D. vulgaricate

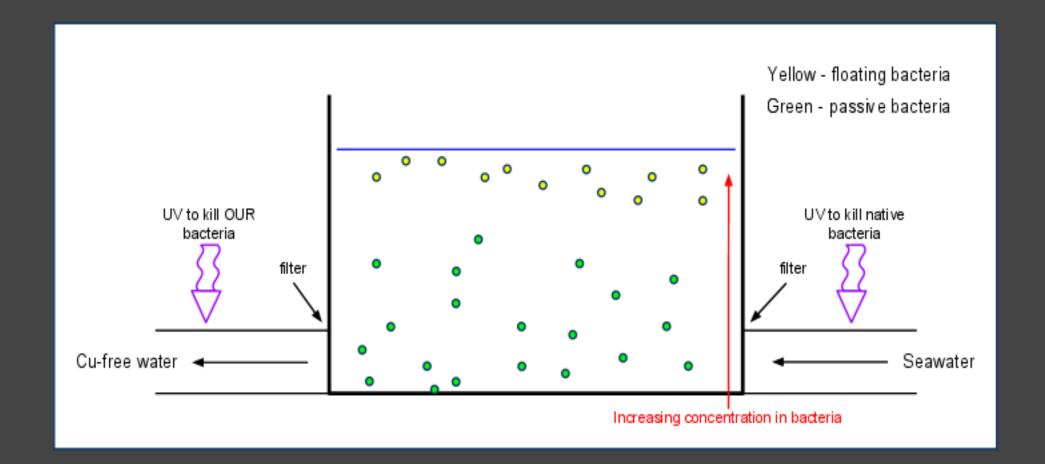
Will Ben

Alvin Jose

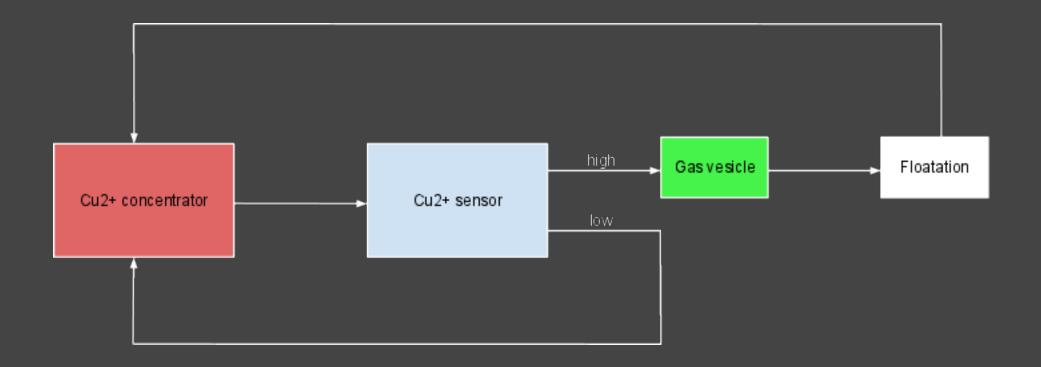
Changes

- Decided to import tunicate abilities to bacteria
- Previous research done on importing to E. coli, but this is not a useful chassis (salt water)
- Cu²⁺ as new target ion
- Shift to systems engineering and feasibility
- Goal: design a constant-throughput system for concentrating metals

Design



Devices



Devices

 Cu²⁺ concentrator: entire point of the project - continually binds Cu²⁺ from the seawater and brings it into the cell

