

Dissecting Complex Trait Variation with the *Drosophila* Synthetic Population Resource

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Motivation

Types of causative variant

Coding or regulatory ?
SNPs, simple InDels,
CNVs ?

Mechanism

Molecular / cellular processes by which natural allelic variation leads to phenotypic change

Evolution

What forces maintain genetic variation ?

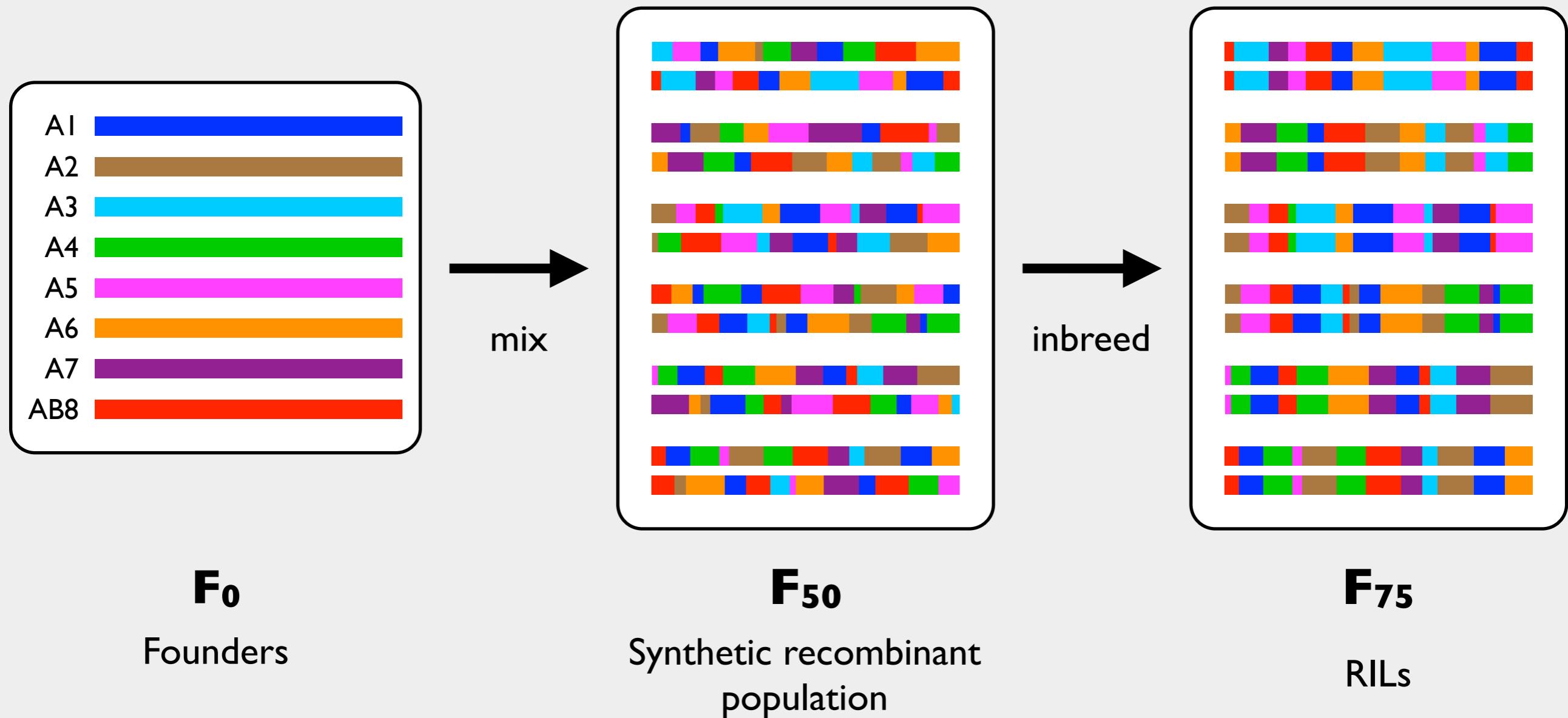
Mutation-Selection Balance

Variants unconditionally deleterious
Very rare (< 1% MAF)
Large effect

Balancing Selection

Variants maintained by selection
Intermediate-frequency
Subtle effect

Drosophila Synthetic Population Resource



Empirical “Positive Control”

Overall activity of Alcohol Dehydrogenase (ADH) enzyme

Quantitative genetic variation of enzyme activities in natural populations of *Drosophila melanogaster*

(population genetics/modifier loci/regulatory elements/protein polymorphism)

C. C. LAURIE-AHLBERG[†], G. MARONI[‡], G. C. BEWLEY[†], J. C. LUCCHESI[‡], AND B. S. WEIR[§]

Quantitative analysis of RNA produced by Slow and Fast alleles of *Adh* in *Drosophila melanogaster*

(gene regulation/alcohol dehydrogenase/molecular evolution/polymorphism)

CATHY C. LAURIE* AND LYNN F. STAM*

Use of *in Vitro* Mutagenesis to Analyze the Molecular Basis of the Difference in *Adh* Expression Associated With the Allozyme Polymorphism in *Drosophila melanogaster*

Madhusudan Choudhary¹ and Cathy C. Laurie²

The Effect of an Intronic Polymorphism on Alcohol Dehydrogenase Expression in *Drosophila melanogaster*

Cathy C. Laurie and Lynn F. Stam

Molecular Dissection of a Major Gene Effect on a Quantitative Trait: The Level of Alcohol Dehydrogenase Expression in *Drosophila melanogaster*

Lynn F. Stam and Cathy C. Laurie

Classic studies on the quantitative genetics of ADH expression (Cathy Laurie)

Adh Locus

Allozyme Polymorphism

Fast AAG Lys
Slow ACG Thr

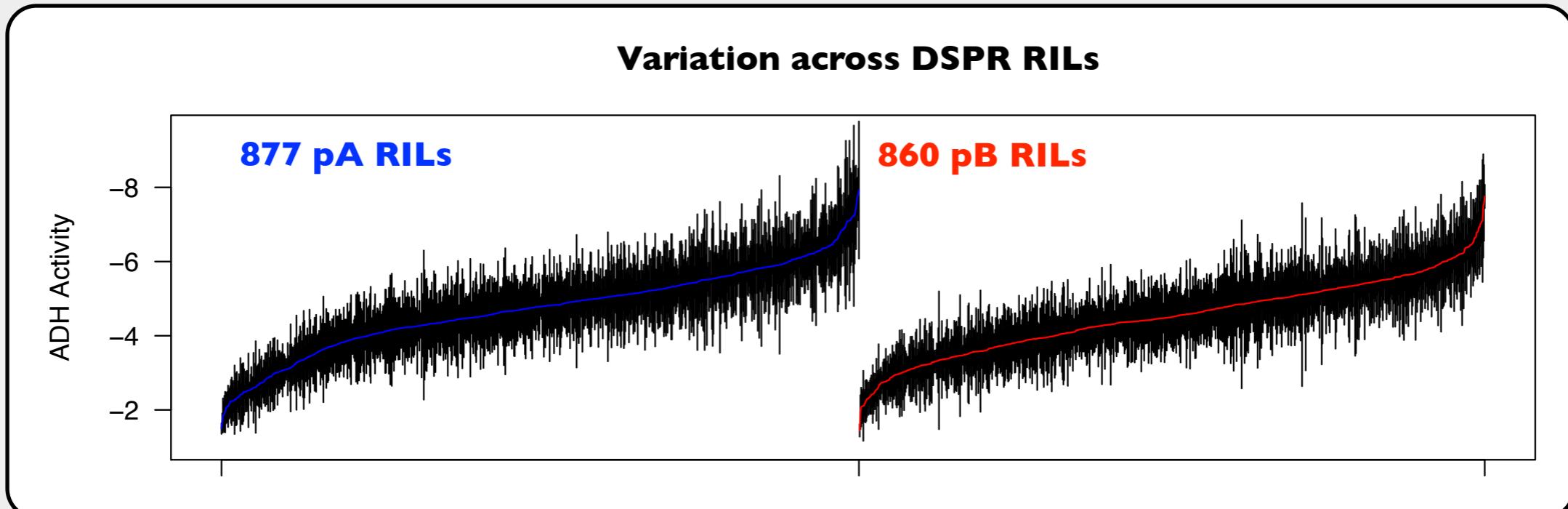
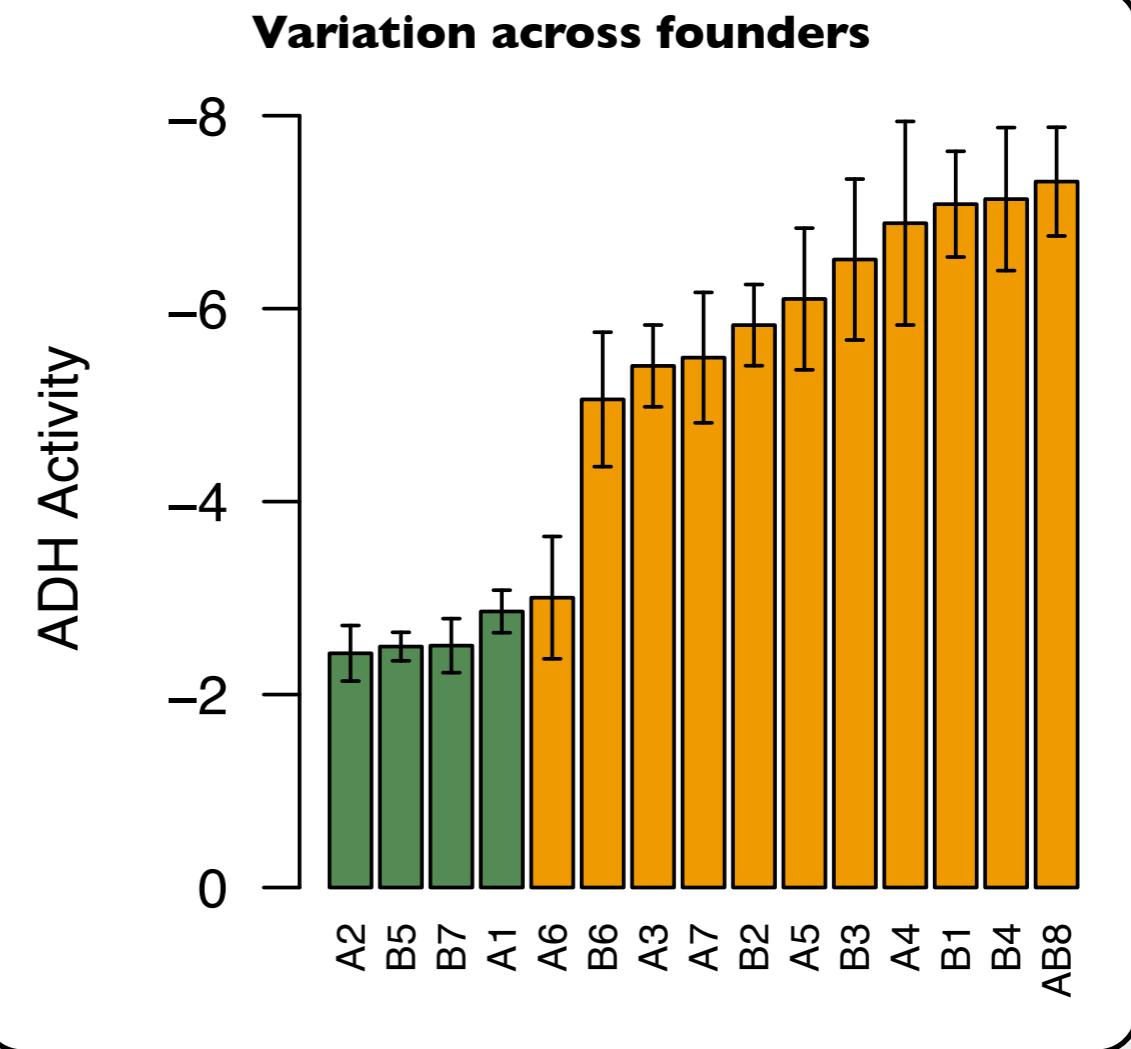


