









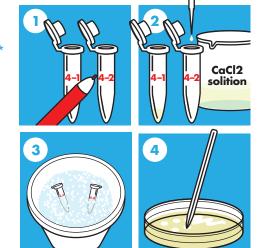


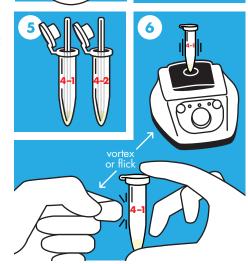


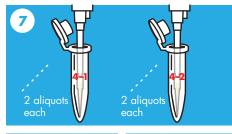
Patch 4-1 and 4-2 bacteria on LB petri dishes **

DAY OF LAB

- 1. Label 2 small microfuge tubes either "4-1" or "4-2."
- 2. Pipet 200 µl of CaCl2 solution into each microfuge tube.
- 3. Place the tubes on crushed ice.
- 4. Using a sterile pipet tip, toothpick or innoculating loop, scrape a patch of cells off the petri dish**. Avoid scraping up the agar.
- 5. Swirl the in cells the appropriate tube with cold CaCl2 then repeat for the other patch of bacteria.
- 6. Gently vortex the cells to resuspend them. If no vortex is available, gently flick and invert the microfuge tubes, then return them to your icebucket.
- 7. Retrieve 2 aliquots of each plasmid for a total of 4 samples (2x pPRL, 2x pGRN).
- 8. Label one of the **pPRL** tubes "4-1" and label the other **pPRL** tube "4-2."
- 9. Label one of the **pGRN** tubes "4-1" and label the other **pGRN** tube "4-2."

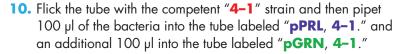












- 11. Flick the tube with the competent "4-2" strain and then pipet 100 µl into the tube labeled "pPRL, 4-2" and an additional 100 µl into the tube labeled "pGRN, 4-1"
- 12. Incubate the tubes on ice for ~5 minutes.
- 13. Heat shock the transformation reactions at 42°C for 90 seconds exactly.
- 14. Move the tubes to a rack at room temperature and add 0.5 ml LB to each. Close the caps, and invert the tubes to mix the contents.
- 15. Label the media-side of the LB + amp petri dishes to indicate the strain you've used ("4-1" or "4-2") and the DNA you've transformed them with ("pPRL," "pGRN")
- 16. Pipet 250 µl of each sample onto the media of the appropriate petri dish. Spread the sample evenly across the dish with a sterile spreader.** Discard spreader and remainder of transformation mix in 10% bleach solution.
- 17. Incubate petri dishes, media side up, for 2 days at 30°C.