 Cu²⁺ sensor: detects levels of Cu²⁺ in the cell, allows properly-timed flotation

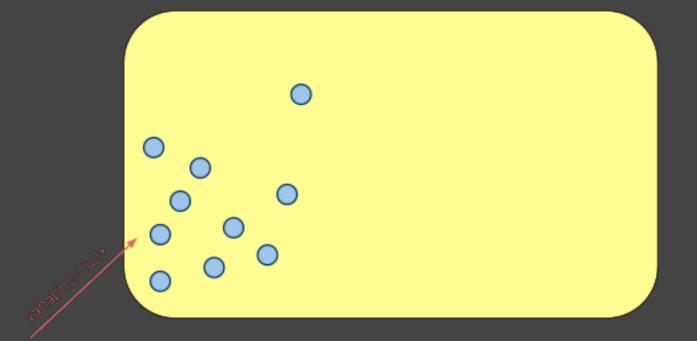
 Flotation device (gas vésicle generator): triggered by Cu²⁺ sensor - brings Cu-rich bacteria to surface for harvesting

Chassis - Desulfovibrio vulgaris

- Salt-tolerant
- Sequenced genome
- Resistant to very high osmotic stress
- Survivable in environments with a variety of heavy metals.

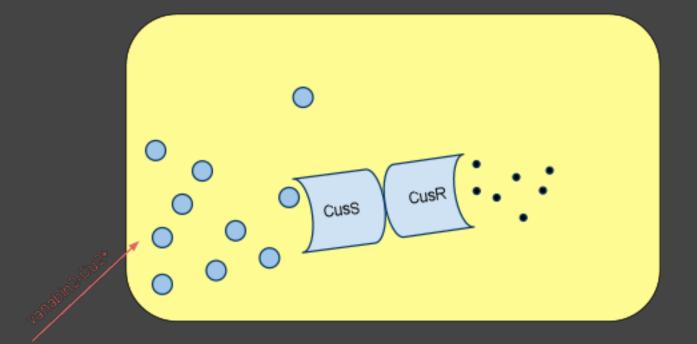
Cu²⁺ binder - MBP-vanabin2

- Vanabin2 protein binds vanadium in tunicate
- Successfully transported to E. coli and fused to MBP (maltose binding protein
- In E. coli, binds Cu²⁺ to concentration of 882ng/mg



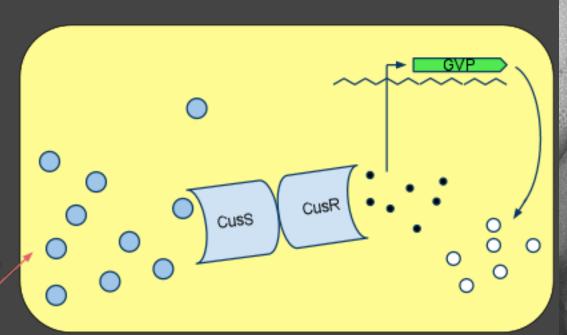
Cu²⁺ sensor - CusR/CusS two component signal system

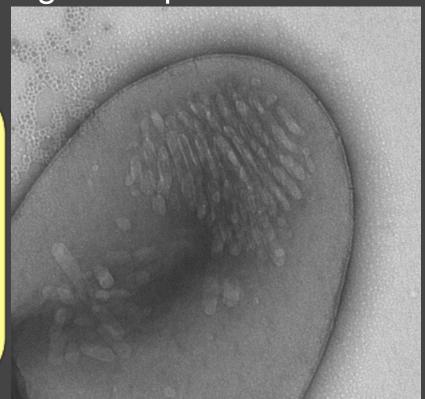
- Originally found in E. coli as a transmembrane sensor
- CusS phosphorylates CusR, which then is an activator
- Hope to bring CusS inside cell, fuse to CusR



Flotation - Gas Vesicle polycistonic gene

- Creates gas-filled organelles inside the cell
- Buoyancy of cell increased, resulting in flotation
- Previously activated by an arsenic-regulated promoter