Intronic InDel Polymorphism

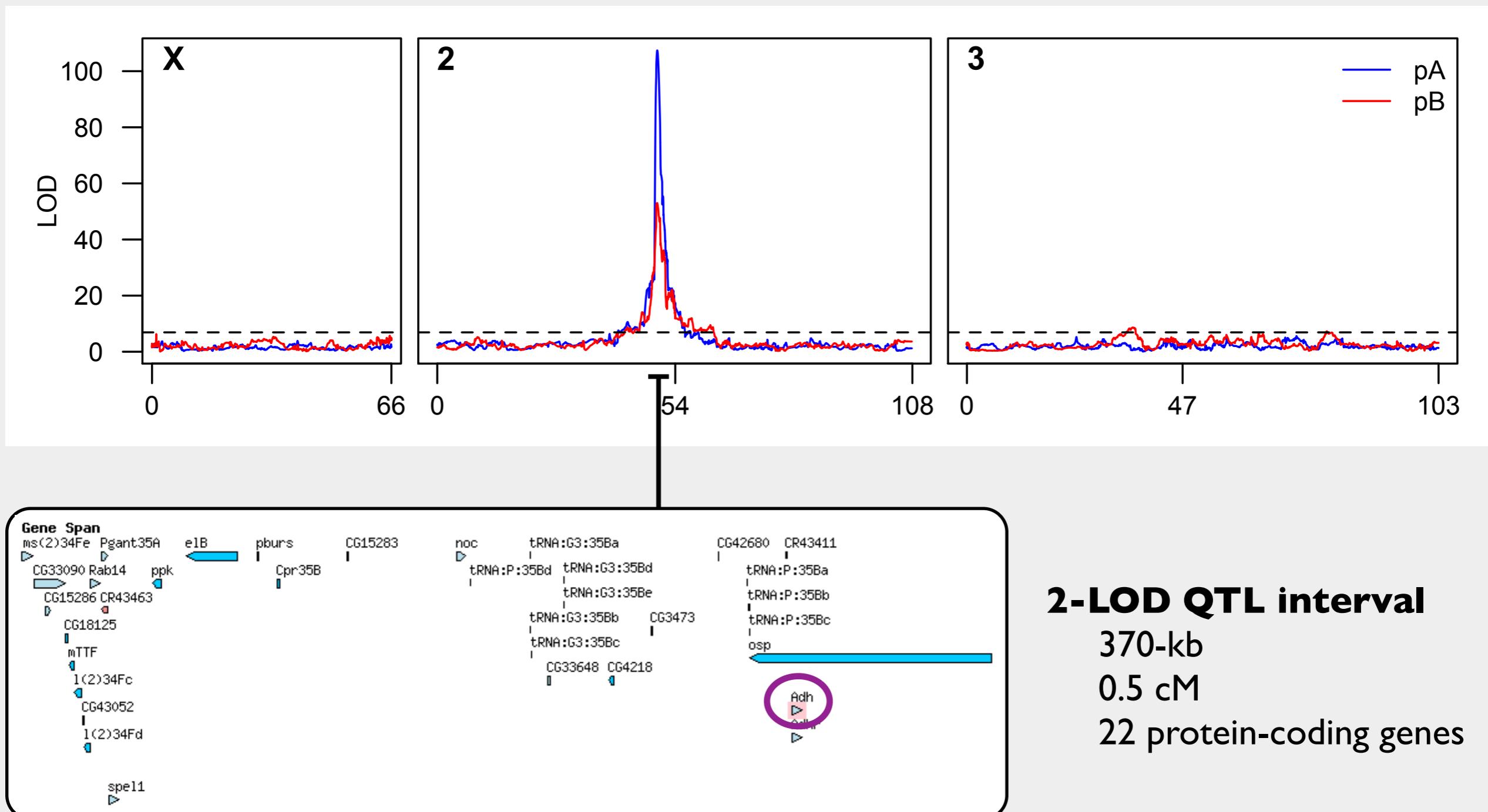
▽I-Low 29-bp
▽I-High 34-bp

ADH Activity Variation

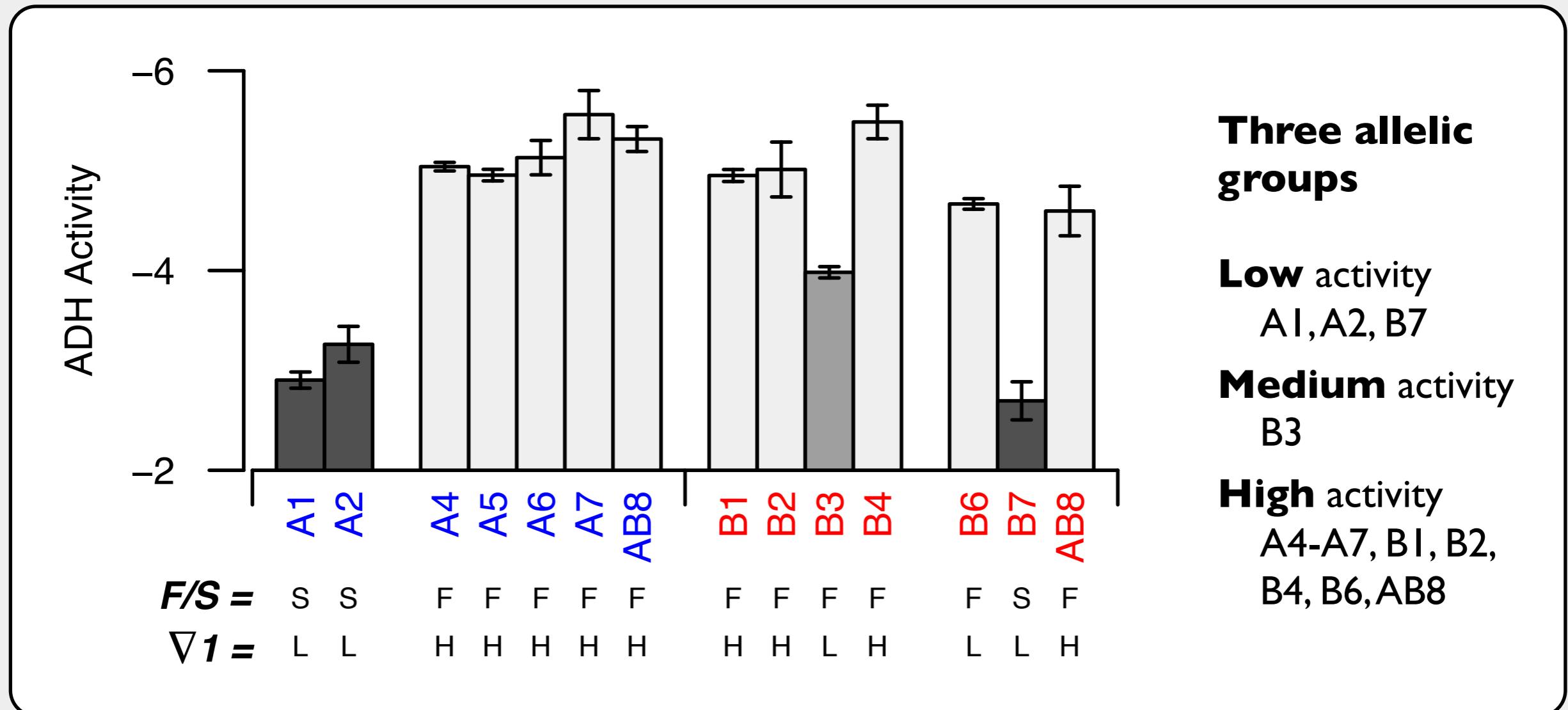
- **Slow** founders show consistently low ADH activity
- Variation among **Fast** founders - implies other loci involved
- Extensive phenotypic variation in DSPR



ADH Enzyme Activity QTL



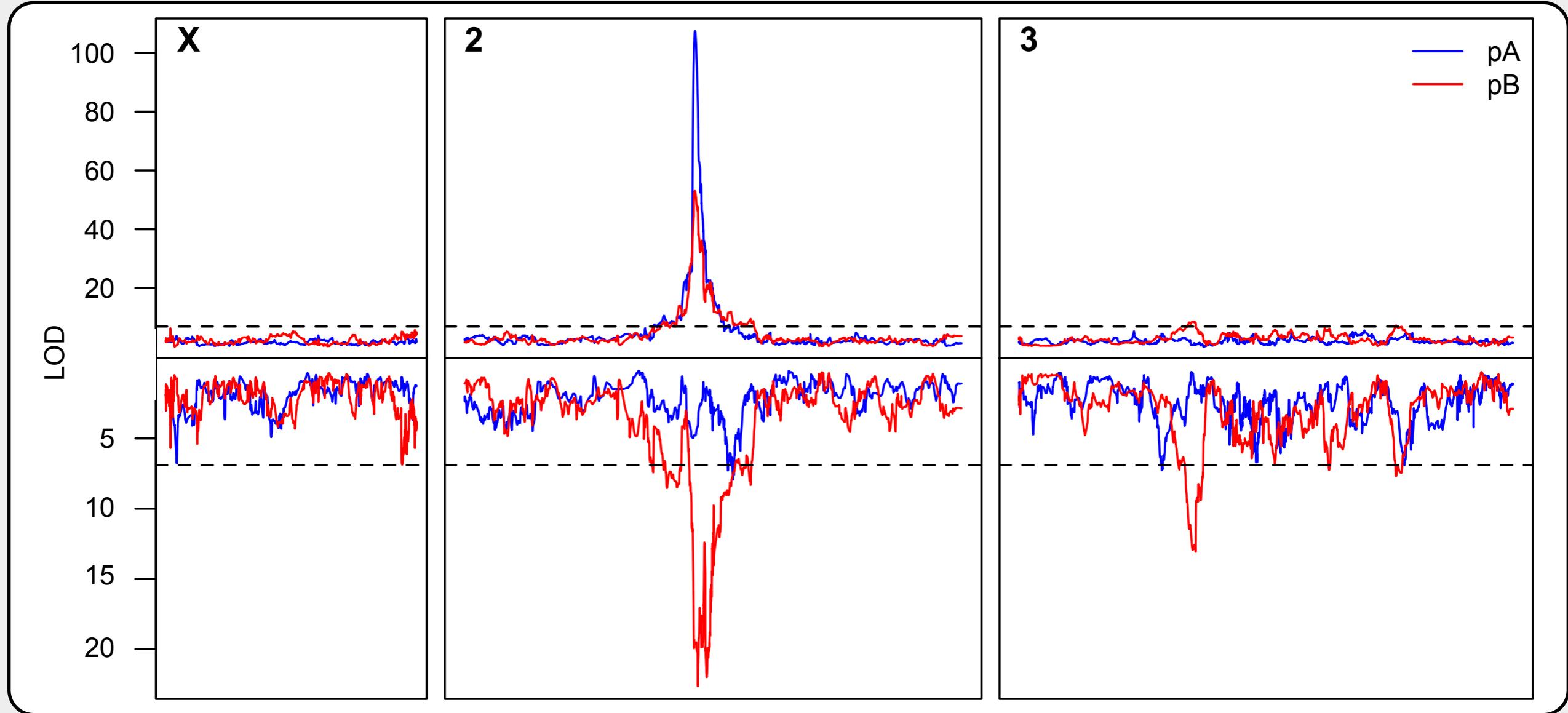
Adh QTL Phasing



Confirm effect of *Fast/Slow*, but other alleles at *Adh* are also involved

Additional ADH QTL

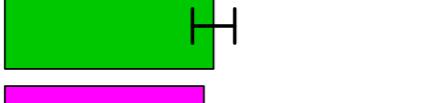
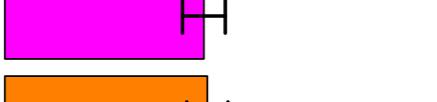
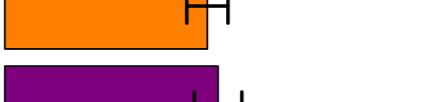
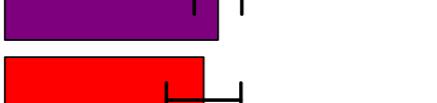
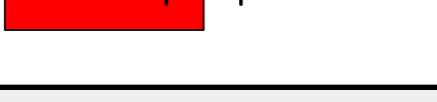
Raw ADH activity QTL



