# Condensed down Mycoplasma Test Protocol – reference protocol = MycoSensor PCR Assay Kit (Agilent Tech - #302109)

## **Extraction of Cell Culture Supernatant and Cells**

- 1. Set up heat block to 95°C
- 2. Take out 100 uL of media from test cell culture into a microcentrifuge tube
- 3. Scrape cells into 5 mL of 1X TC PBS and spin down 3 min at 1000 RPM in 15 mL centrifuge tube
- 4. Wash again with 1 mL 1X TC PBS and spin down 3 min at 1000 RPM
- 5. Resuspend again in 1 mL 1X TC PBS AND COUNT CELLS
- 6. Count out 100,000 cells into a new tube and again spin down (3 min at 1000 RPM)
- 7. Aspirate supe and add 100 uL of sterile dH<sub>2</sub>O
- 8. Boil tubes with 100 uL of media and cells in heating block set earlier. Media goes for 5 min. Cells go for 10 min.
- 9. Spin the boiled tubes briefly to get liquid off sides
- 10. Resuspend the kit-included StrataClean resin (**Green** lid) by vortexing the tube until all pellet is suspended in liquid
- 11. Add 10 uL of StrataClean to each tube and flick tubes to mix the resin and extracts
- 12. Spin all cells for 1 min to pellet the resin
- 13. Transfer supes to new tubes
- 14. Both can be stored at 4°C for several days

## **PCR** of the Samples

### SET UP:

#### **RXN MM**

Component	Volume Added (uL)	2 samples
Sterile dH₂O	19.5	39 /
MgCl <sub>2</sub> (25 mM)	8	16 🗸
5X Taq Buffer	10	20 🗸
dNTP mix (from kit)	2	4 🗸
Mycoplasma Primer Mix (from kit – red lid)	1	2 🗸
Internal Ctrl template (from kit)	4	8 🗸
> Taq polymerase	0.5	1
RXN MM VOLUME	45 uL (each)	Template = add 5 uL

#### TOTAL RXN VOLUME = 50 uL

- Use Thermocycler program for Mycoplasm test (takes ~ 2 hours)
- Run on 2% agarose gel (0.8g agarose + 40 mL of NEW 1X TAE buffer) + 1.5 uL EtBr in small gel box
  - o Ladder Recipe: 1 uL LMW ladder + 2 uL 6X Loading Dye + 7 uL dH₂O (TOT = 10 uL)
  - o Sample Recipe: 17 uL + 3 uL Loading Dye
  - o Run at 120V for about 30 min.