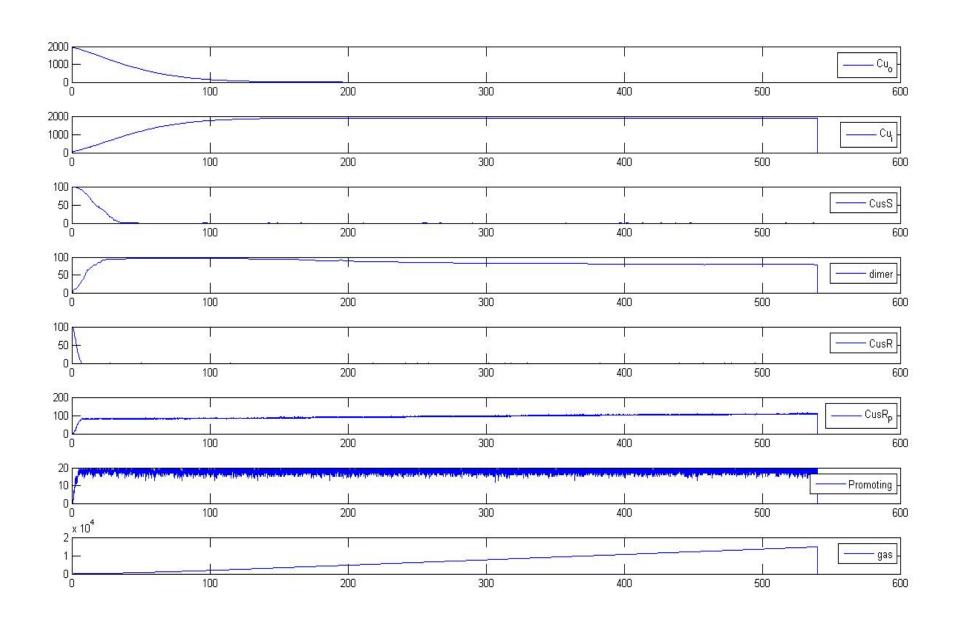




Equations:

```
1) Cu^{2+} + Vanabin2 = Cu Complex
2) Cu Complex is brought inside cell
3) Cu Complex = Cu^{2+} + Vanabin2
4) Vanabin2 is exported
5) Cu^{2+} + CusS = CusS Cu
6) CusS Cu + CusS Cu = CusS Cu 2
7) CusS Cu 2 + CusR = CusR P + CusS Cu 2
8) CusR P binds to GVP Promoter
9) Promoter then increases expression of
GVP
```

Timing Diagram



Error-Checking and Testing

- Implement one part at a time!
- test components in *D. vulgaris* individually first.
- 1. Insert gene(s) for Cu²⁺ concentrator.
 - 1. If cells die, the Cu overload is probably killing them, therefore new chassis.
 - 2. if no death occurs, preform assay to determine amount of Cu²⁺ in cells
- 2. Insert normal genes for signaling pathway, attach promoter to GFP or LacZ
 - 1. if death occurs, try again
 - 2. if no death occurs, culture on X-Gal or check for GFP to see if it functions
 - 3. if it does not function, assay for CusS and CusR to see what is wrong

Error Checking and Testing

- 3. place GVP genes into chassis with promoter known to work
 - 1. if cell dies, find new chassis
 - 2. if cell lives, check if it floats or not
- 3. if it doesn't float, assay for proteins to see if GVP is being made
- 4. Hook up concentrator and normal signaling pathway to check for denaturing of proteins
- 5. Hook up concentrator, normal pathway and GVP, determine if you have expression
- 6. Start mutagenizing CusS until you have one that works inside the cell membrane

Impact

- Proof of concept using copper
- Hope to tune the system for other metals.
- Not environmentally destructive
- Tremendous resources in ocean
- Mesh with current lithium extraction techniques



Outstanding Issues

Internalizing Cu²⁺ two-component system

- Current state, is transmembrane
- Unknown if removing CusS from membrane, fusing to CusR is feasible
- Possibility of crosstalk between sensor and native features in cell
- Alternative: batch processing of seawater, measure concentration drop outside cell

Outstanding Issues

Implementation

- Vanabins untried in our chassis
- D. vulgaris has complex interactions with many ions which give it its durability
- Promoting GVP with CusR
- Sufficiency of GVP to overcome random turbulence

GO