Control for *Adh* haplotypes defined by *F/S* and ∇I



QTL to Causative Variants

Founder means at QTL	Predicted allelic configuration	Sequence alignment within QTL interval
	H	GGCCAG T AAAAAATTAAATT C ACACT
	H	GGCCAG T AAAAAACATTAAATT C ACACT
	H	GGCCAG T AAAAAATTAAATT C ACAGT
	L	GACCAG C AAAAAATTAAATT T ACAGT
	L	GACCAG C AAAAACATTAAATT T ACAGT
	L	GGCCAG C AAAAAATTAAATT T ACACT
	L	GGCCAG C AAAAACATTAAATT T ACACT
	L	GGCCAG C AAAAAATTAAATT T ACAGT

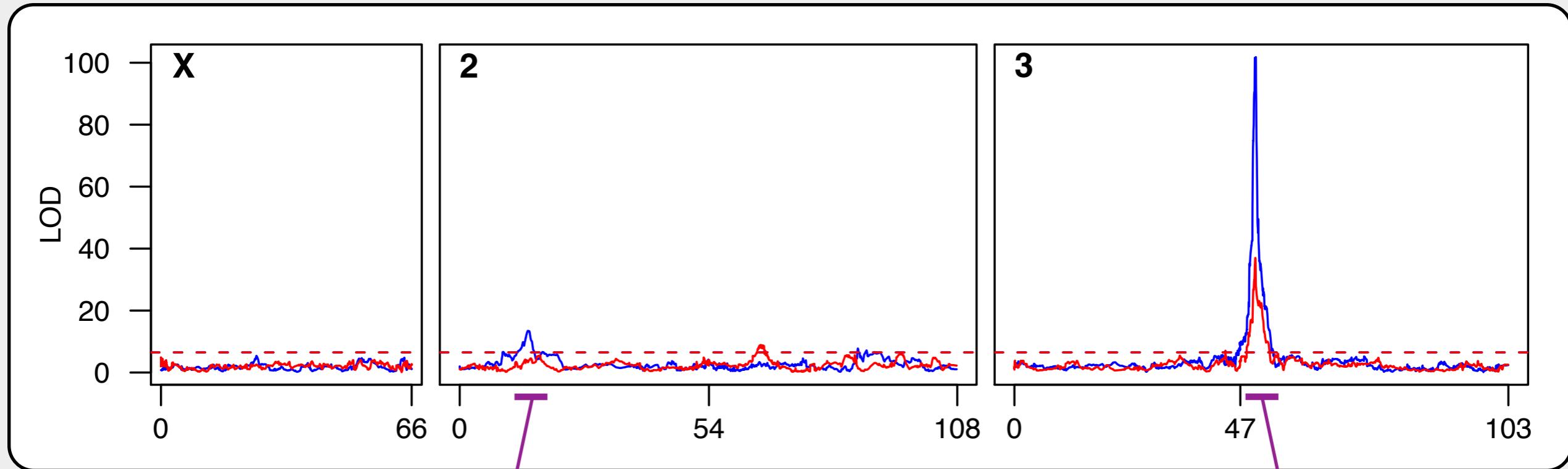
Few “in-phase” Polymorphisms

	SNP	nsSNP	InDel
QI	169	3	48
QR2	6	0	8
QR3	102	25	89
QR4	12	1	12
QR6	537	233	490
QR7	2	6	11
QR8	37	2	33

QI
Includes known
F/S Adh variant

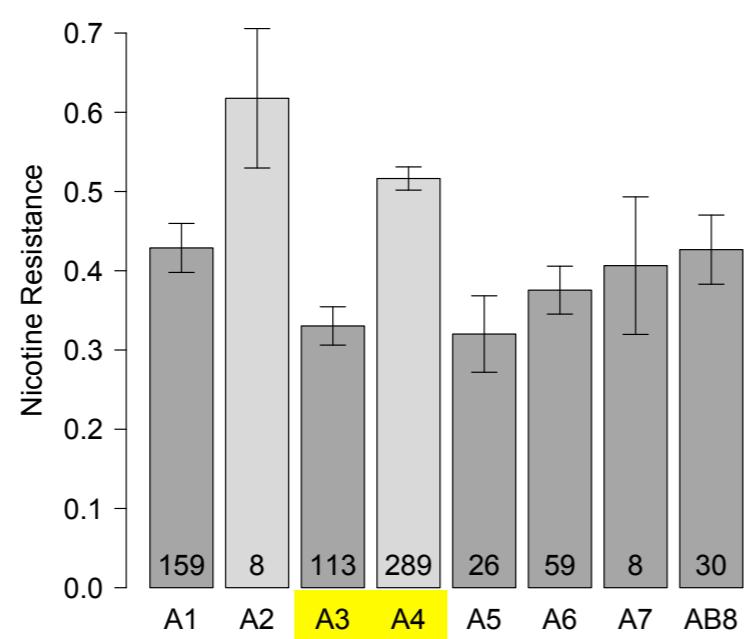
QR2
Includes frameshift
mutation in CG7377
and intronic deletion
in CG6024

Nicotine Resistance QTL



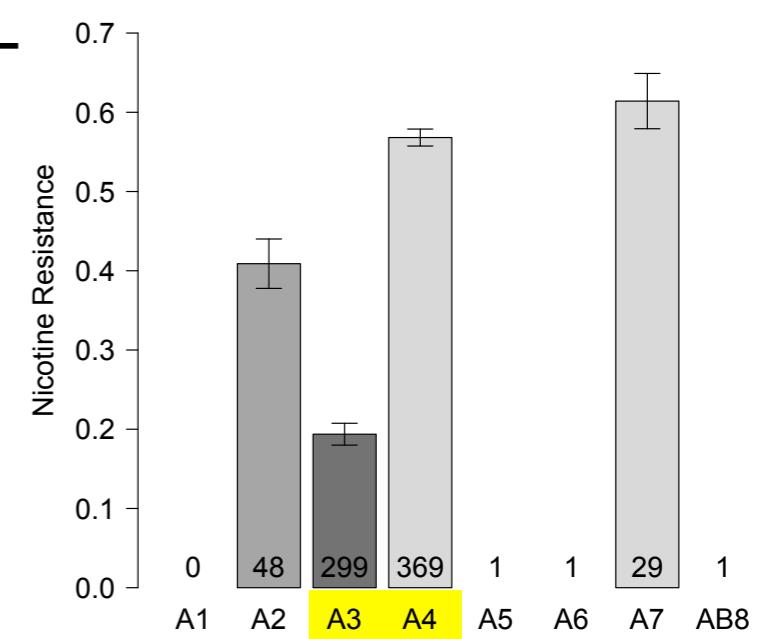
Biallelic QTL
14.4% variation

220-kb
0.84 cM
27 genes

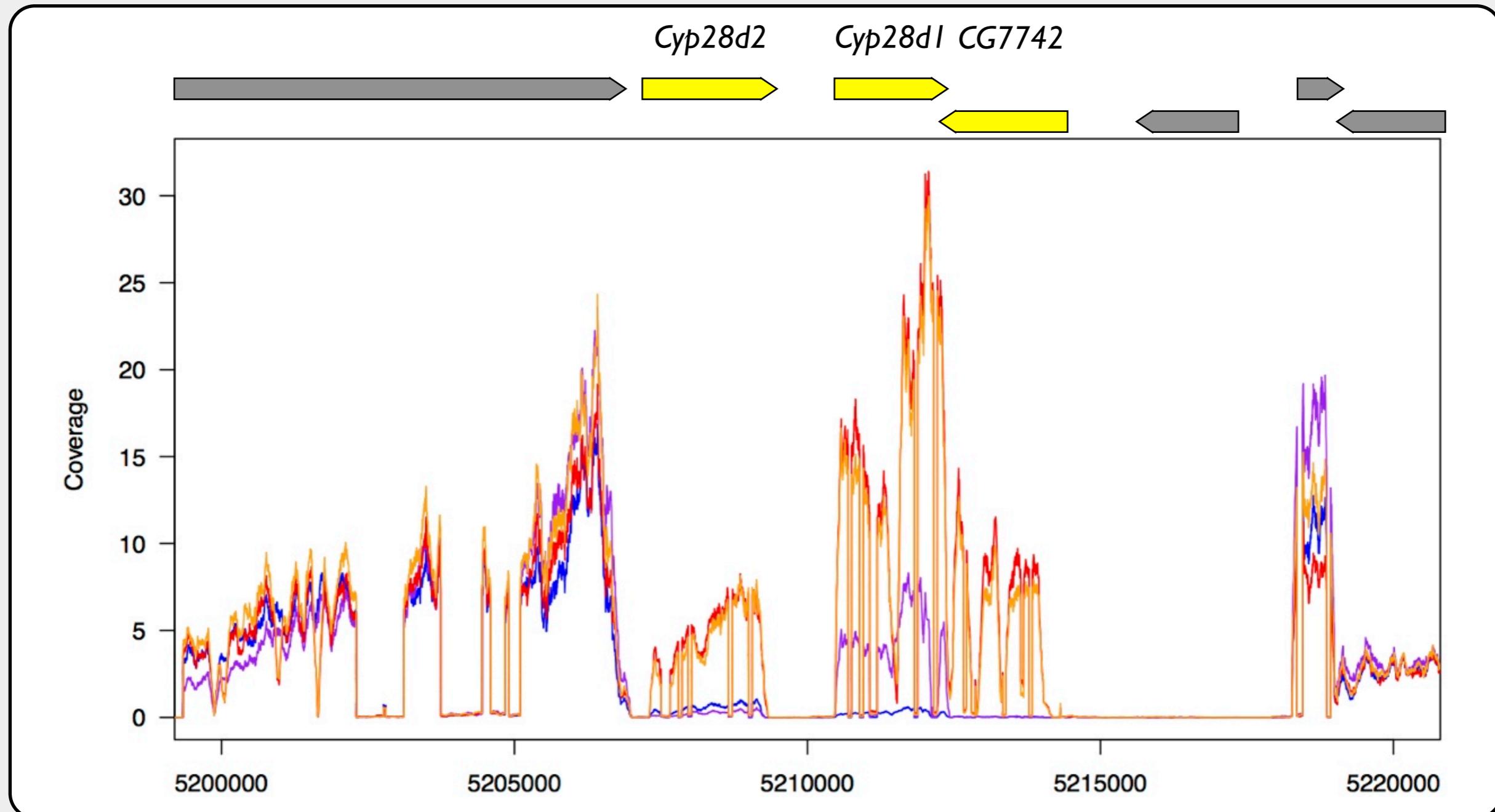


Multiallelic QTL
51.5% variation

190-kb
0.21 cM
32 genes



Expression Candidates (2L)



A3 (regular)



A3 (nicotine)

