# Illuminating Phaeodactylum tricornutum Cell Biology with a Genetically **Encodable Tag for Electron Microscopy and Subcellular Proteomics**





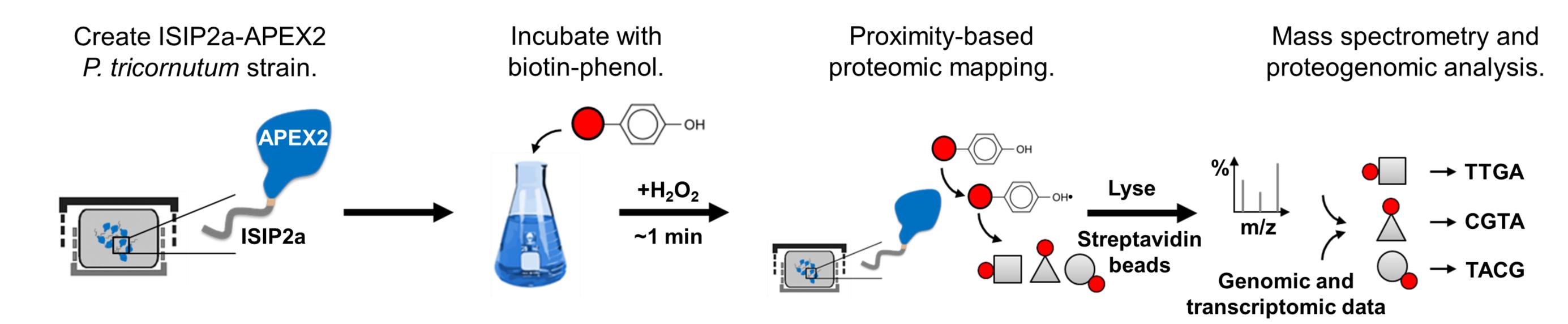




### 1. Background

Iron plays a crucial role in many key enzymes linked to photosynthesis, respiration and nitrogen fixation. Iron availability limits primary productivity in ~30% of the modern oceans. Our laboratory has identified phytotransferrins as a new group of high affinity ferric iron-binding proteins widespread among marine microeukaryotes. Phytotransferrin ISIP2a from a model diatom Phaeodactylum tricornutum internalizes ferric iron via endocytosis, but the molecular details behind ion liberation, chemical speciation and intracellular allocation remain elusive.

