## **In-solution Digests (revised 8-10-07)**

## **Reagents:**

- 1. 4X digestion buffer: 8 M electrophoresis grade urea, 1.0 M Tris (pH 8.5), 8 mM CaCl<sub>2</sub> and 0.2 M methylamine. (freeze in 50 μL aliquots)
- 2. 0.2 M Dithiothreitol (DTT) solution (31 mg into 1.0 mL of water). Make up fresh.
- 3. 0.5 M Iodoacetamide (IAA) solution (23 mg into 0.25 mL of water). Make up fresh (protect from light).
- 4. 0.1 μg/μL Promega modified sequencing grade trypsin stock solution. Prepare by dissolving a 20 μg vial of trypsin (Cat # V5111) in 200 μL of 1 mM HCl. Store frozen in 20 μL aliquots for up to 3 months at -20°C, or one year at -70°C. **Do not freeze thaw.**

## **Protocol:**

- 1. Start with a protein solution of known concentration that is detergent free. Save a portion to run on a gel to assess the extent of digestion.
- 2. Add the protein solution to a 0.5 mL centrifuge tube. Take the protein sample to dryness in a speed vac. Up to 100 μg of protein may be digested using the volumes given below, unless high salt concentrations are present that would prevent the sample from being redissolved in 10 μL of digestion buffer.
- 3. Thaw the 4X digestion buffer. Add  $10 \mu L$  to the dried protein sample and vortex to dissolve.
- 4. Add 1  $\mu$ L of the DTT solution, vortex, spin down to the bottom of the centrifuge tube and incubate at 50°C in the thermocycler or water bath for 15 min.
- 4. Remove the sample from the thermocycler, let it cool for several min, add 1  $\mu$ L of the IAA solution, vortex, and incubate in the dark at room temp for 15 min.
- 5. Add an additional 2  $\mu$ L of DTT solution to the mixture, vortex, and incubate for 15 min at room temp.
- 6. Calculate how much trypsin to add for a final 1:25 enzyme to substrate ratio. For example, if starting with 10  $\mu$ g of protein, you would use 4  $\mu$ L of 0.1  $\mu$ g/ $\mu$ L trypsin stock. However, before adding the trypsin, add sufficient water so that the final volume of the digestion mixture is 40  $\mu$ L (in this example you would add 22  $\mu$ L of water).
- 7. After addition of water (step 6 above), vortex the sample, and add the trypsin solution.
- 6. Vortex gently to mix, centrifuge to the bottom of the tube and incubate overnight at 37°C.
- 7. Add 2 µL of neat 88% formic acid to stop the digestion.
- 8. If needed, run equal amounts of undigested and digested samples on a gel to assess the extent of digestion.

9. Freeze at -20°C if LC/MS analysis will not be performed within one week.	