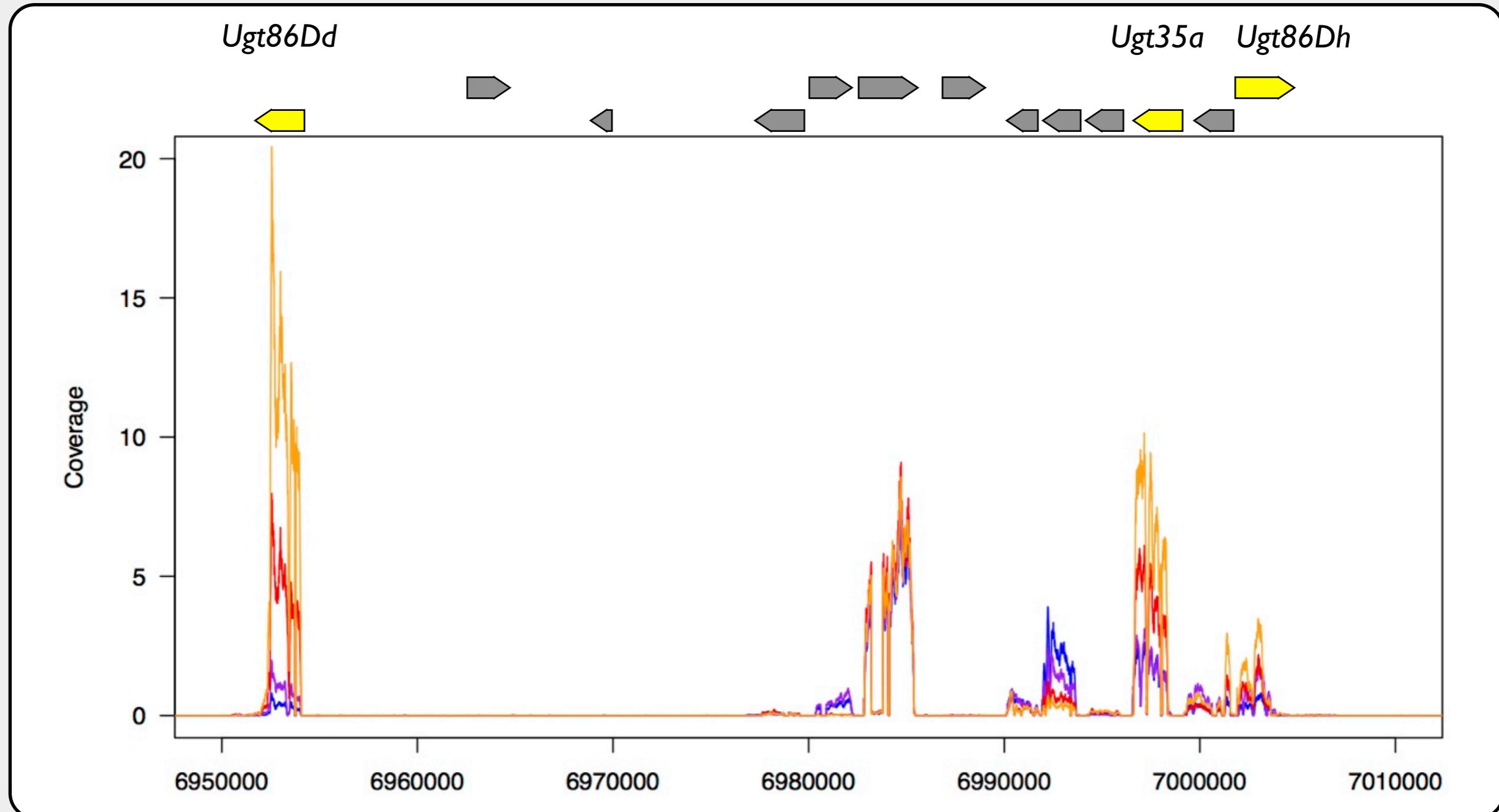


A4 (regular)



A4 (nicotine)

Expression Candidates (3R)



A3 (regular)



A3 (nicotine)

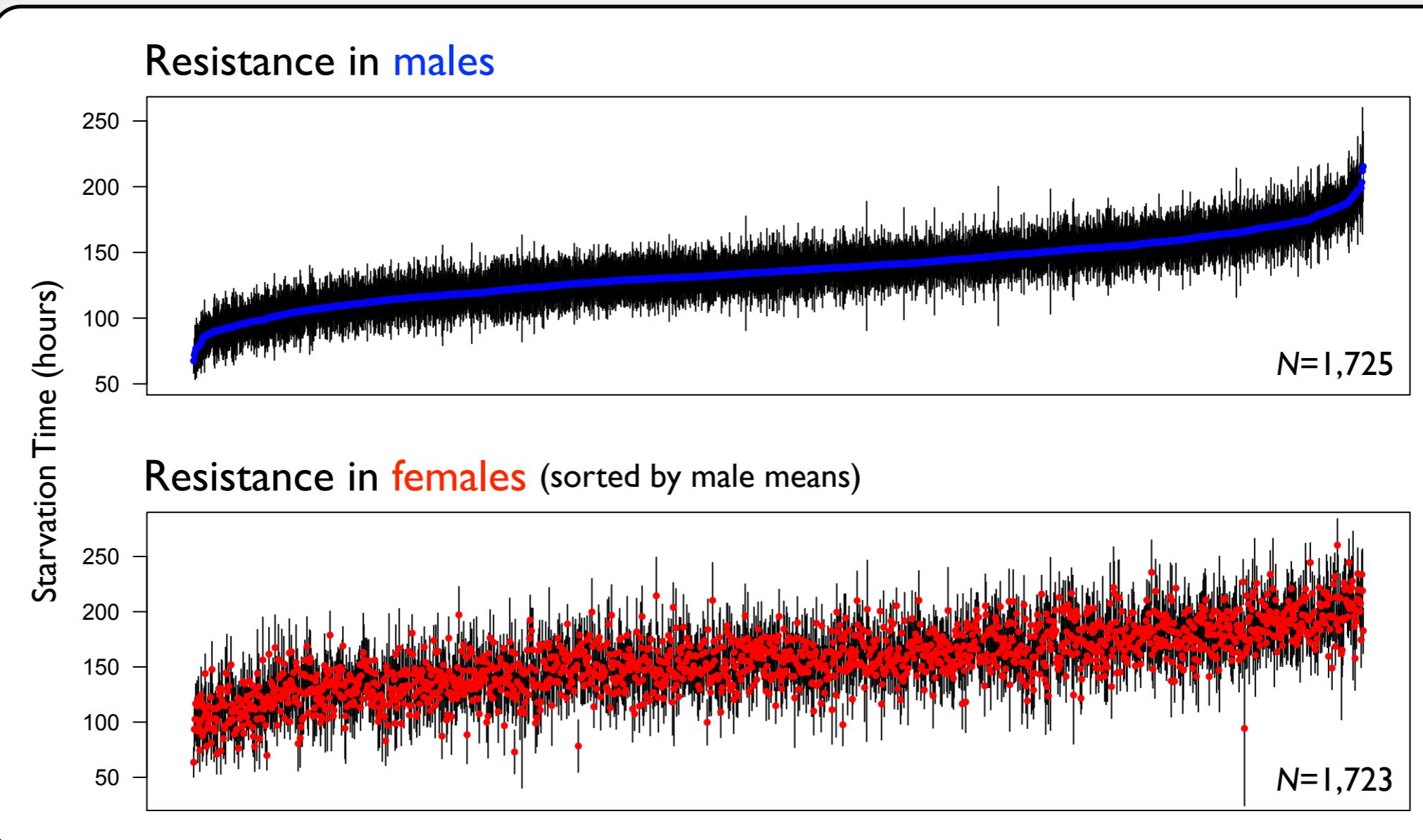


A4 (regular)



A4 (nicotine)

