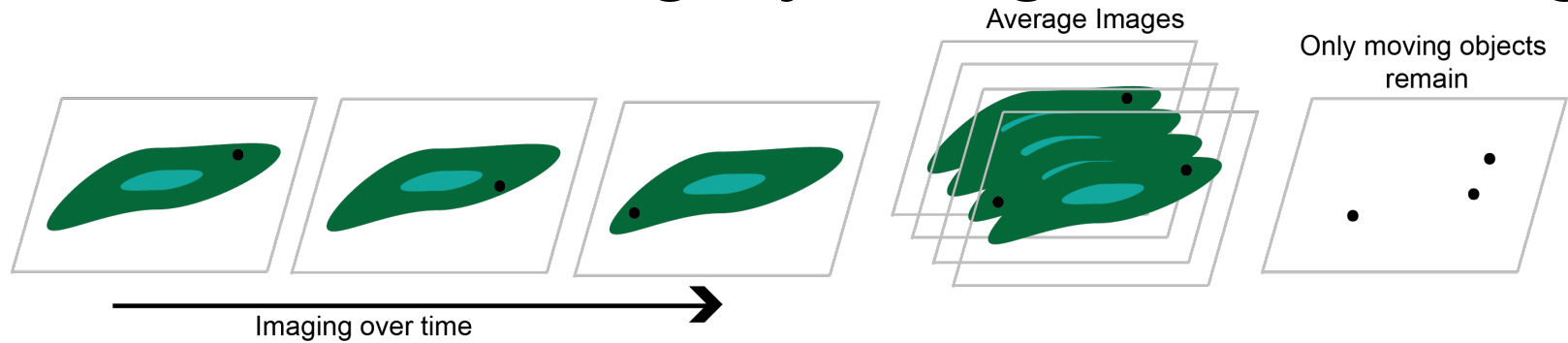
## *Image Processing*



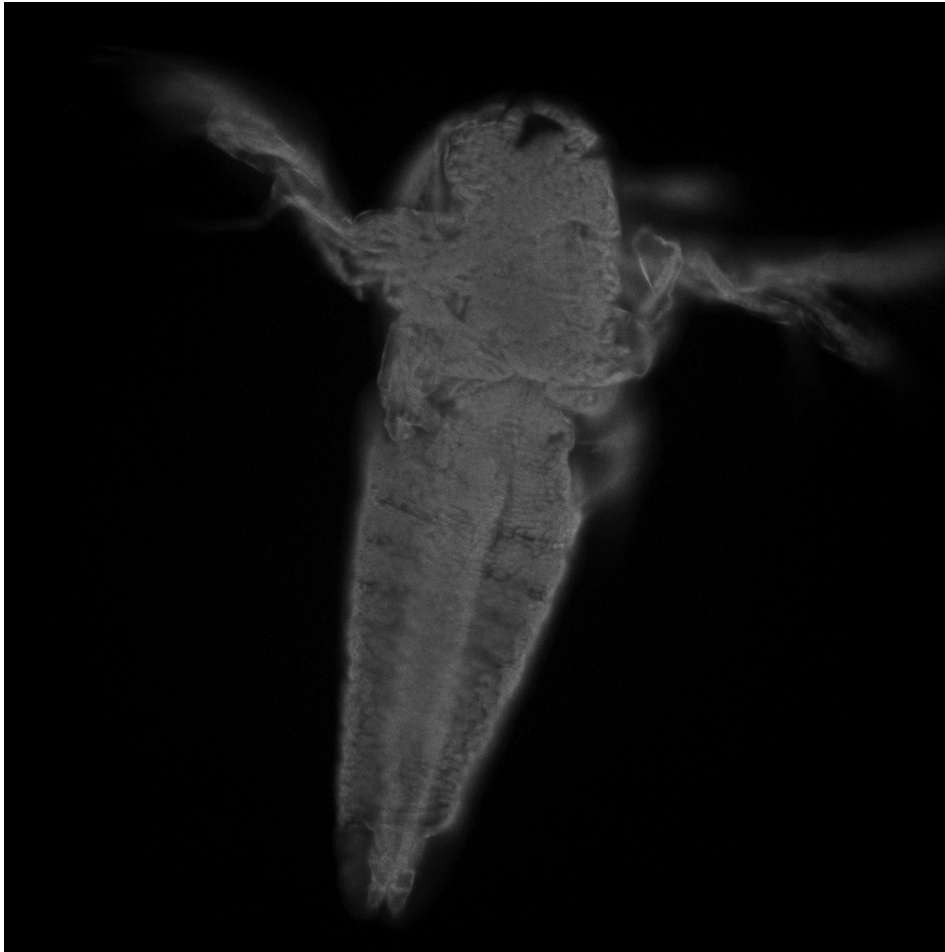
## Photo bleaching/activation



# Feature Extracting by Image Processing



# Selective Photo bleaching by binary selection





# QFM Class Information

- [http://www.mdibl.org/courses/  
Quantitative\\_Fluorescence\\_Microscopy/159/](http://www.mdibl.org/courses/Quantitative_Fluorescence_Microscopy/159/)