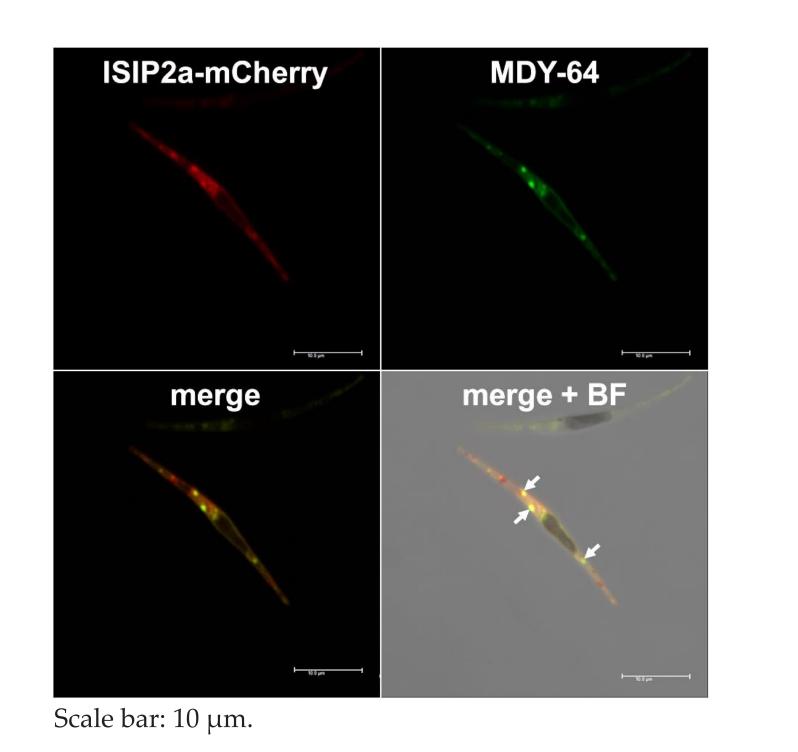
## 2. Approach



ISIP2a-APEX2

#### 3. Results

3.1 ISIP2a-mCherry colocalization with MDY-64 (membrane stain) further supports ISIP2a endocytosis model.



**PEAKS Studio 8.5** 

3.2 Confirming in vivo peroxidase activity of APEX2 with Amplex UltraRed assay.

13000

12000

11000

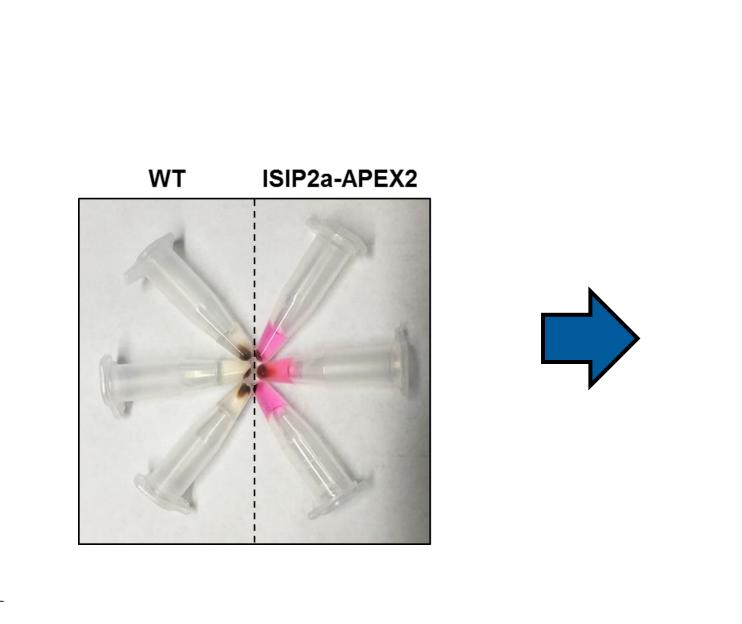
10000

9000

5000

4000

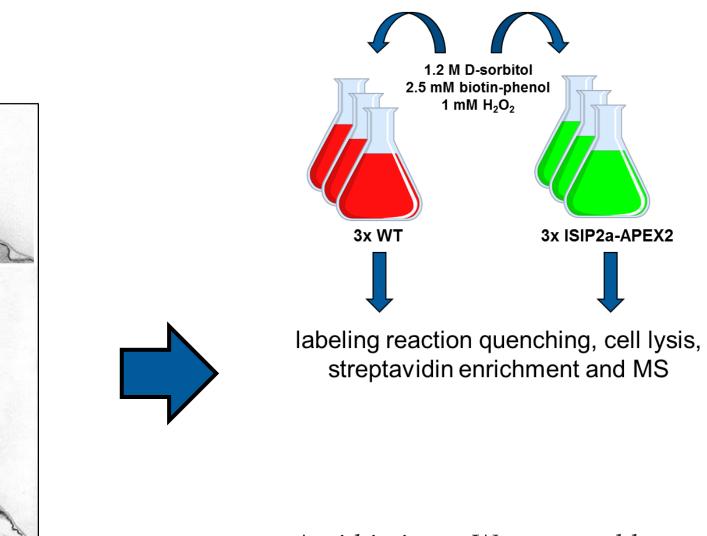
3000



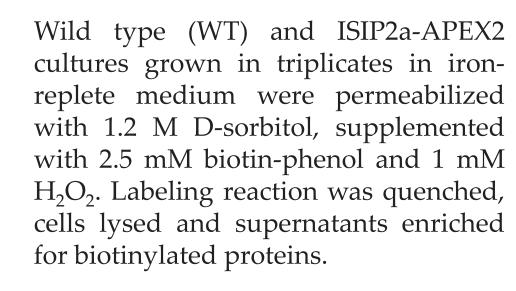
3.3 Visualizing ISIP2a-APEX2 using transmission electron microscopy (TEM).

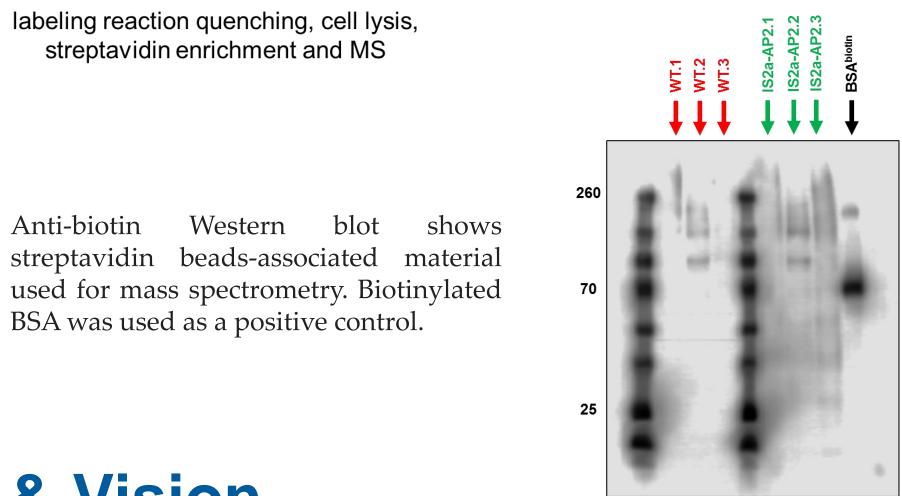
WT

Scale bar: 1 µm.