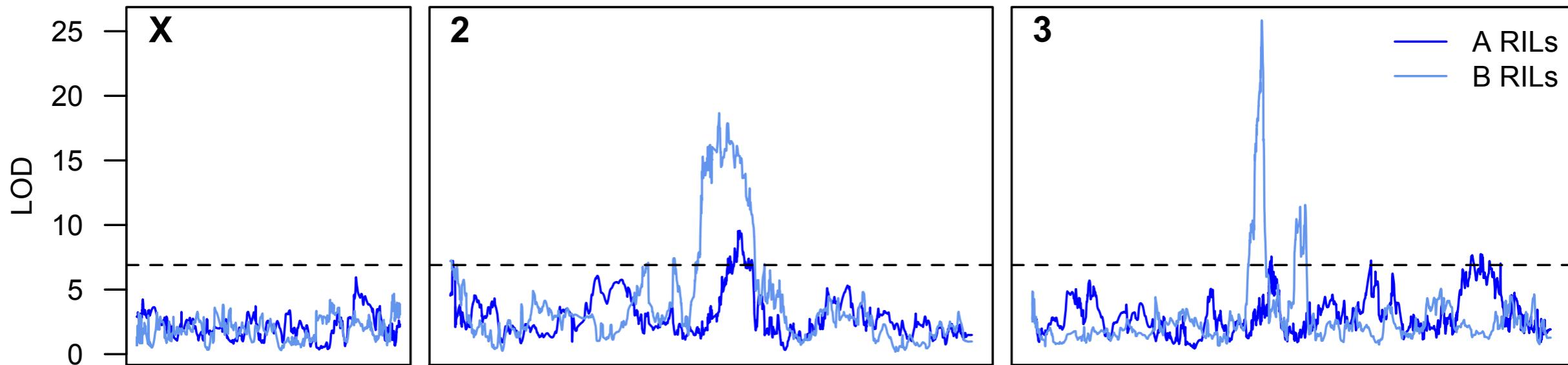
Starvation Resistance in DSPR



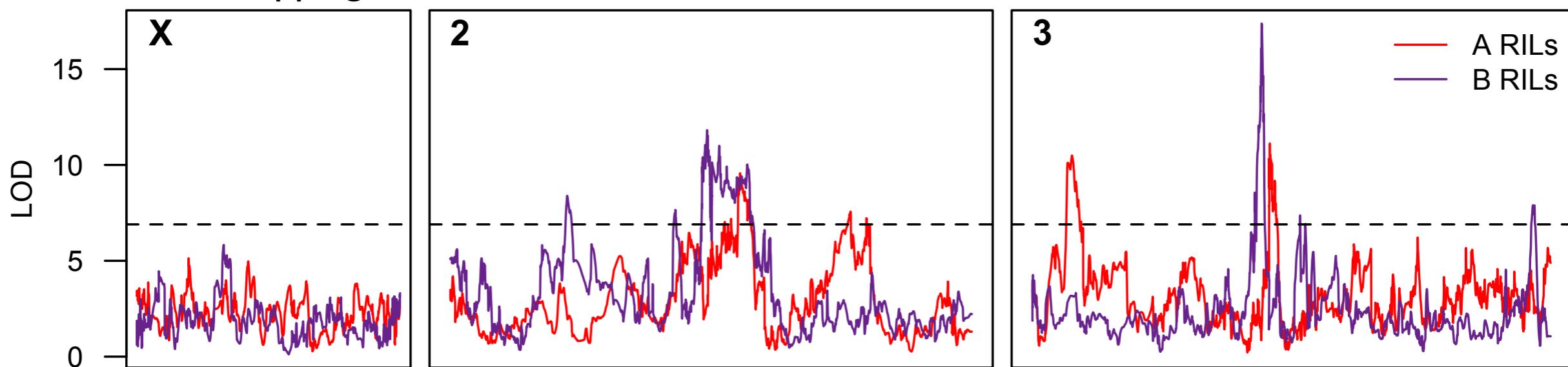
- Heritability, $H^2 = 52-55\%$

Numerous Starvation QTL

QTL mapping in **males**



QTL mapping in **females**

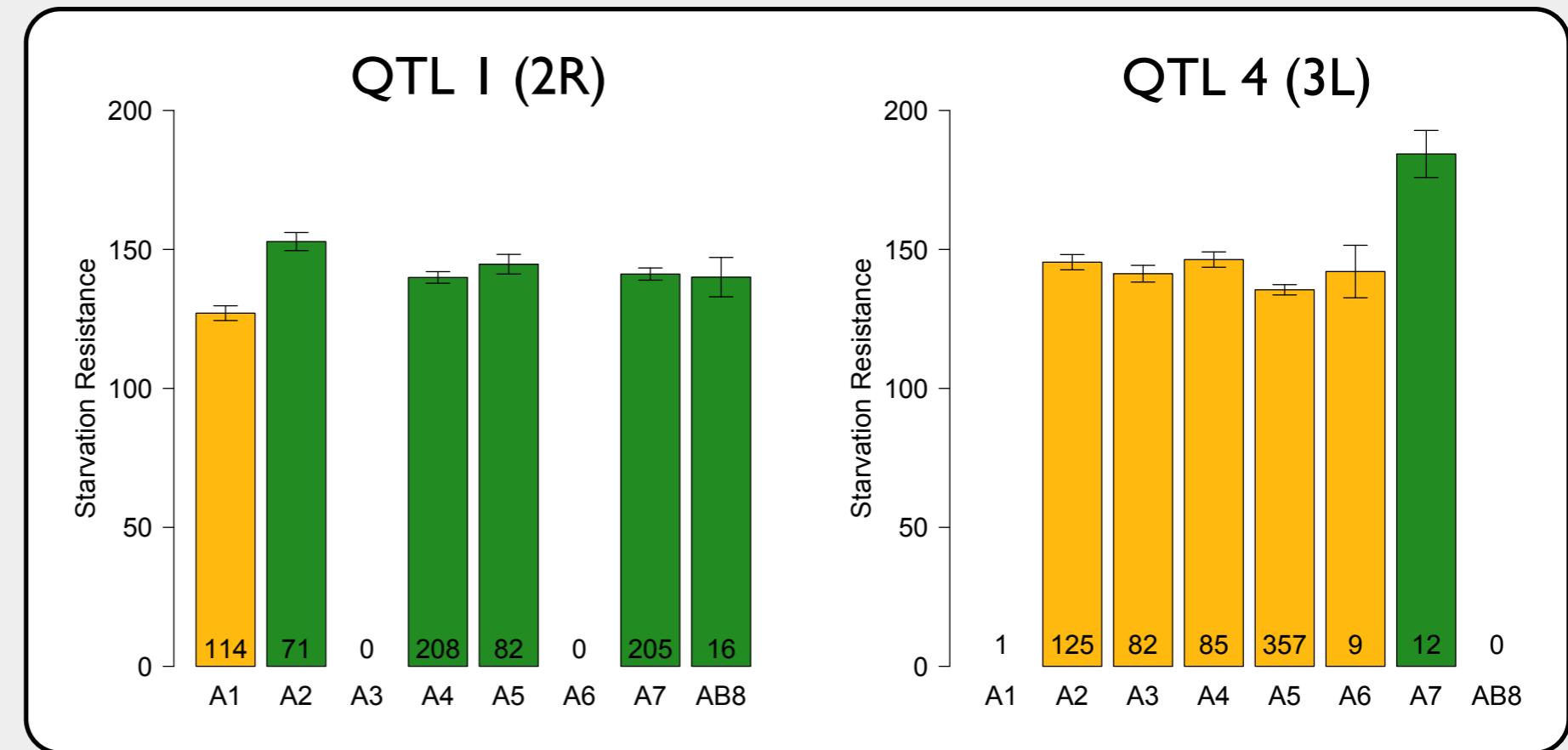


Female QTL Summary

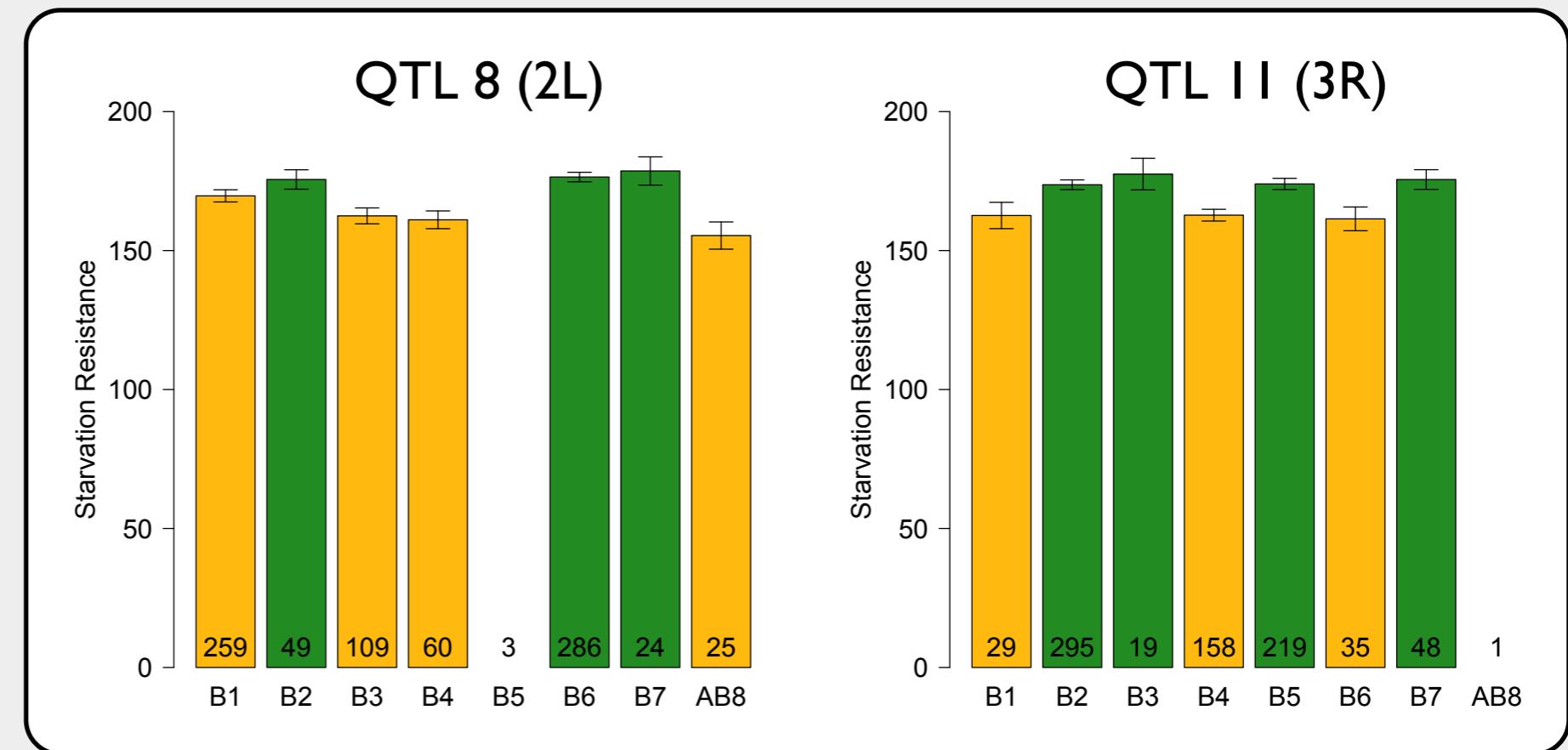
QTL	Popn	Chr	% Var Expl	Size (Mb)	# Genes
1	A	2R	5.8	1.4	220
2	A	2R	4.6	0.9	179
3	A	2R	4.4	0.6	103
4	A	3L	6.4	0.5	41
5	A	3R	6.7	1.5	208
6	B	2L	4.6	0.4	28
7	B	2L	4.3	0.5	40
8	B	2L	7.1	2.5	243
9	B	3L	10.5	0.5	43
10	B	3R	4.9	0.5	64
11	B	3R	4.4	0.5	104
			5.8 ± 1.87	0.9 ± 0.66	116 ± 81.8

Strain Effects

Rare QTL

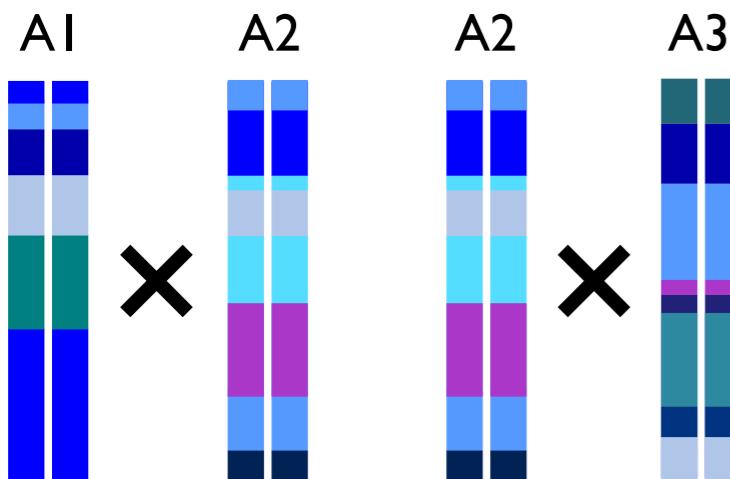


Common QTL



Heterozygous Mapping Designs

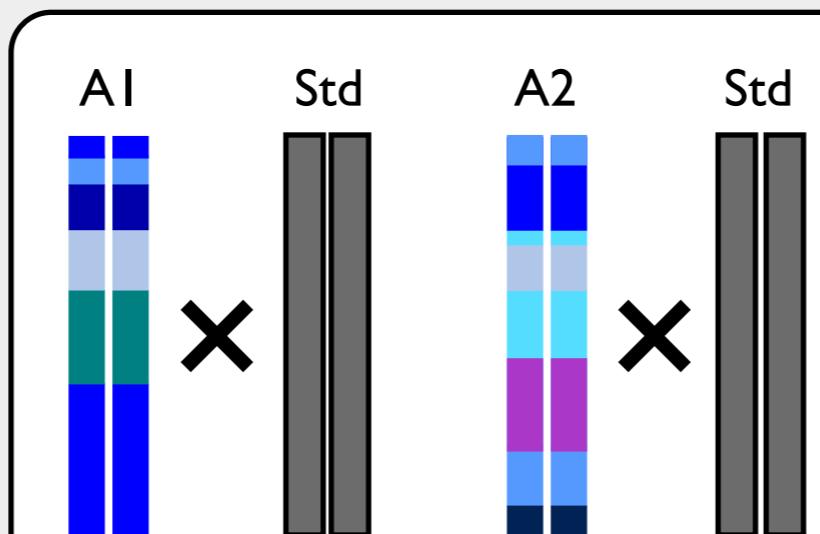
- Map using *trans*-heterozygous cross progeny
- Minimizes inbreeding depression



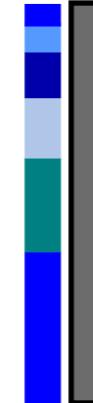
A1*2



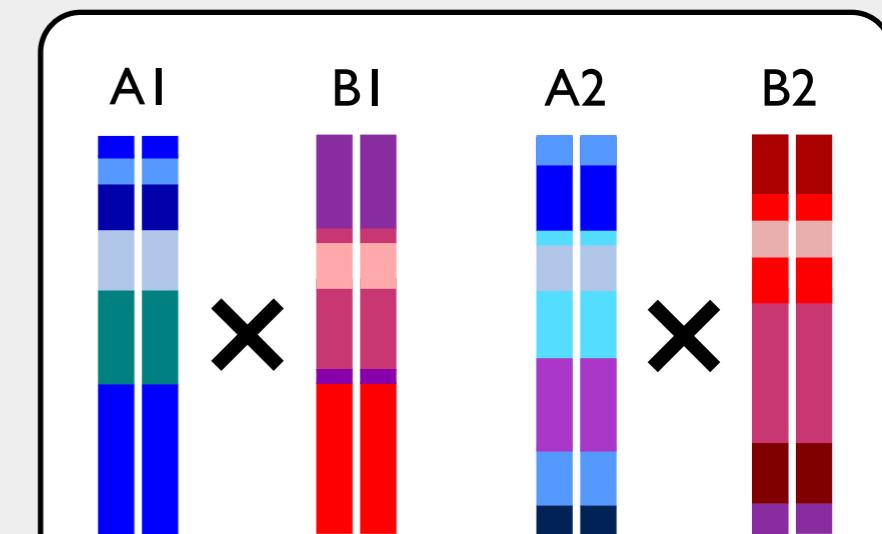
A2*3



A1*Std



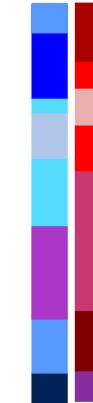
A2*Std



A1*BI



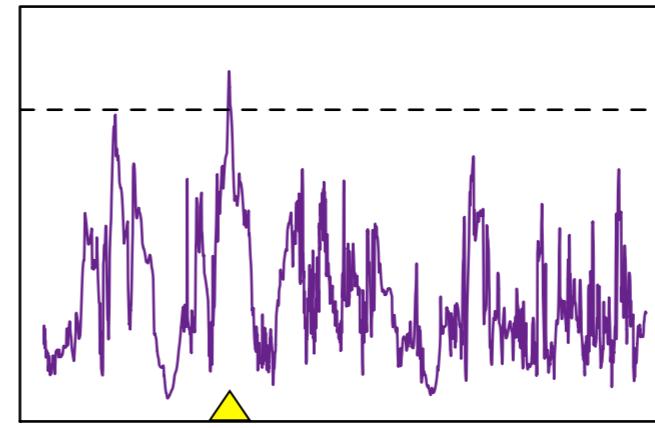
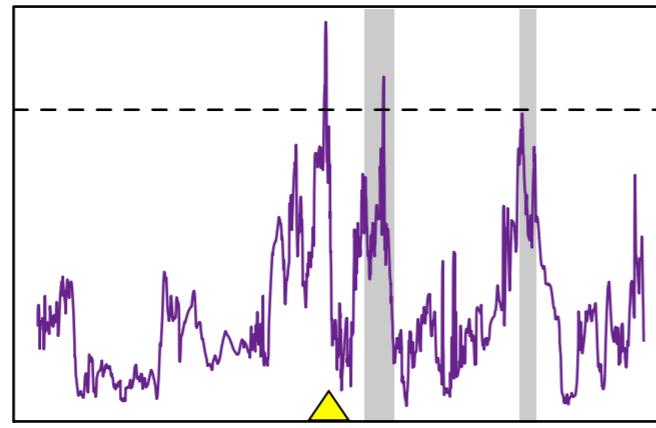
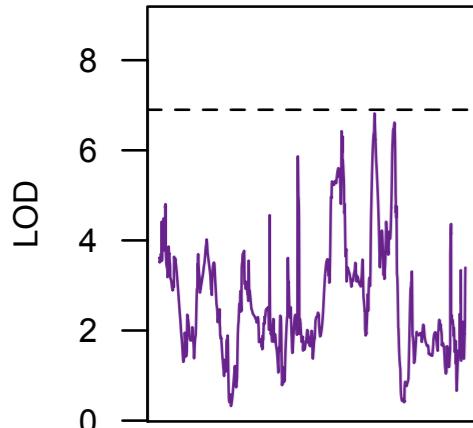
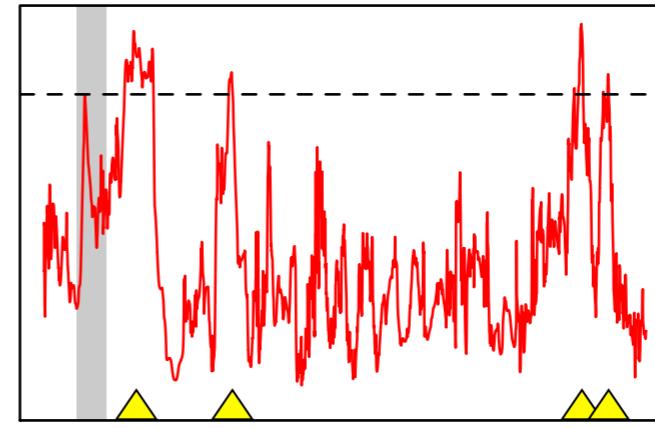
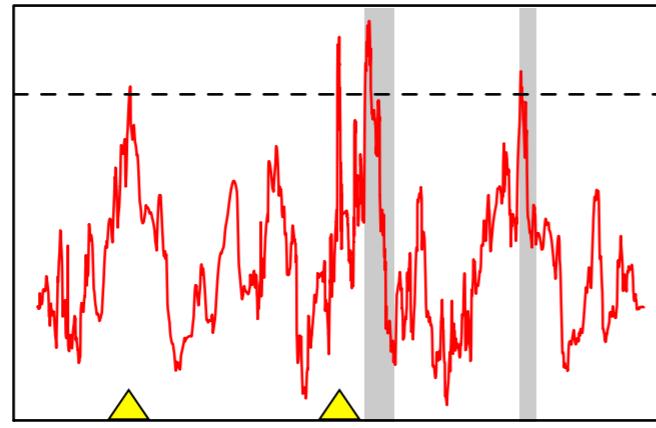
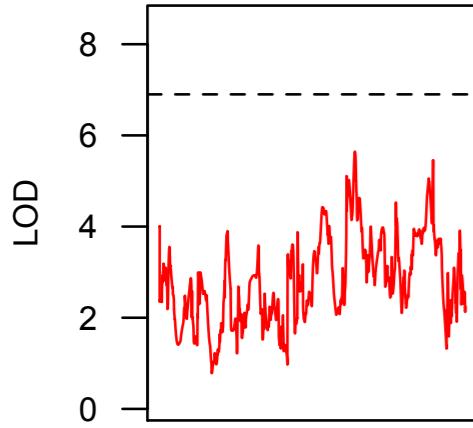
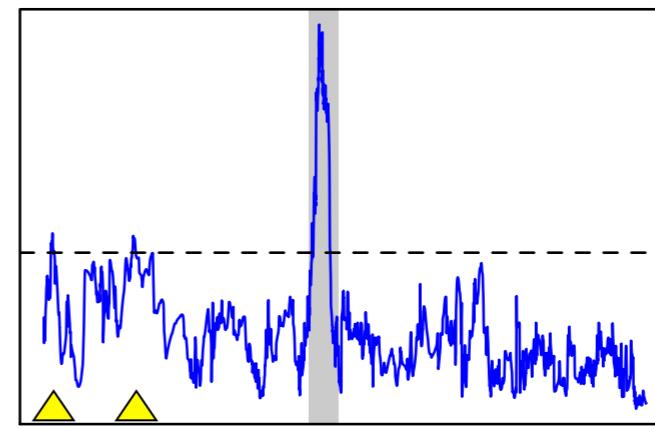
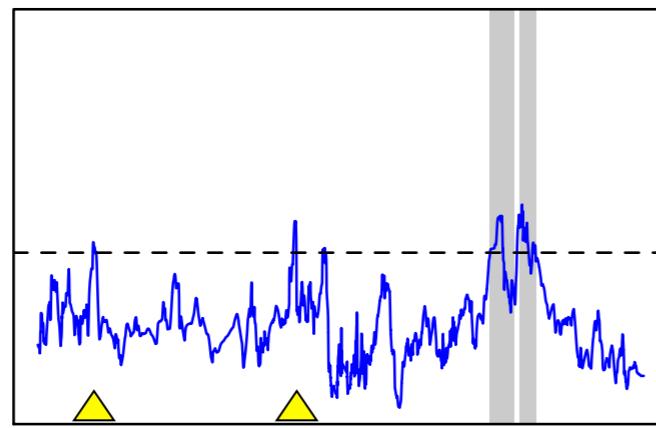
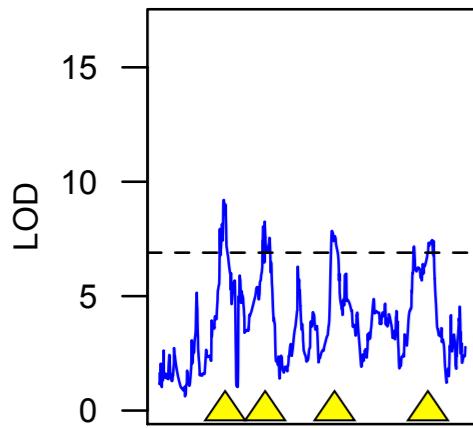
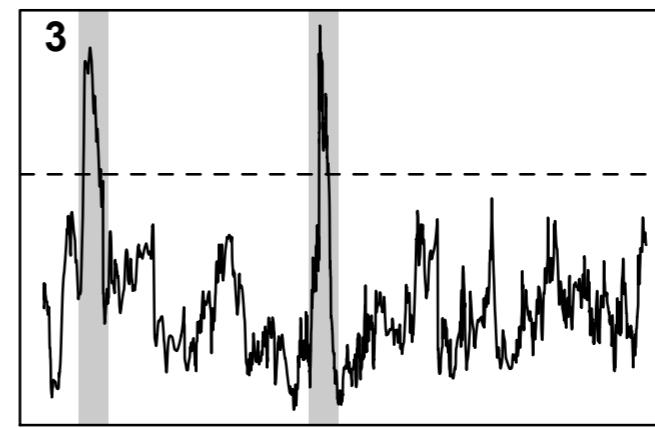
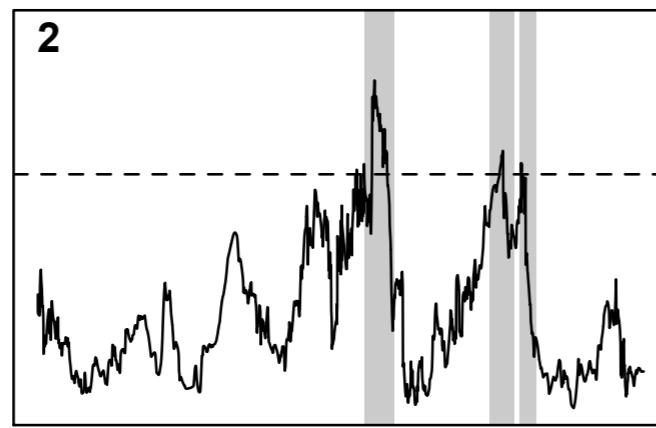
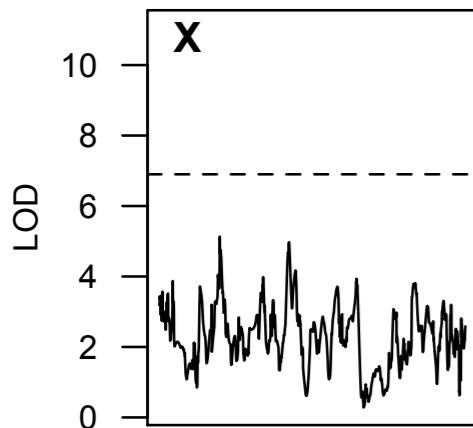
A2*B2