3.4 Subcellular proteomic experiment summary.





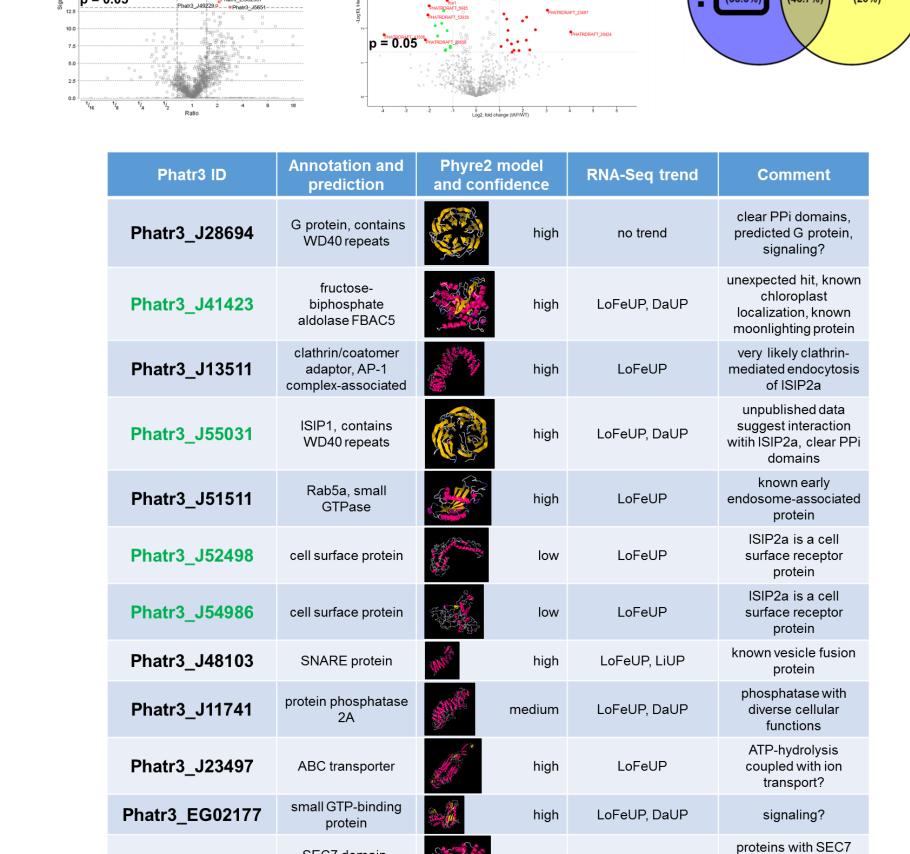
3.5 Label-free quantification, putative ISIP2a interactors and vicinal proteins.

MaxQuant

**Proteome Discoverer** 

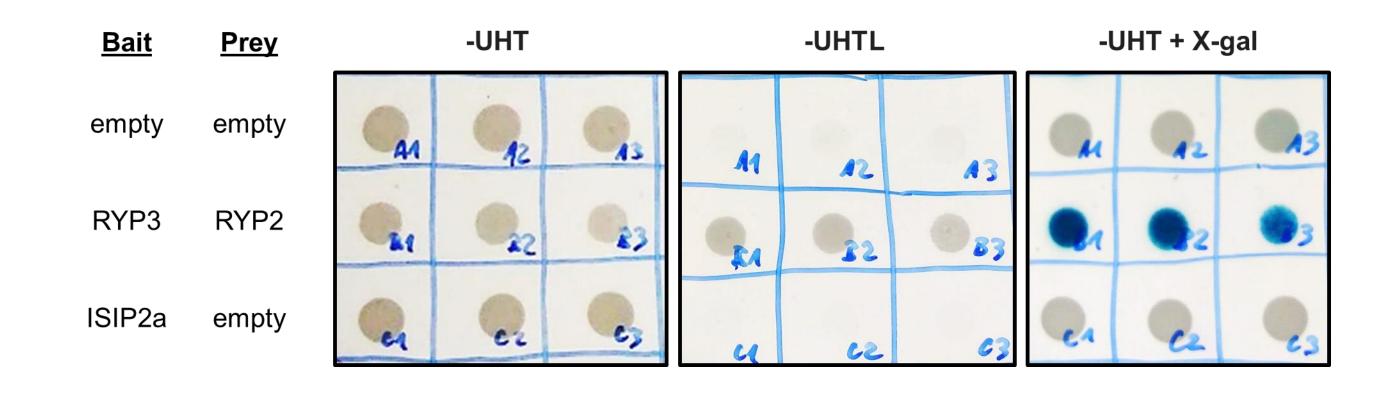
domains are involved

ISIP2a-APEX2



Highlighted in green are proteins that were highly enriched (at least 2-fold above WT) in ISIP2a-APEX2 strains in an independent Tandem Mass Tag proteomic experiment performed in quintuplicates. LiUP: upregulated in light, DaUP: upregulated in dark, LoFeUP: upregulated at low Fe (20 or 40 pM Fe').