pA RIL × **pA** RIL
Crosses

pA RIL × Standard
Crosses

pA RIL × **pB** RIL
Crosses



Expression Profiling

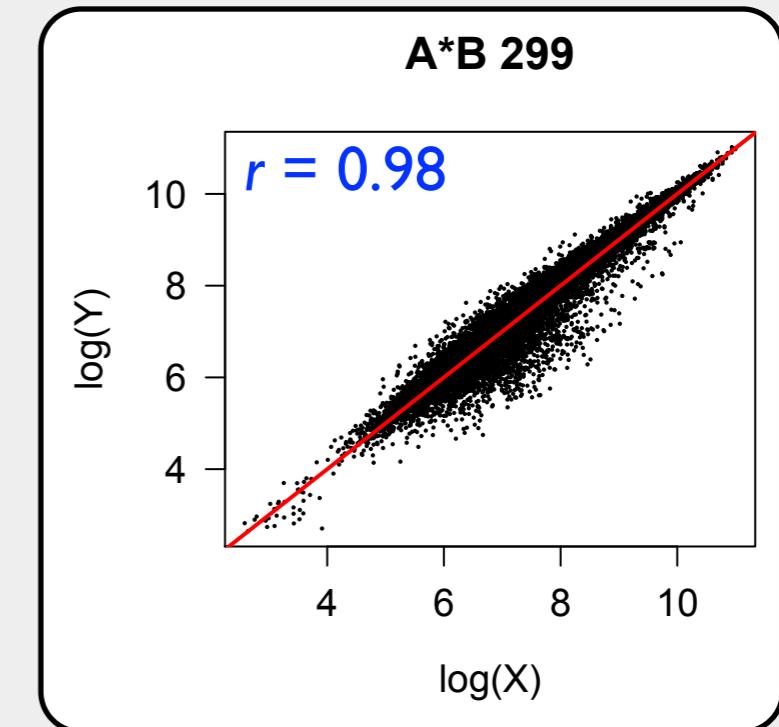
- RNA from ~300 3-5 d.o. mated female heads per genotype

- NimbleGen 12 × 135K microarrays

16,637 target genes
(8 probes/target)

- Robust multi-array (RMA) analysis

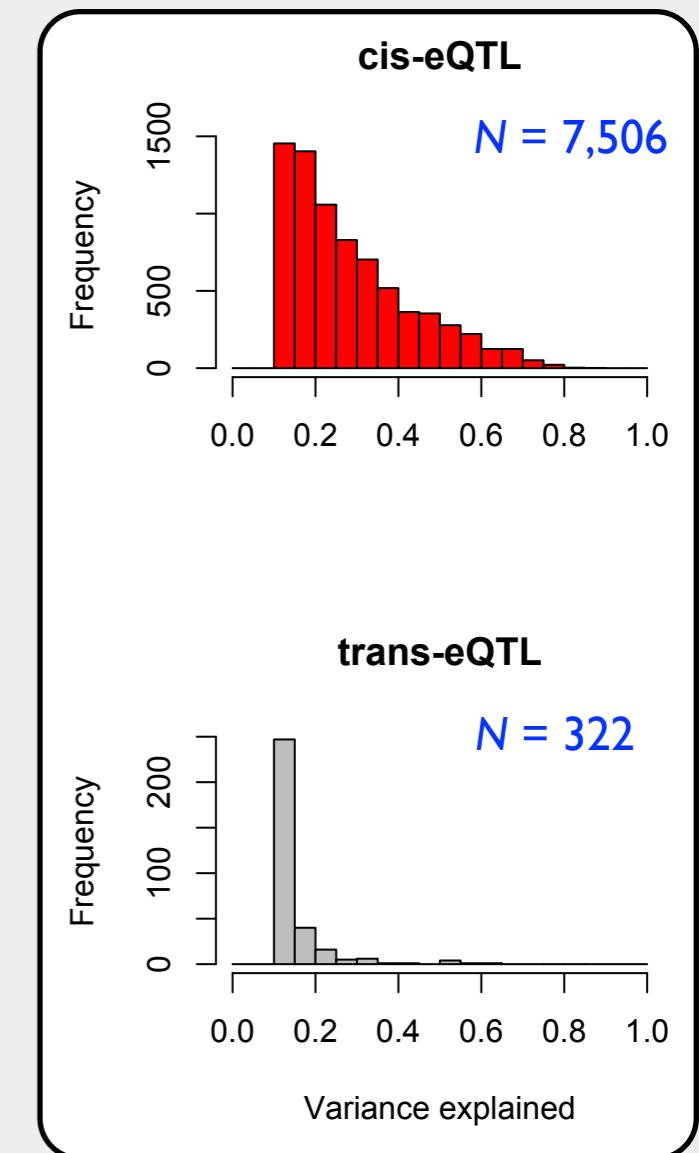
Account for mismatches in probes



Strong correlation between
replicate arrays

Mapped eQTL

Symbol	Name	FlyBase #	eQTL
<i>Clk</i>	<i>Clock</i>	FBgn0023076	<i>cis</i>
<i>tim</i>	<i>timeless</i>	FBgn0014396	<i>cis</i>
<i>rho</i>	<i>rhomboid</i>	FBgn0004635	<i>cis</i>
<i>cyc</i>	<i>cycle</i>	FBgn0023094	<i>cis</i>
<i>Oda</i>	<i>Ornithine decarboxylase antizyme</i>	FBgn0014184	<i>trans</i>
<i>Slob</i>	<i>Slowpoke binding protein</i>	FBgn0264087	<i>trans</i>
<i>DopR</i>	<i>Dopamine receptor</i>	FBgn0011582	<i>cis</i>
<i>Ssk</i>	<i>Snakeskin</i>	FBgn0036945	<i>trans</i>
<i>Pka-R2</i>	<i>cAMP-dependent protein kinase R2</i>	FBgn0022382	<i>cis</i>
<i>5-HT1B</i>	<i>Serotonin receptor 1B</i>	FBgn0263116	<i>cis</i>

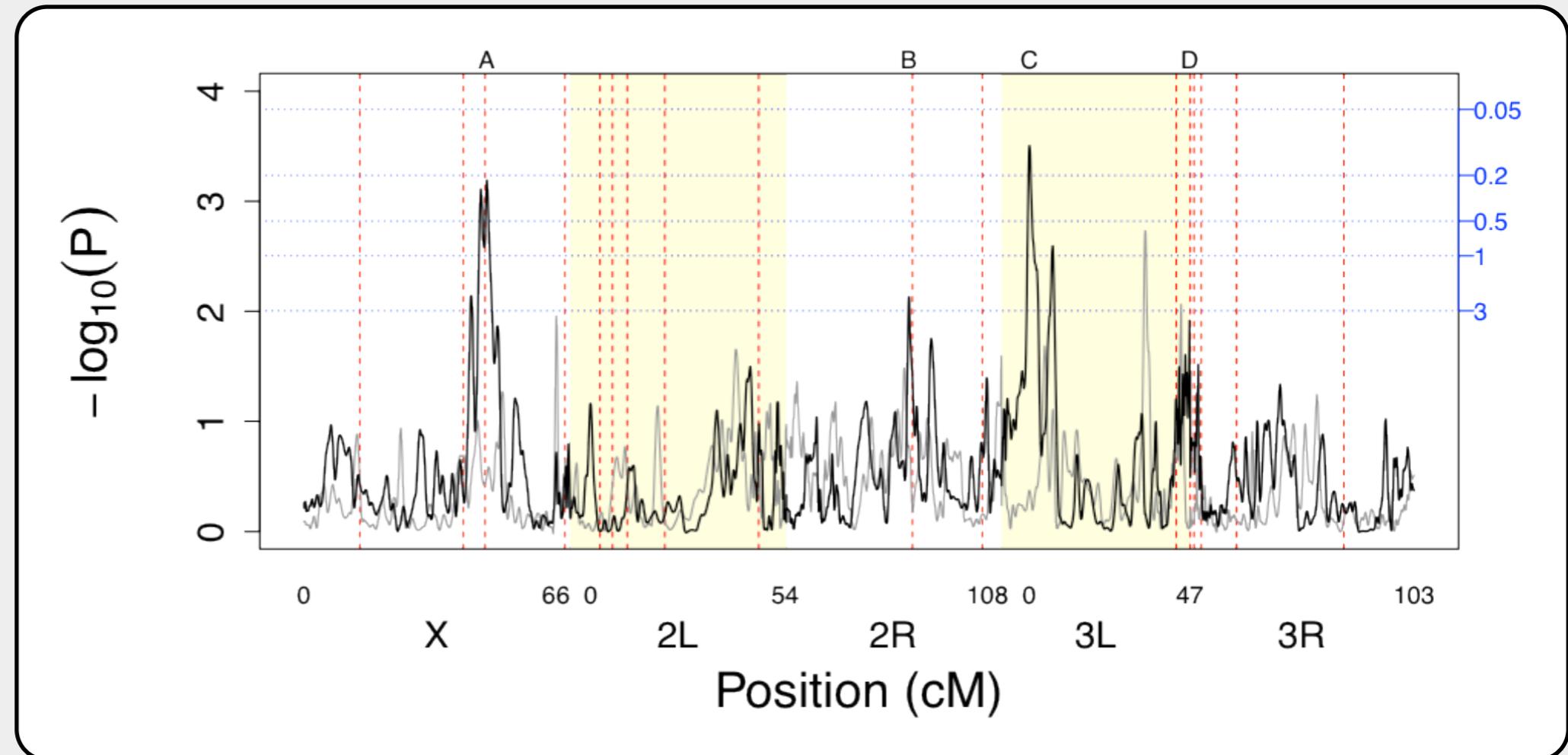


Behavioral / Neural Gene eQTL

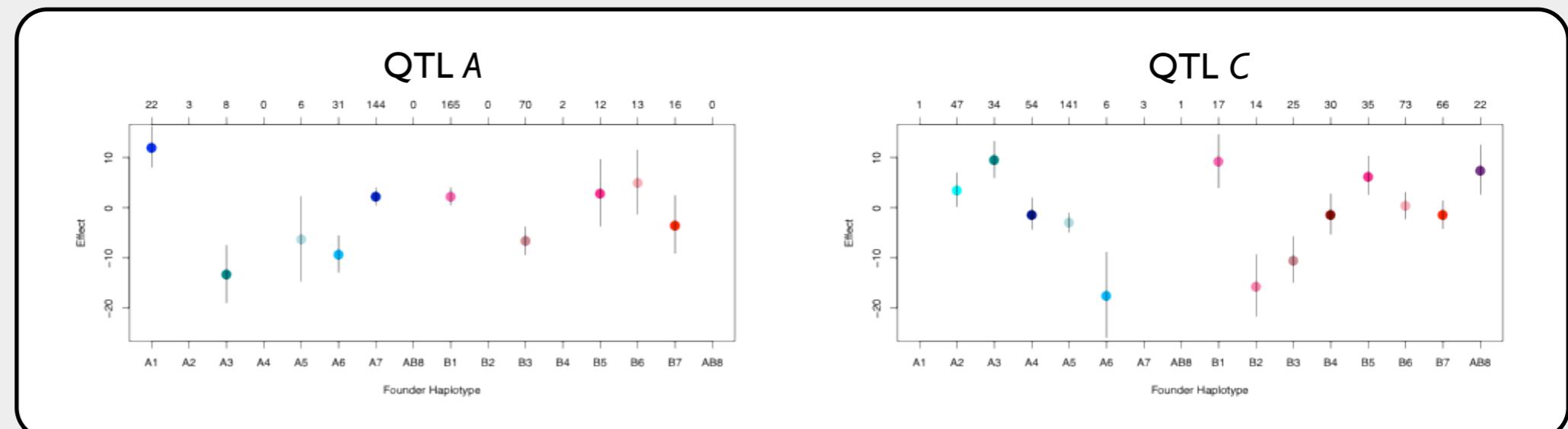
7,828 eQTL mapped for
7,422 target gene
expression measures

Multiallelic QTL ?

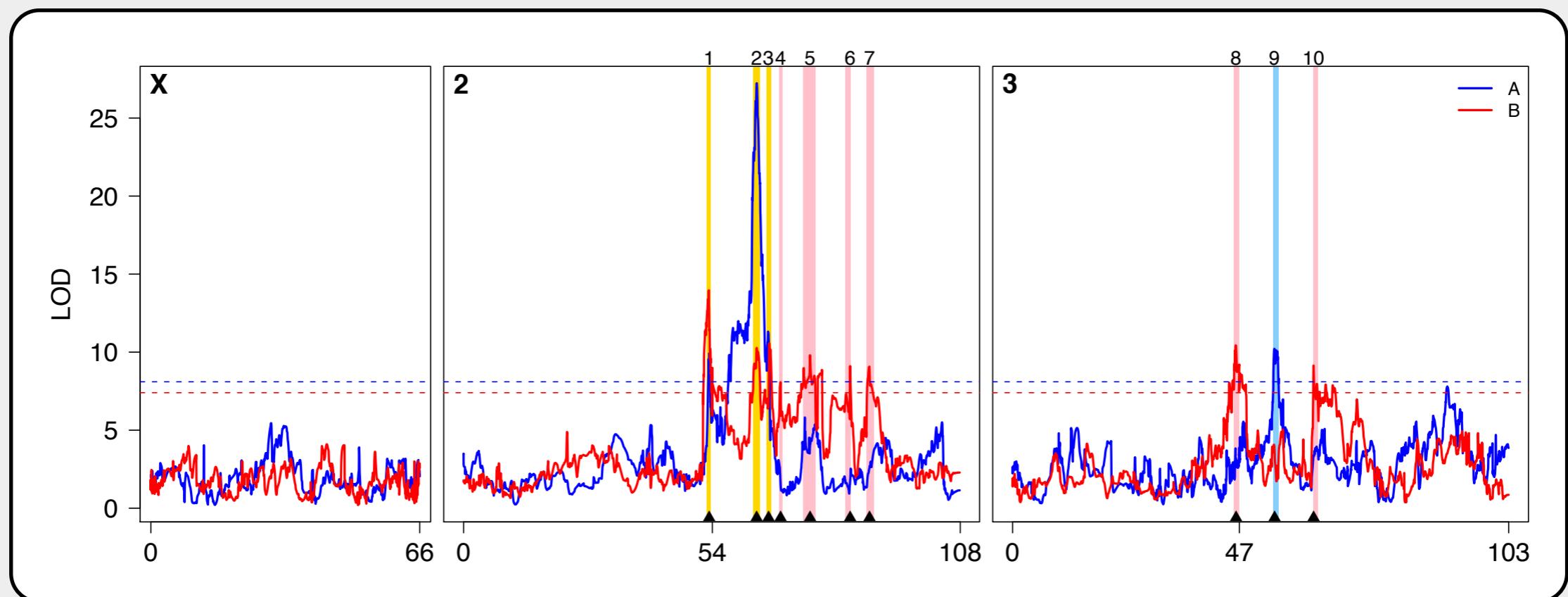
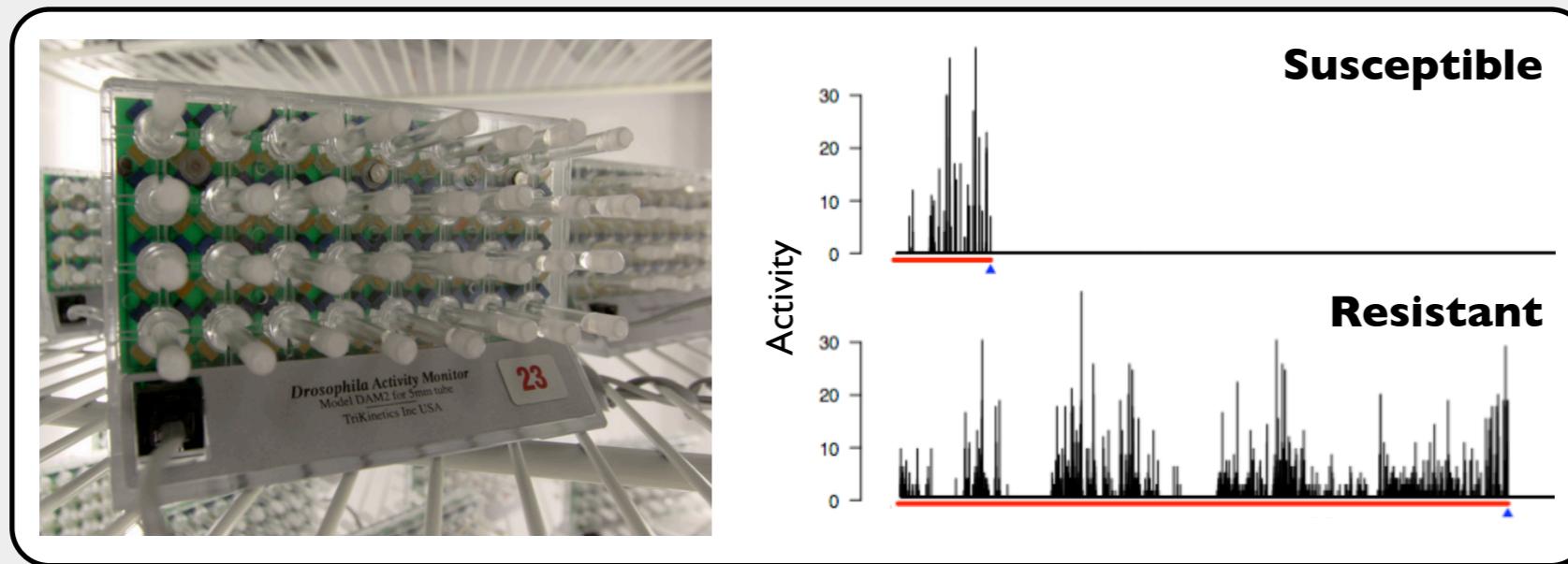
QTL for
methotrexate
toxicity



No clear
biallelic pattern
of strain effects



Caffeine Resistance QTL

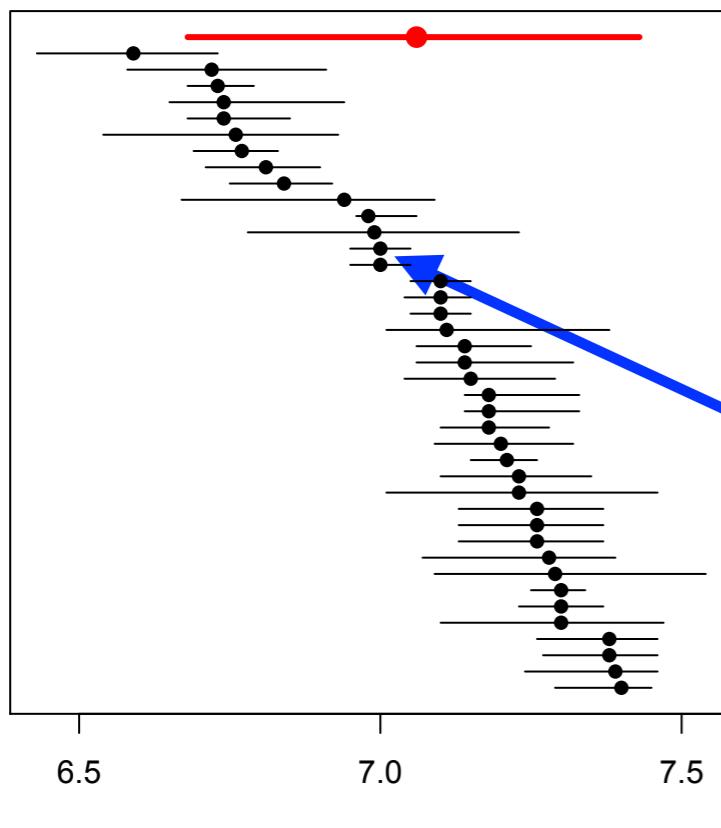


pQTL-eQTL Overlap

→ Combine pQTL and eQTL to help define loci contributing to phenotypic variation

Q2.2R.AB

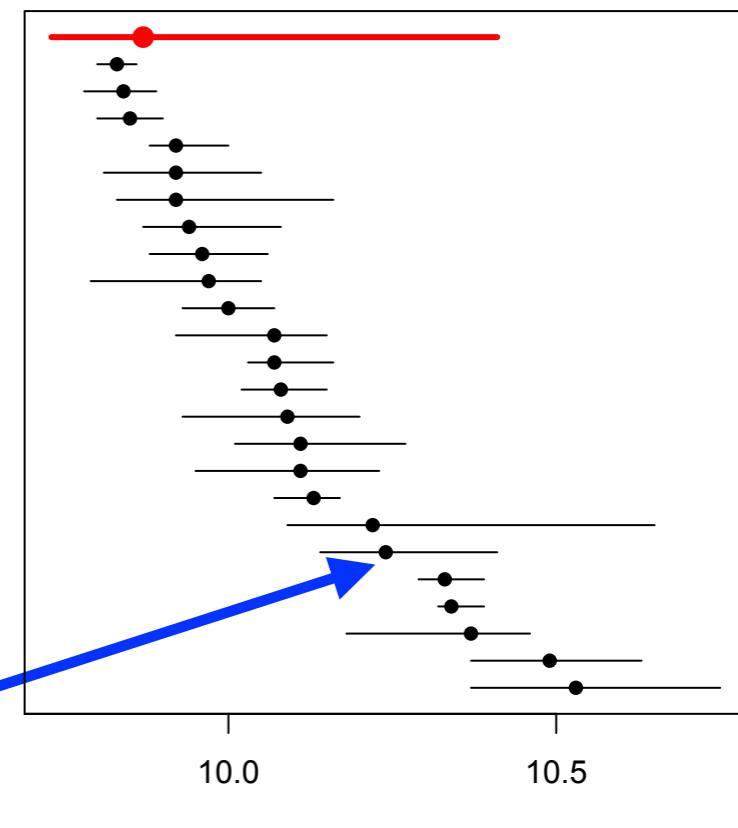
Implicates 128 genes
44 have *cis*-eQTL



Cyp12d1-d

Q9.3R.A

Implicates 81 genes
24 have *cis*-eQTL



Cyp313a1

Acknowledgements

Current Lab Members

Jenny Hackett
Chad Highfill
Sophia Loschky
Brittny Smith

Kristen Cloud*
Matthew Turner*
Kenna Whitley*

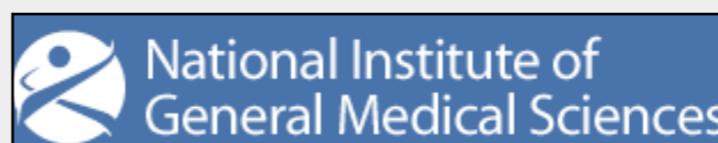
Former Lab Members

Steve Hoofer
Tara Marriage
Casey McNeil
Theresa Melhem
Chris Merkes
Michael Najarro
Brian Sanderson

Clint Bain*

Collaborators

Karl Broman (UW-M)
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Libby King (UCI)
Tony Long (UCI)
Erik Lundquist (KU)
Lynda Morrison (SLU)
Saunak Sen (UCSF)
Andrew Symons (KU Med)



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NIH P20-GM103418
(PI: Doug Wright)

NIH R01-GM085260

NIH R21-NS070417
(PI: Erik Lundquist)