#### 3.6 Successful implementation of a yeast two-hybrid (Y2H) screen to study protein interactions with ISIP2a.



In our Y2H screen, a protein-protein interaction leads to restored ability of yeast cells to grow in the absence of leucine as well as their ability to break down X-gal. Cells were spotted on each of the following three plates: -UHT (all strains will grow), -UHTL (only strains where protein-protein interaction occurs will grow), -UHT + X-gal (all strains will grow; strains where protein-protein interaction occurs will turn blue). Plates were incubated for 2 days at 30 °C and imaged. U: uracil, H: histidine, T: tryptophan, L: leucine. RYP3 and RYP2 are known interacting proteins from a fungal pathogen. ISIP2a alone doesn't activate reporter genes.

# 4. Next steps

- Y2H screen using a prey library containing proteins identified with MS is in progress.
- Co-localization studies in *Phaeodactylum tricornutum*.
- His tag/Strep-tag II pull-downs for additional cross-validation.

### 5. Conclusions & Vision

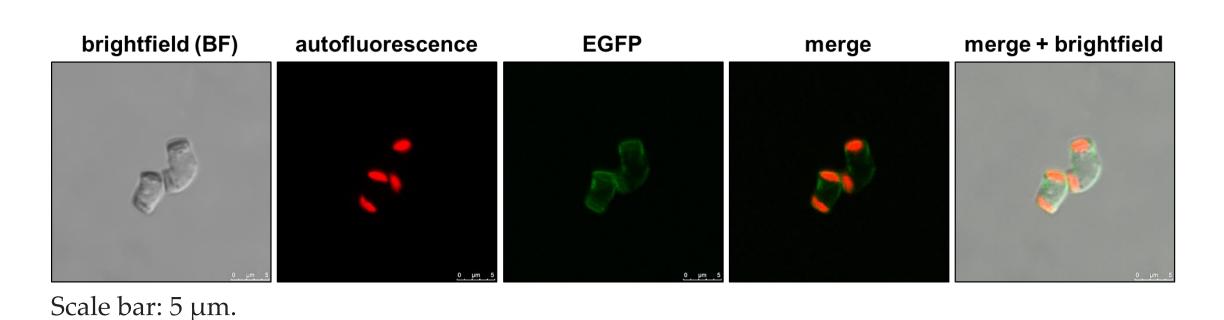
1 micron

We have implemented APEX2, a genetically encodable ascorbate peroxidase, in *Phaeodactylum tricornutum*, to **promote high resolution** protein imaging and cataloging of biological pathway-specific proteomes. This work represents the first application of APEX2 technology in a photosynthetic host. We envision it will allow us to dissect a range of outstanding cell biology questions in Phaeodactylum tricornutum and other diatoms as they relate to their prominent role in global biogeochemical cycles, unique evolutionary history, and biotechnological potential.

BSA was used as a positive control.

# 6. Future applications of APEX2 in diatoms

Proteomic characterization of a silica deposition vesicle (SDV) in *Thalassiosira* pseudonana (Tp) – a biomineralization model system.



Ornamented silicified cell walls are the hallmark of diatom biology. Biosilica morphogenesis proceeds inside a silica deposition vesicle (SDV). A revisited Thalassiosira pseudonana conjugation protocol was used to localize a known SDV-associated protein. We are currently preparing strains expressing this protein fused to APEX2.

# Acknowledgements

We thank Pardis Gholami for help with diatom culture work; Tom Deerinck, Mason Mackey, Andrea Thor, Daniela Boassa and Mark Ellisman for help with TEM; Majid Ghassemian, Yanbao Yu, Marian Kalocsay and Steven Gygi for help with mass spectrometry and proteomic data analysis; Sinem Beyhan for help with Y2H work.

## Funding

This research is supported by the Department of Energy, Office of Biological and Environmental Research (BER), grant DE-SC0018344 (AEA), and by the Gordon and Betty Moore Foundation (GBMF), grant GBMF4958 (JT).

#### Affiliations

<sup>1</sup>Harvard Medical School, Department of Systems Biology, Boston, MA 02115 <sup>2</sup>Microbial and Environmental Genomics, J. Craig Venter Institute, La Jolla, CA 92037 <sup>3</sup>Integrative Oceanography Division, Scripps Institution of Oceanography, UC San Diego, La Jolla, CA 92037

